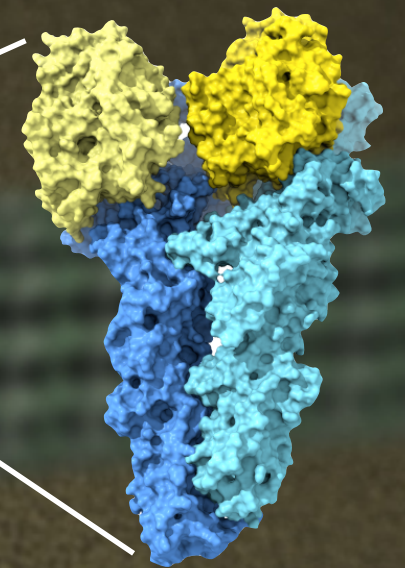
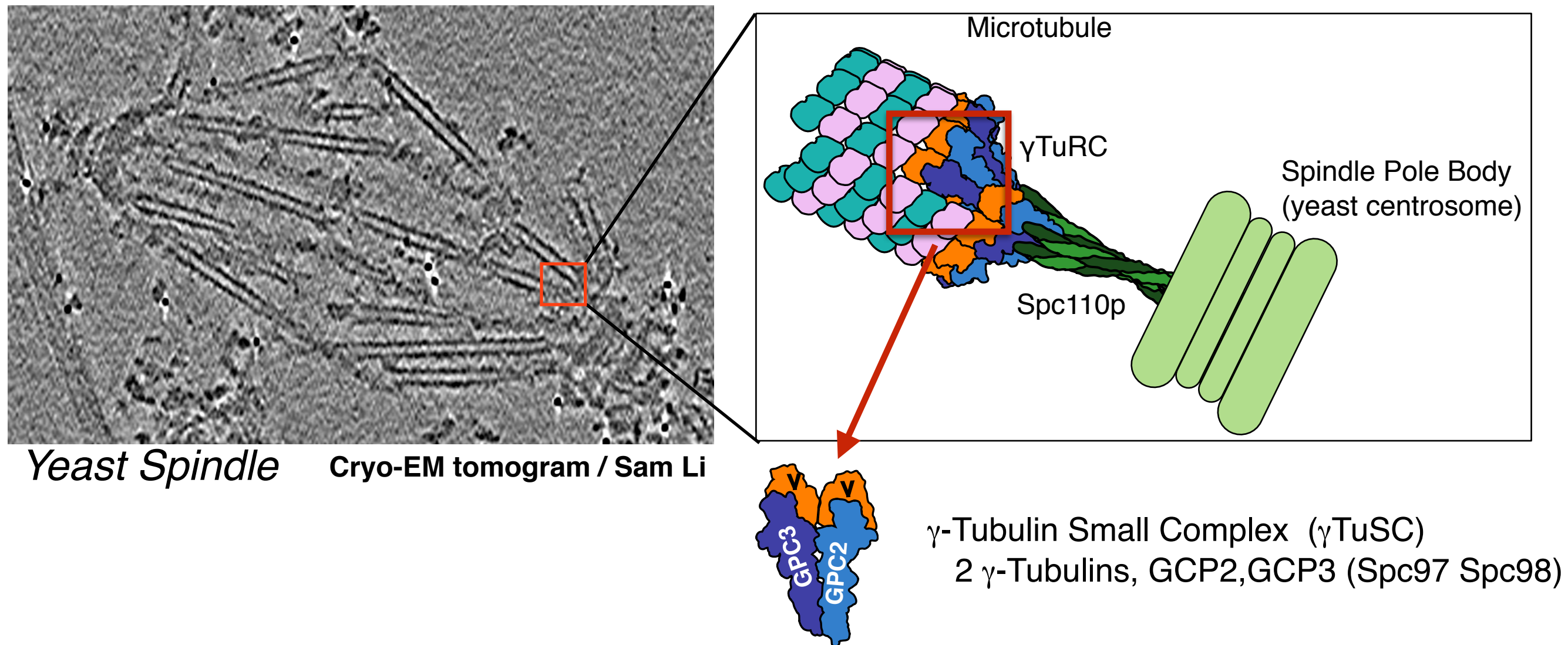


The Atomic Structure of the Microtubule Nucleating γ -tubulin Small Complex and its Implications for Regulation

*Axel Brilot
Agard Lab, UCSF
in collaboration with
Davis Lab, UW*

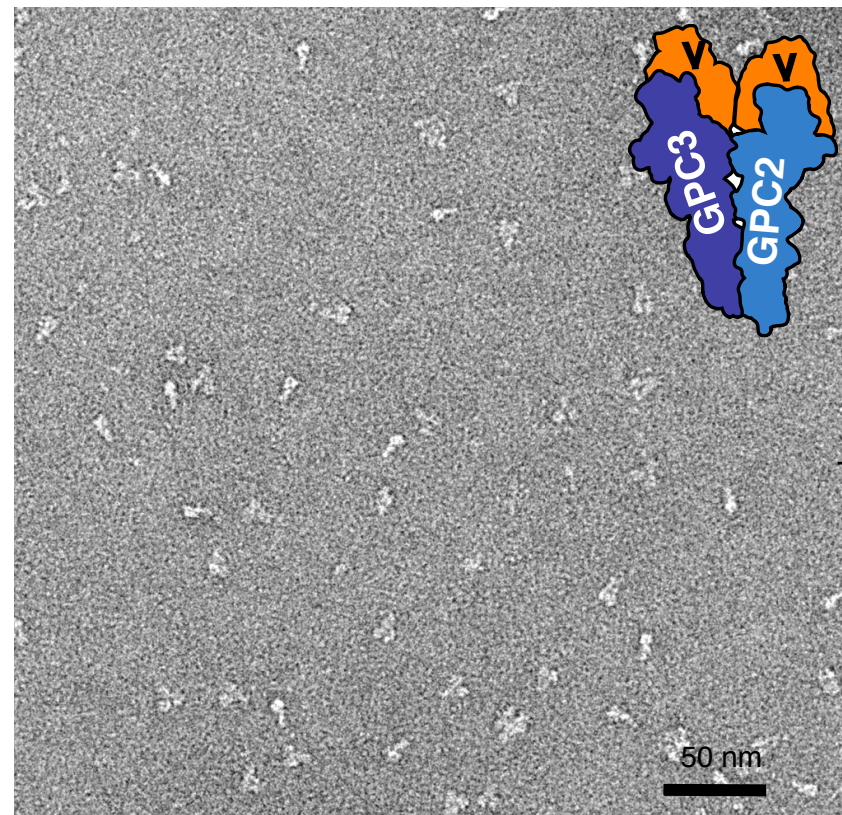
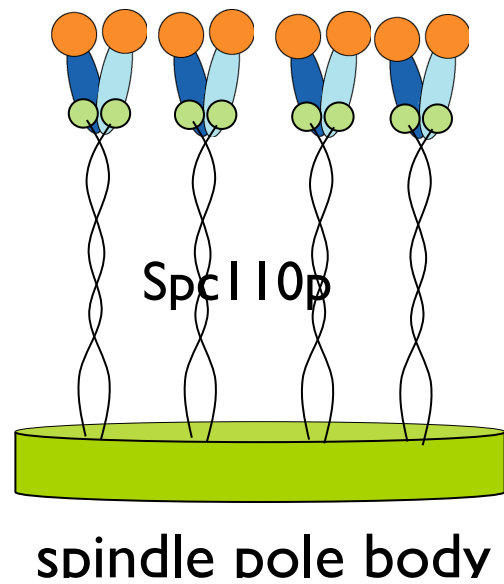


Microtubule nucleation by γ -tubulin Complexes



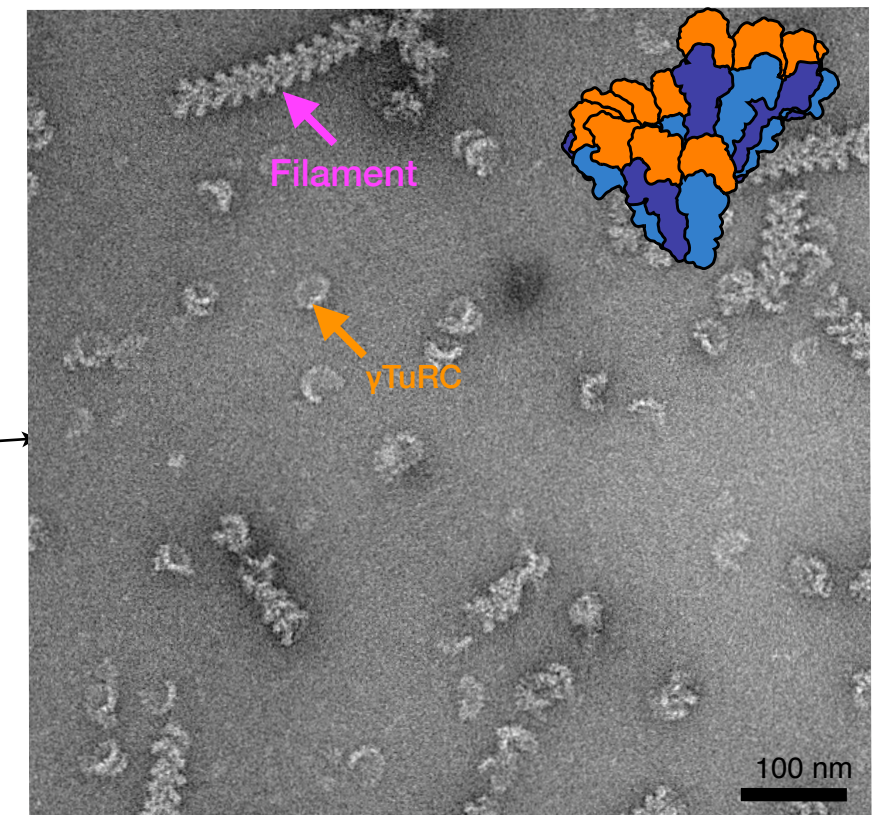
γ TuSC/ γ TuRC structure, assembly, activation

Attachment factor *Spc110* stabilizes γ TuSC assembly

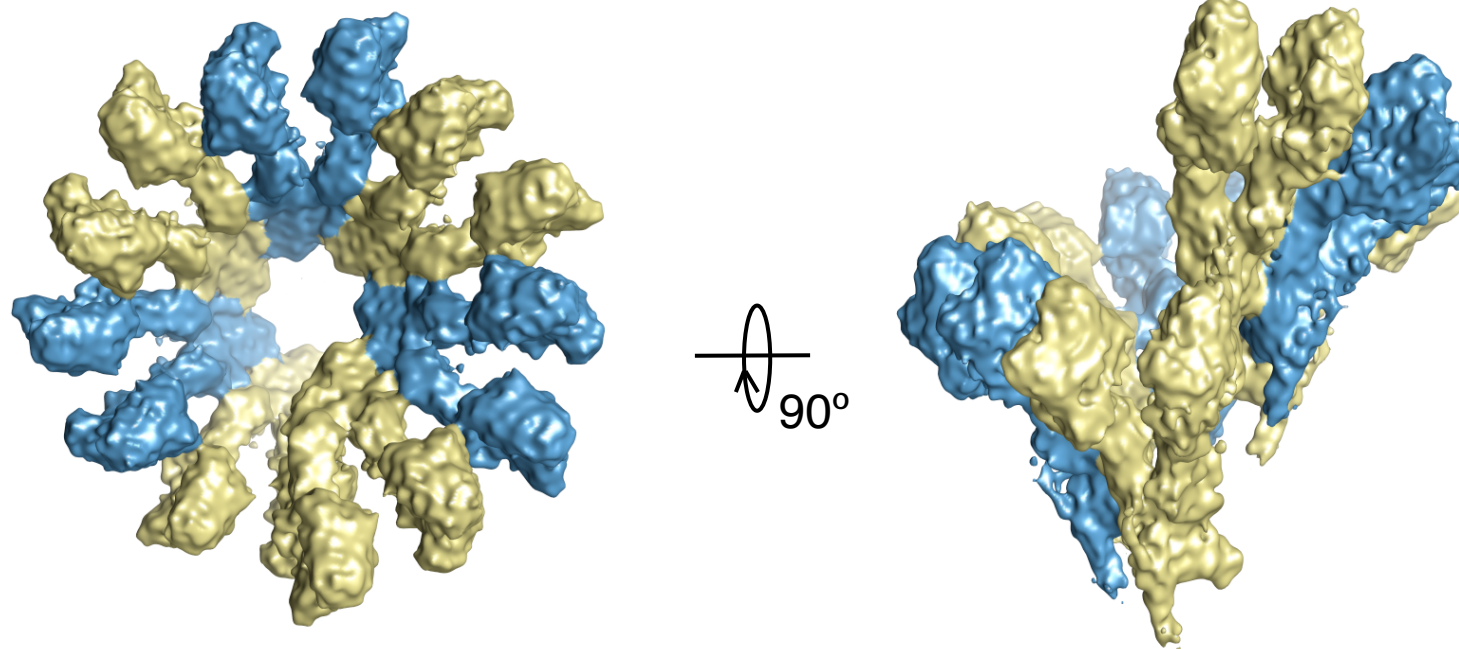


Individual γ TuSCs

+ Spc110p¹⁻²²⁰

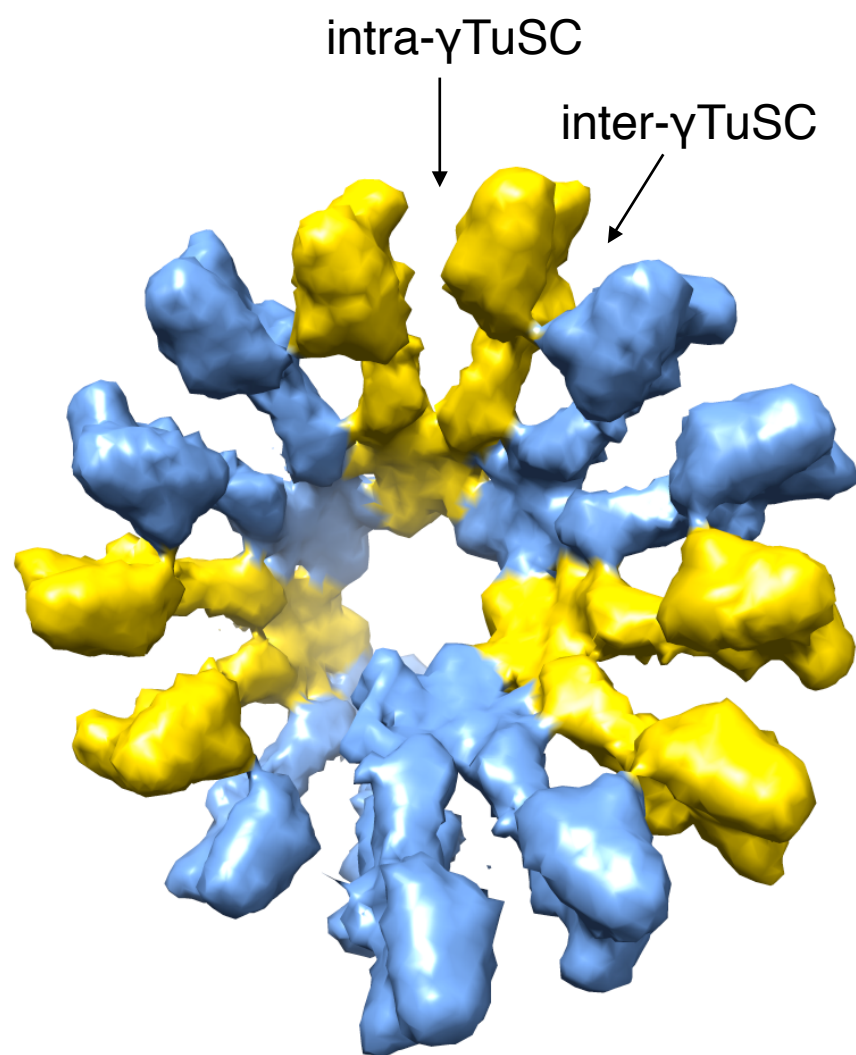


γ TuRCs/filaments

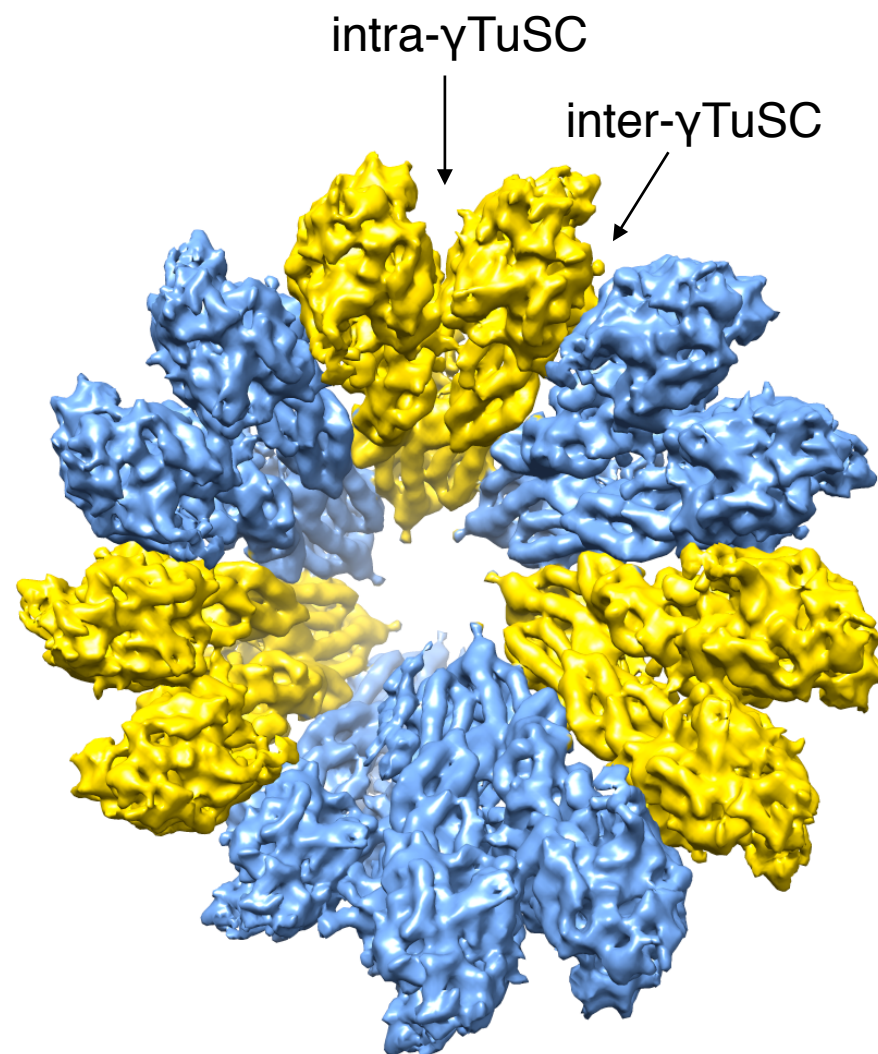


6.5 γ TuSCs/turn = 13 γ -tubulins = *in vivo* MT protofilament #

Open - closed transition enhances γ -TuRC MT nucleation



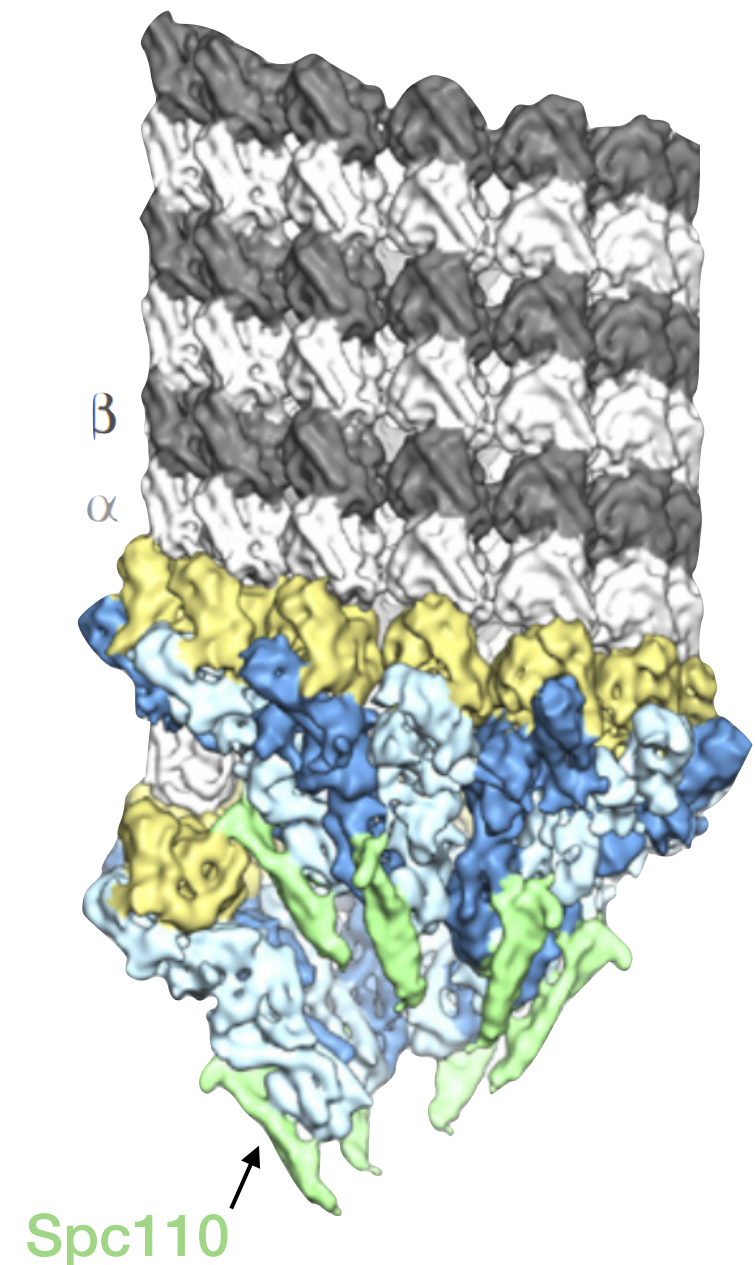
Open state



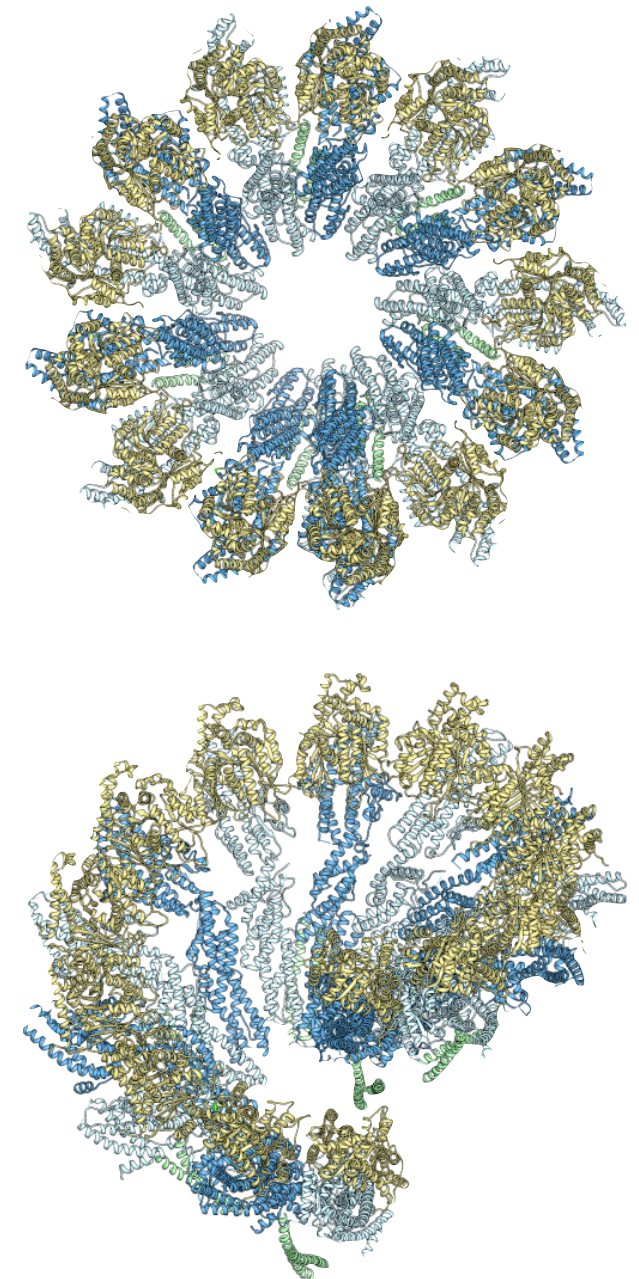
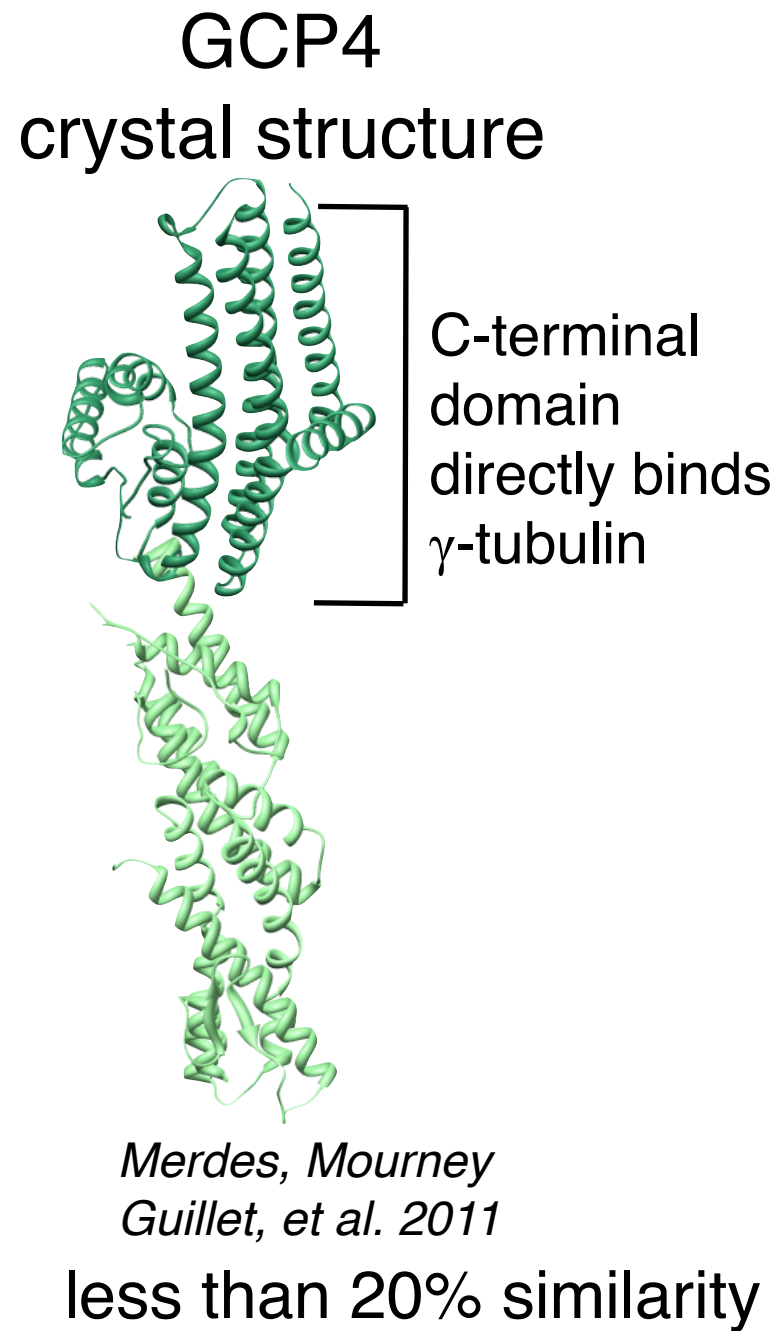
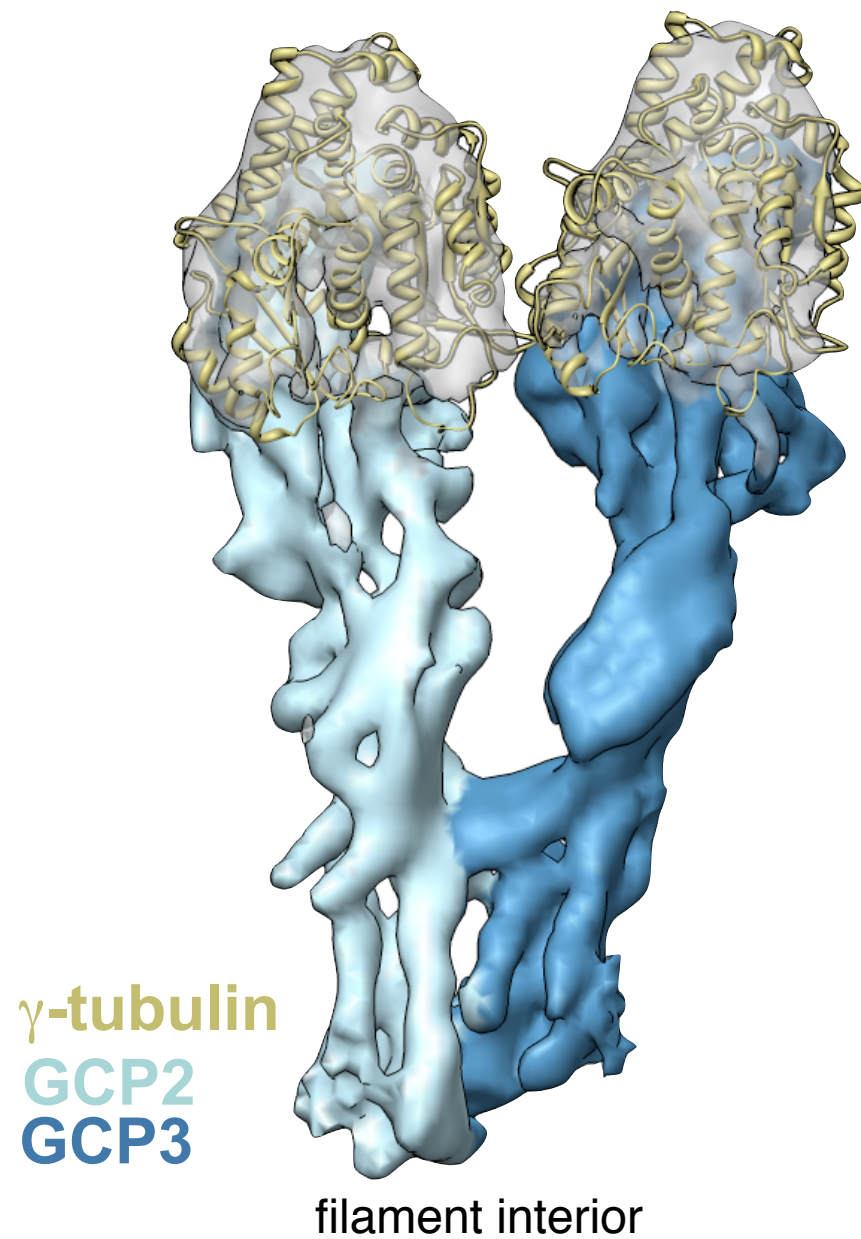
Closed state
(disulfide stabilized)

- *closed state better MT nucleator*
- *suggests closure as a regulatory mechanism*

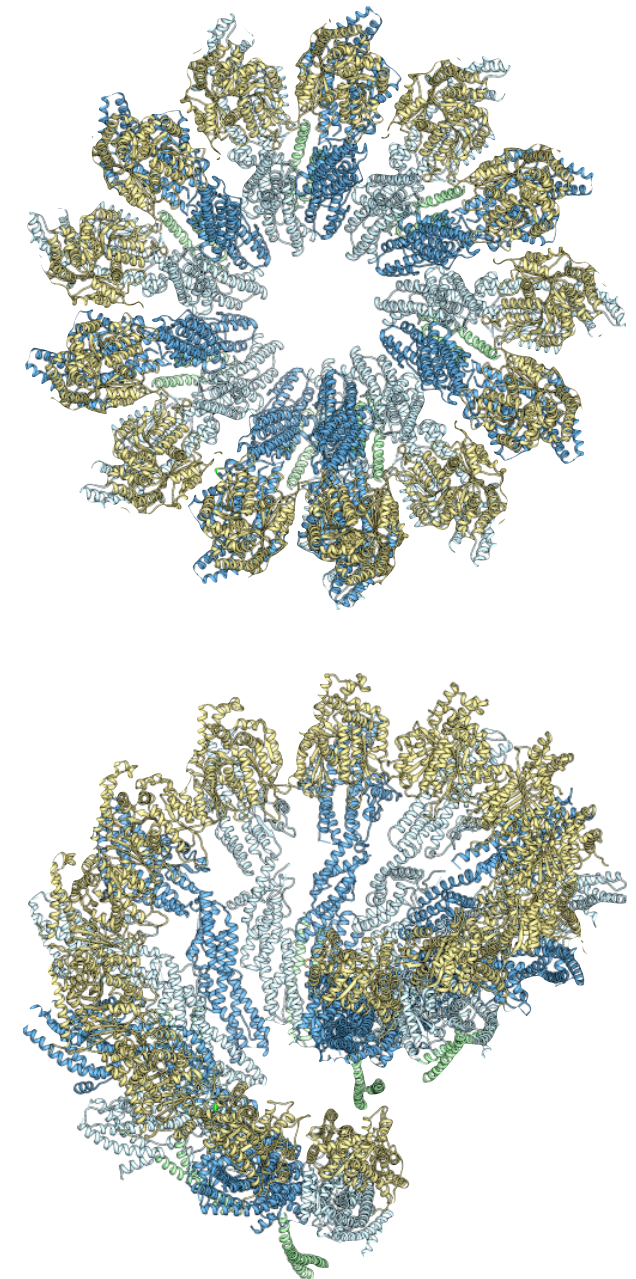
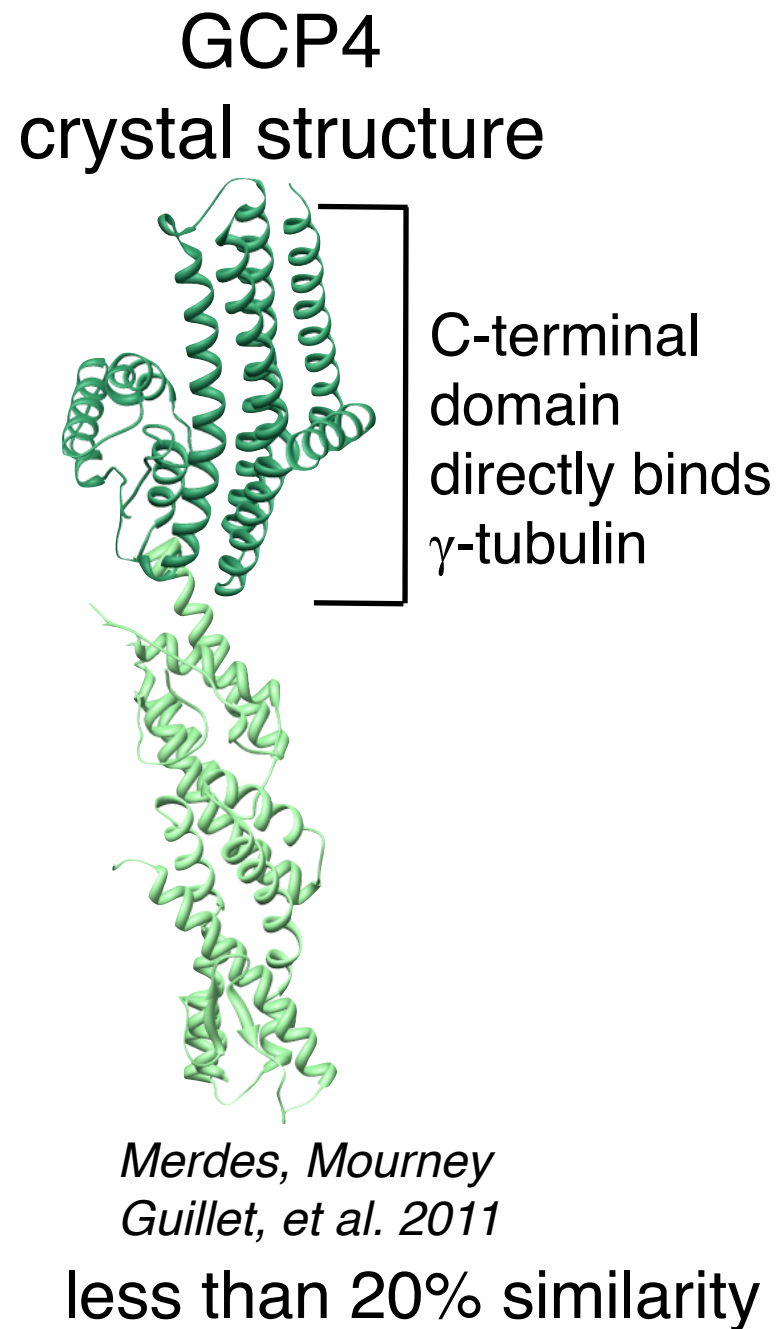
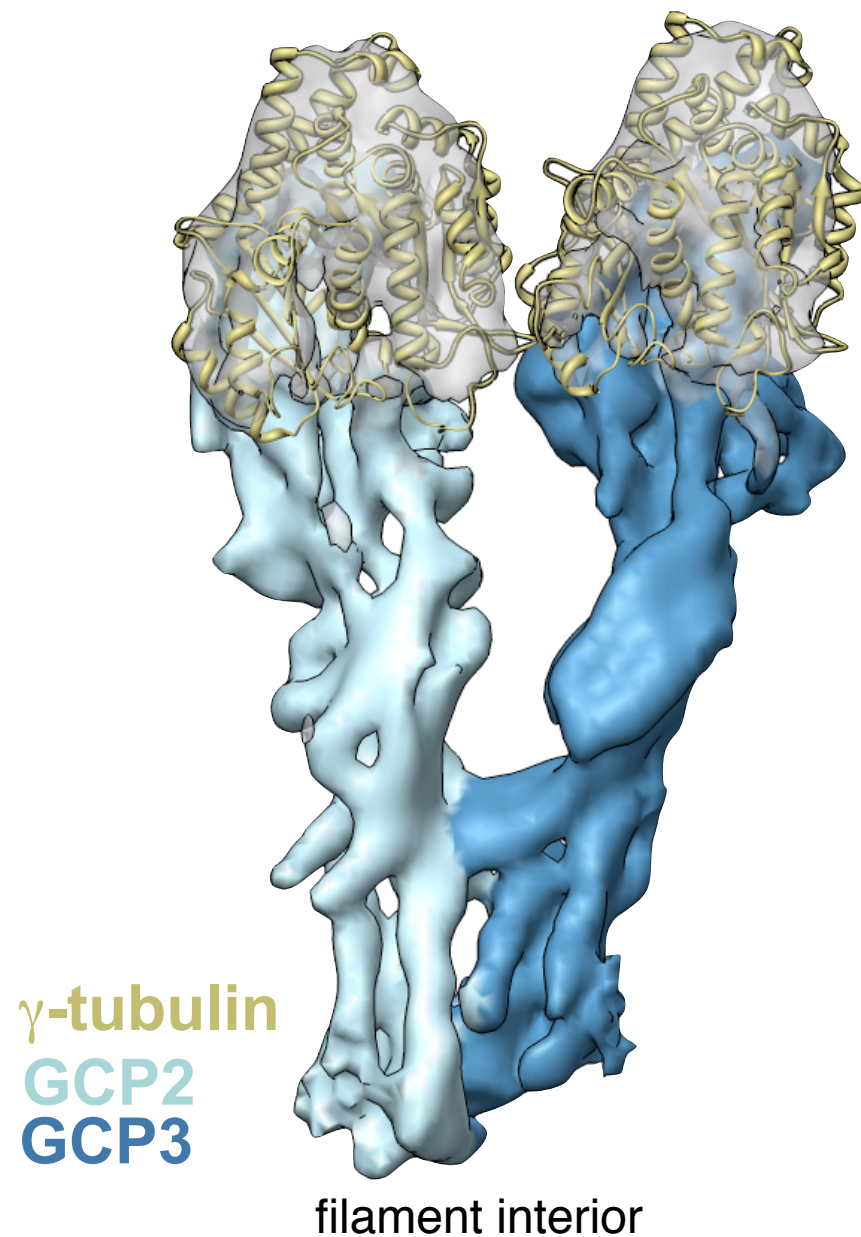
Model:
 γ TuRC nucleating microtubule



γ TuSC pseudo-atomic model built using 6.5Å oxidized map

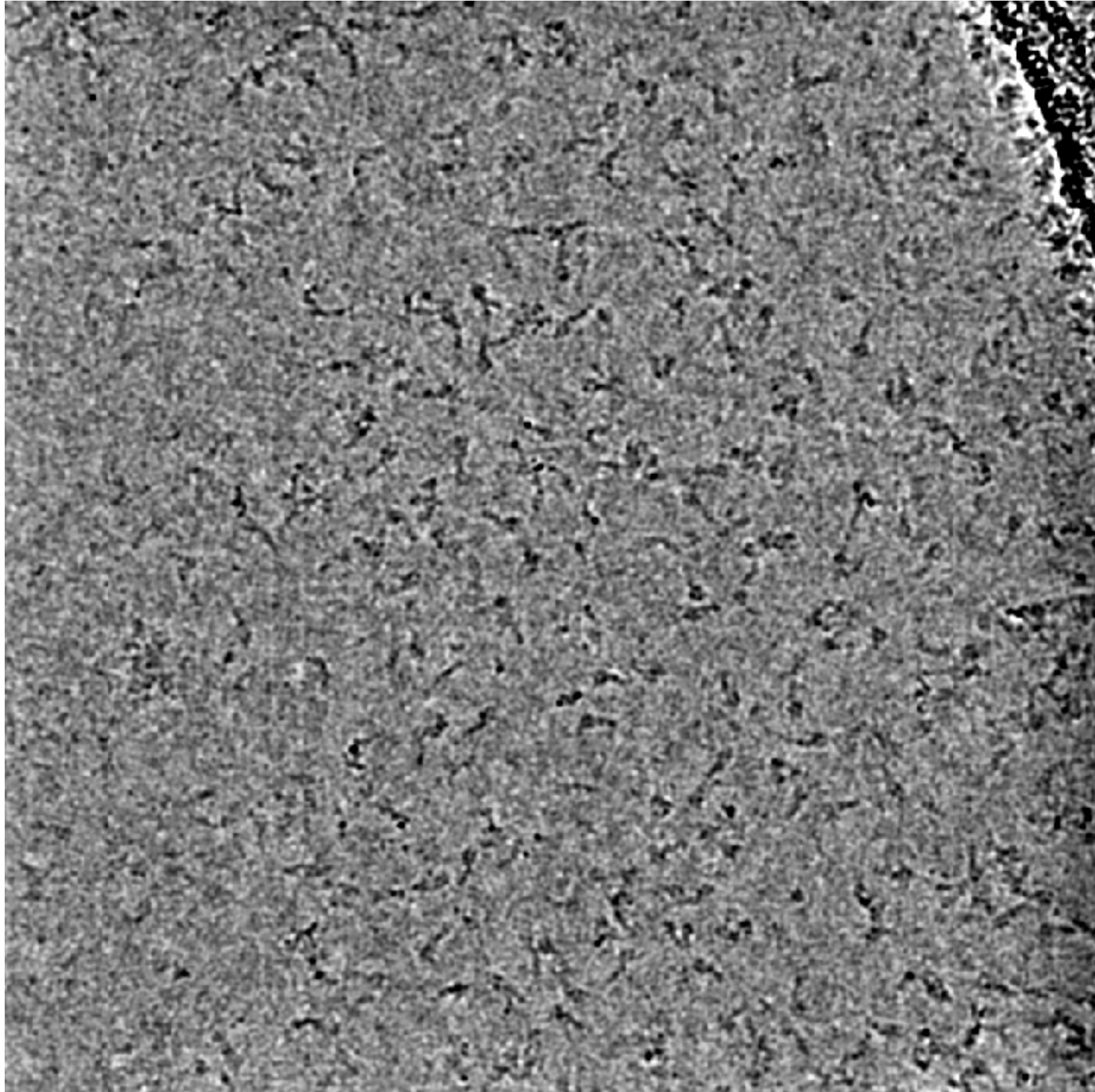


γ TuSC pseudo-atomic model built using 6.5Å oxidized map



*Missing 234 aa from gcp2, 275 aa from gcp3
Built into a ~6.5 Å map*

The Image Data



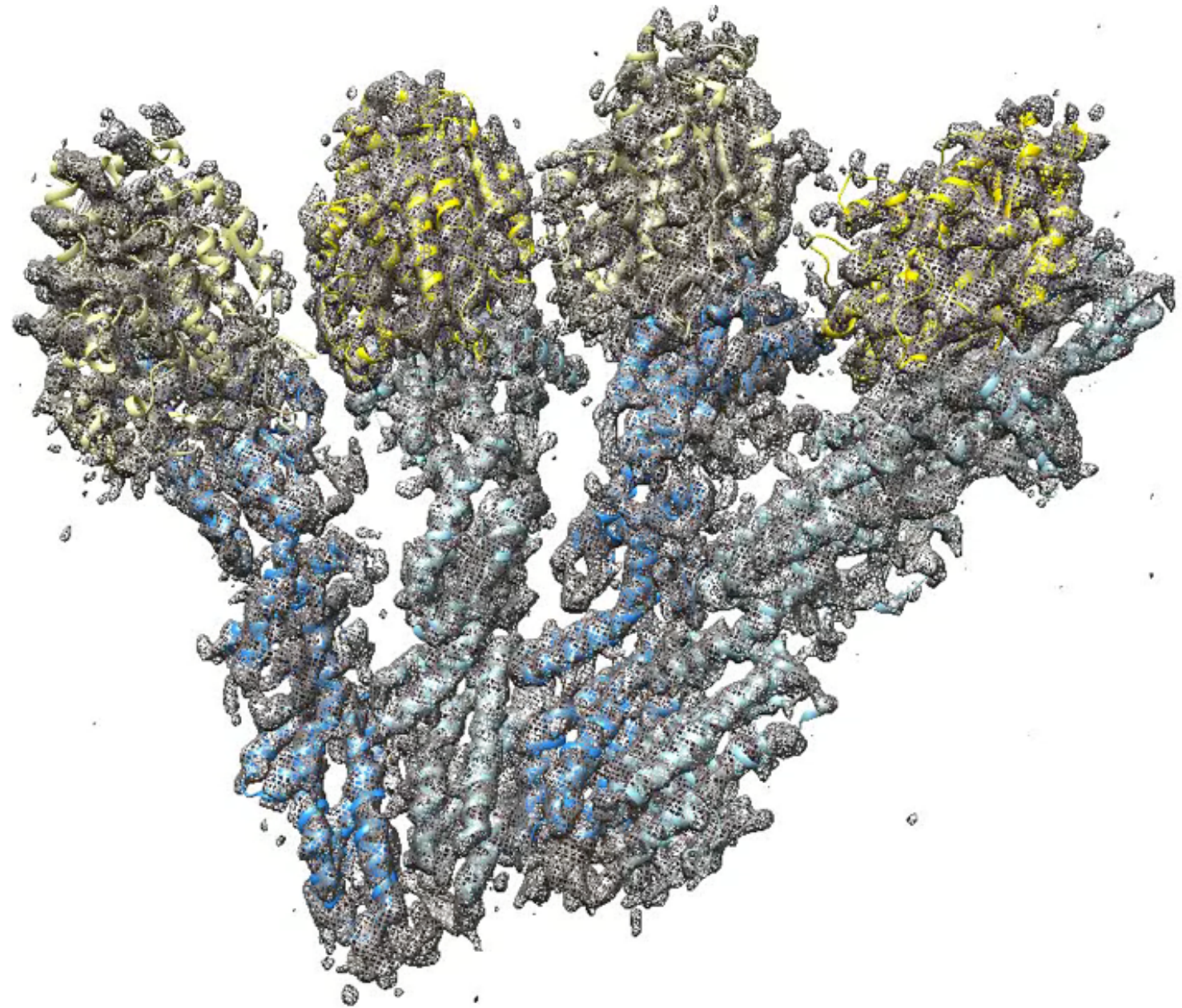
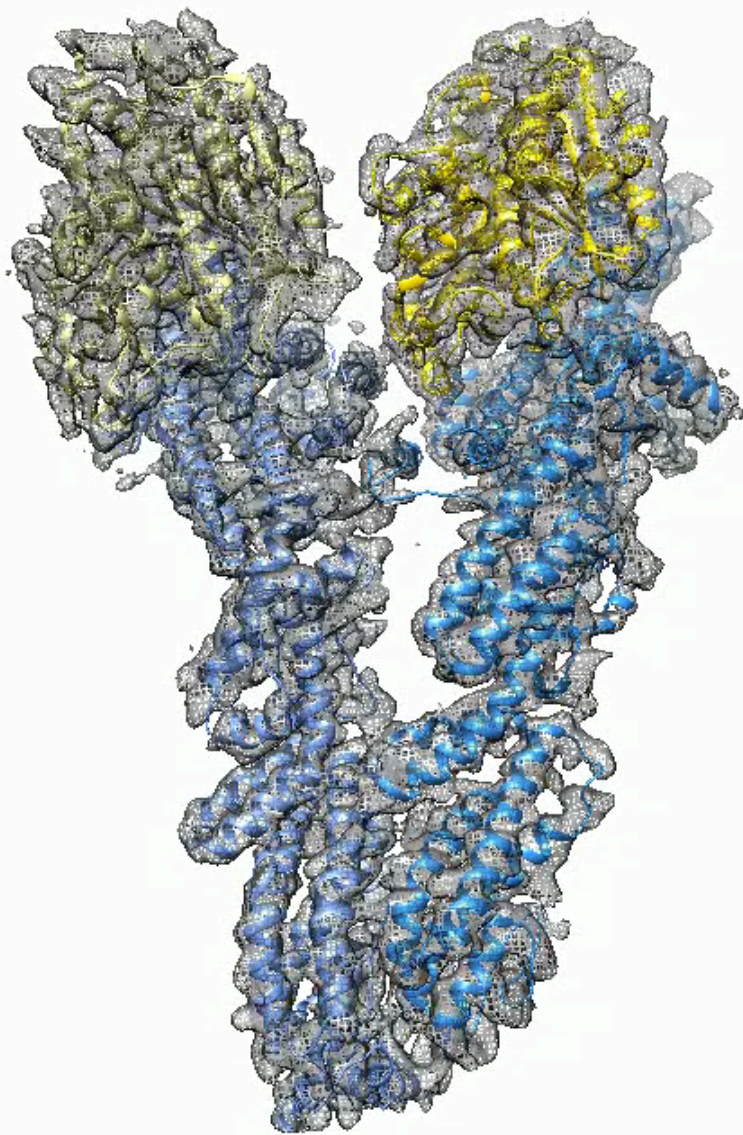
Polara Data

~80 e-/Å²

Dose filtered &
aligned with MotionCorr2

Thon rings 5Å
or better

γ TuSC monomer/dimer by single particle cryoEM (3.8Å)



- first true atomic description of γ TuSC, numerous inserts, etc
- differences in the interfaces between the γ TuSCs vs internal interface
- conformational changes in γ -tubulin upon assembly into γ TuSC
- interpretation of phosphorylation sites, mutations

Workflow

Drift correct & pick



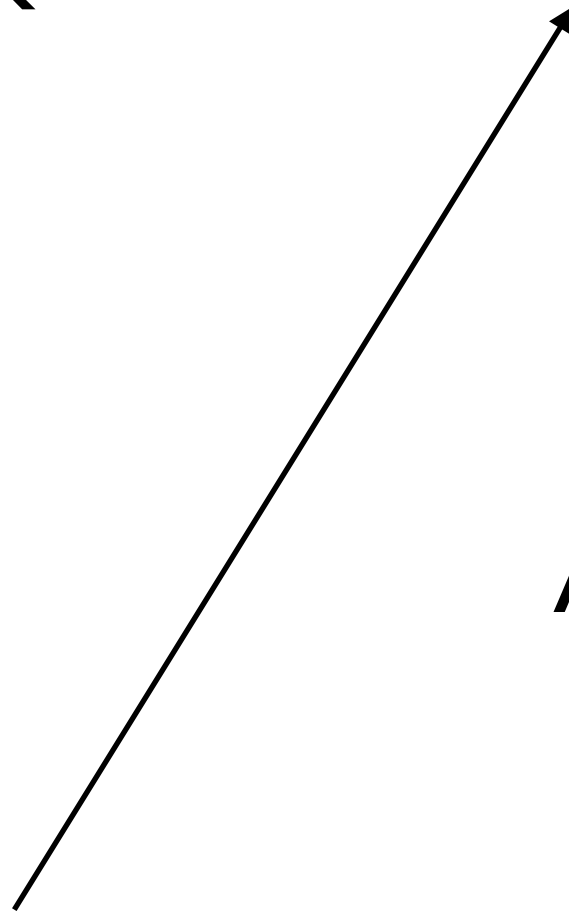
Determine CTF



extract particles



2D Classification



3D Classification



Extract classes



Align into one class



3D Classification

Improving the Map

Increase Dataset size (+0.5M particles)

Various Programs (Relion, cryosparc)

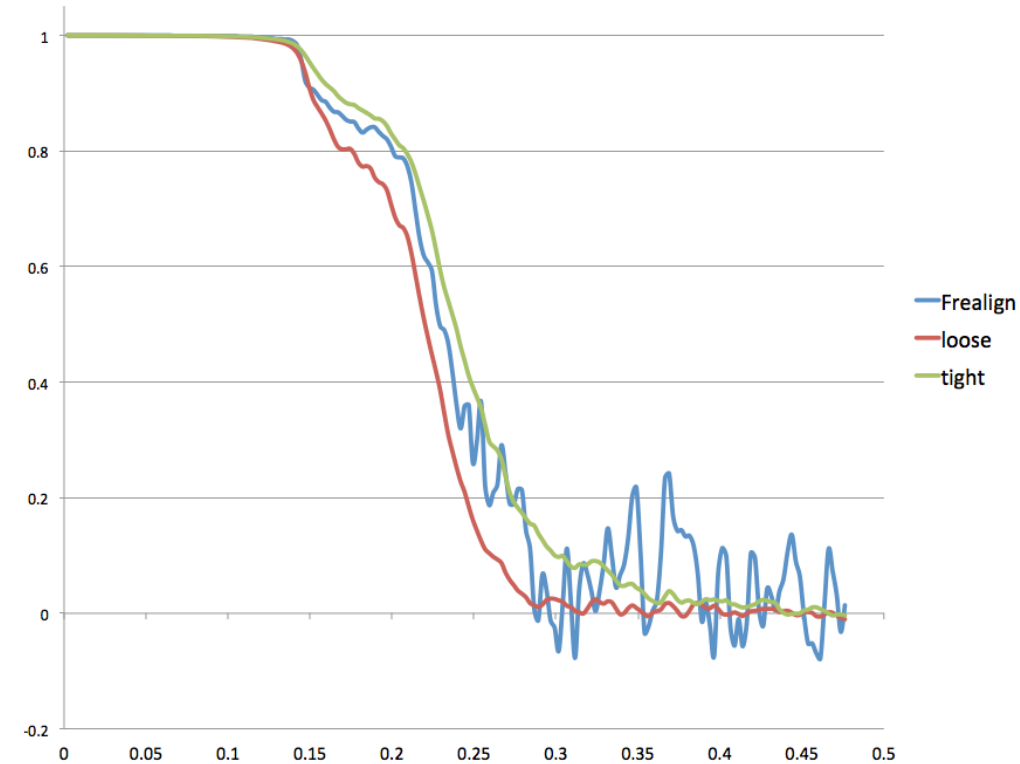
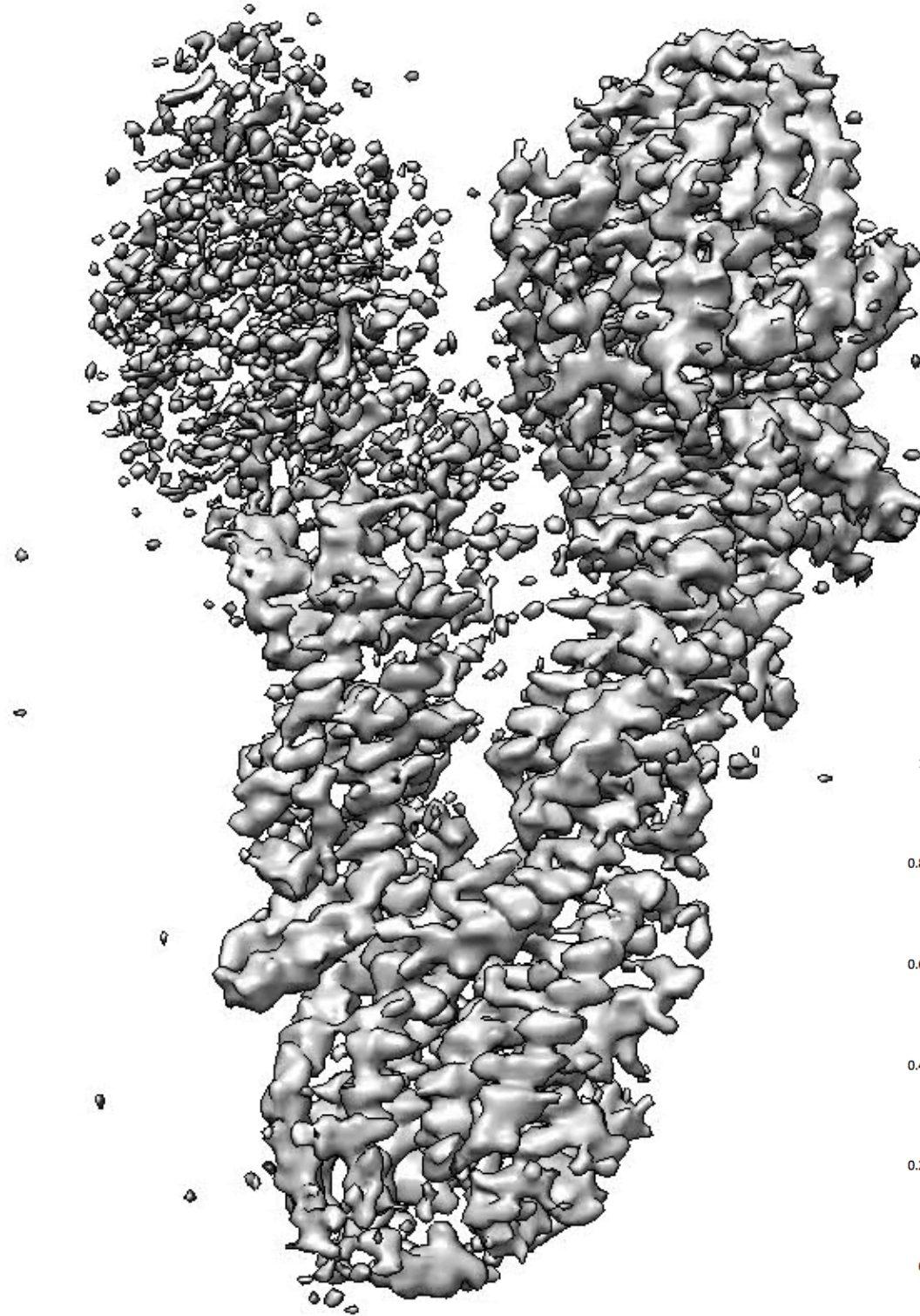
Full workflow, as well as feeding them classification results from Frealign

Focused Classification in Frealign

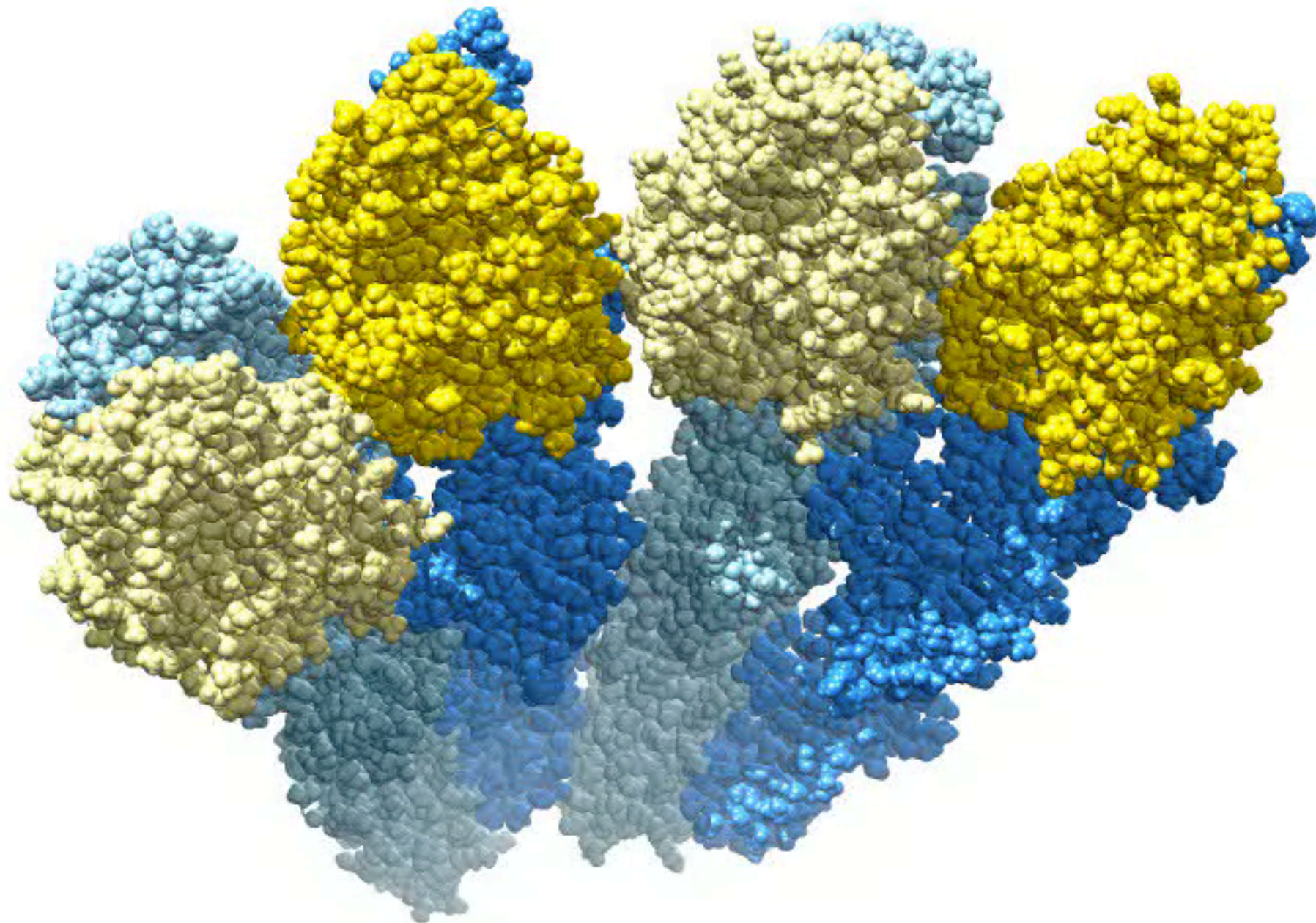
Various Masks

Half-Tusc, Base only, Base plus one tubulin arm

Improving the Map - Frealign, Shaped Masks and Weighting

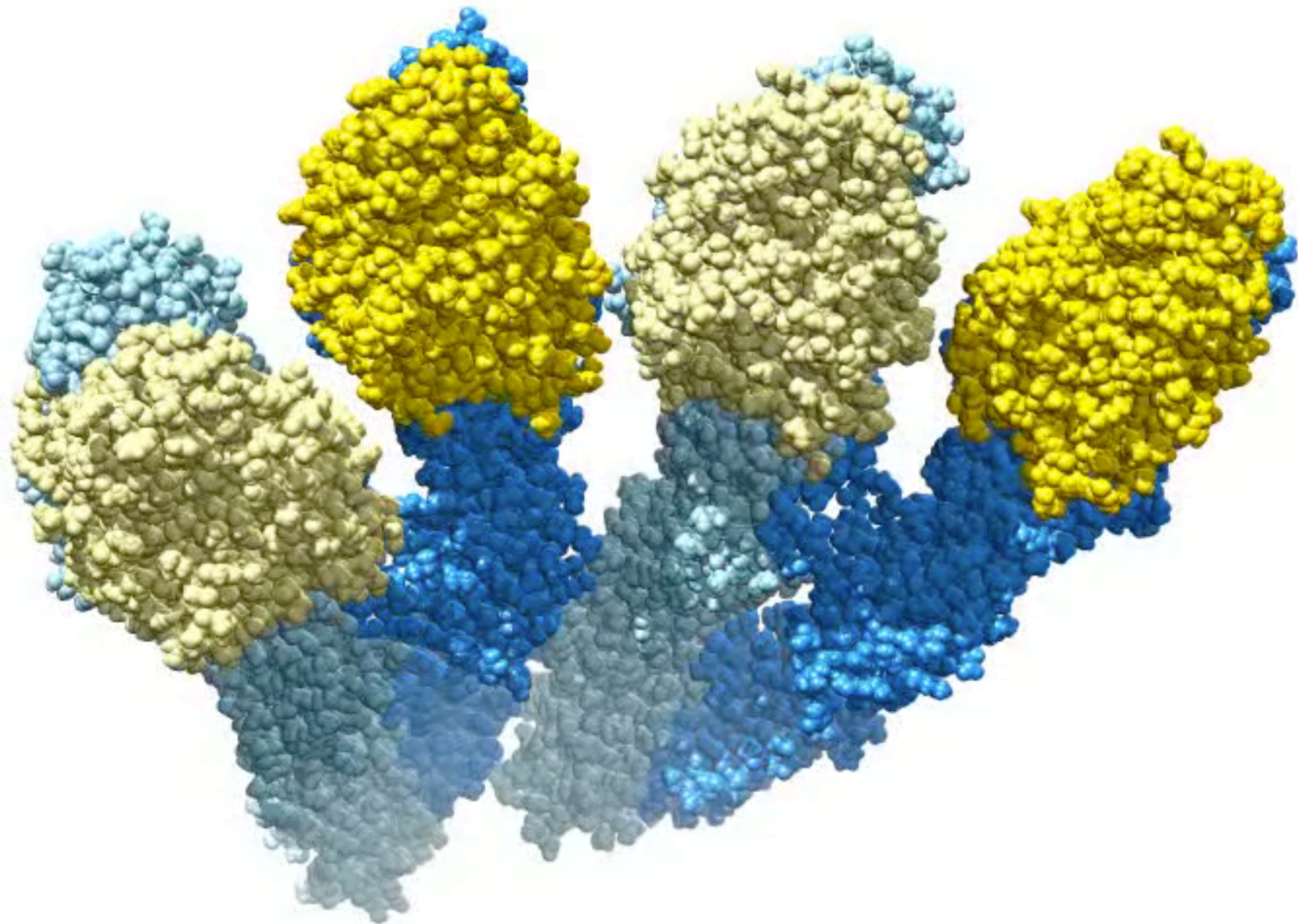


Assembly driven global conformational changes



both assembly & allosteric conformational changes required

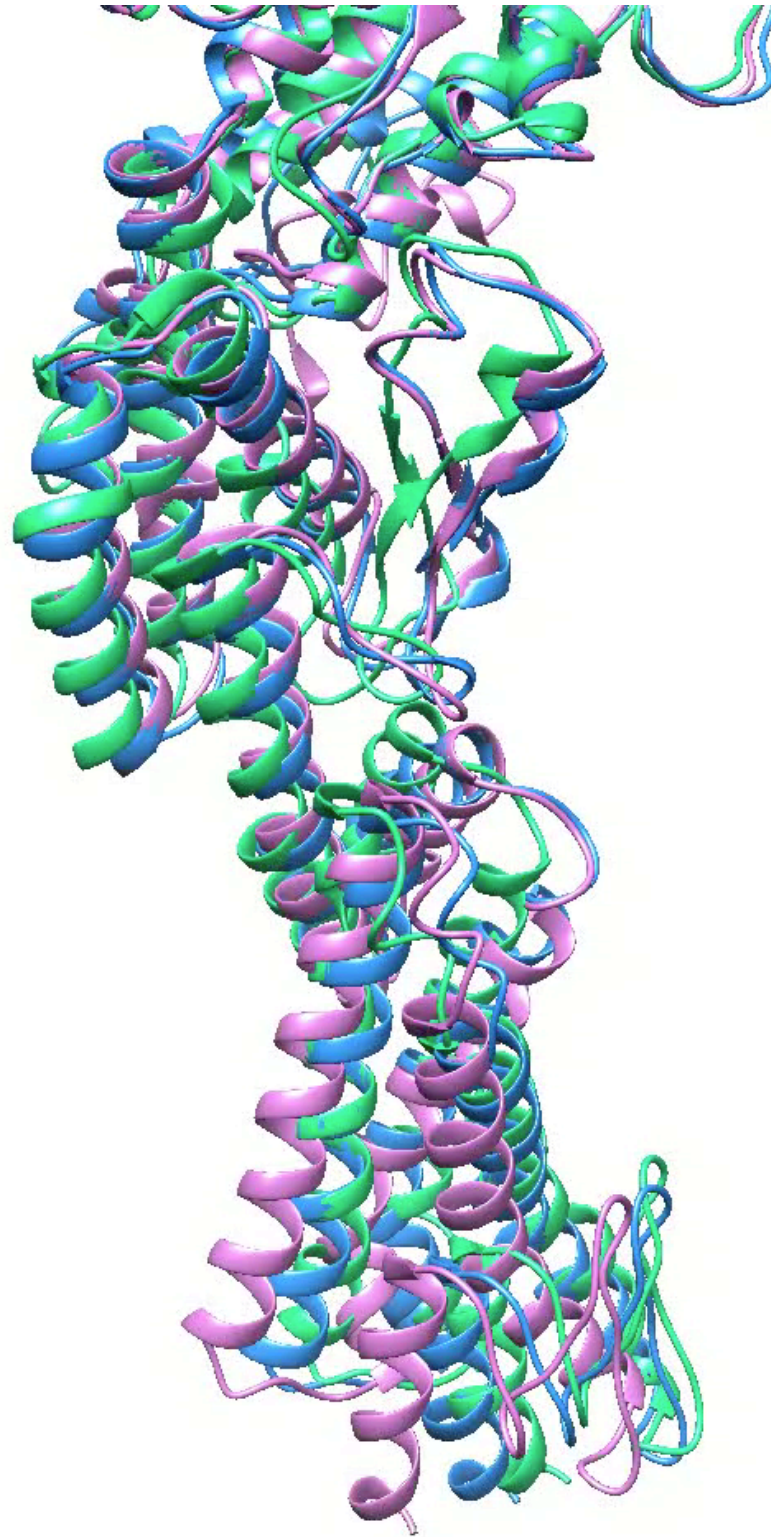
Assembly driven global conformational changes



both assembly & allosteric conformational changes required

Assembly driven global conformation changes

Monomer
Closed
Open

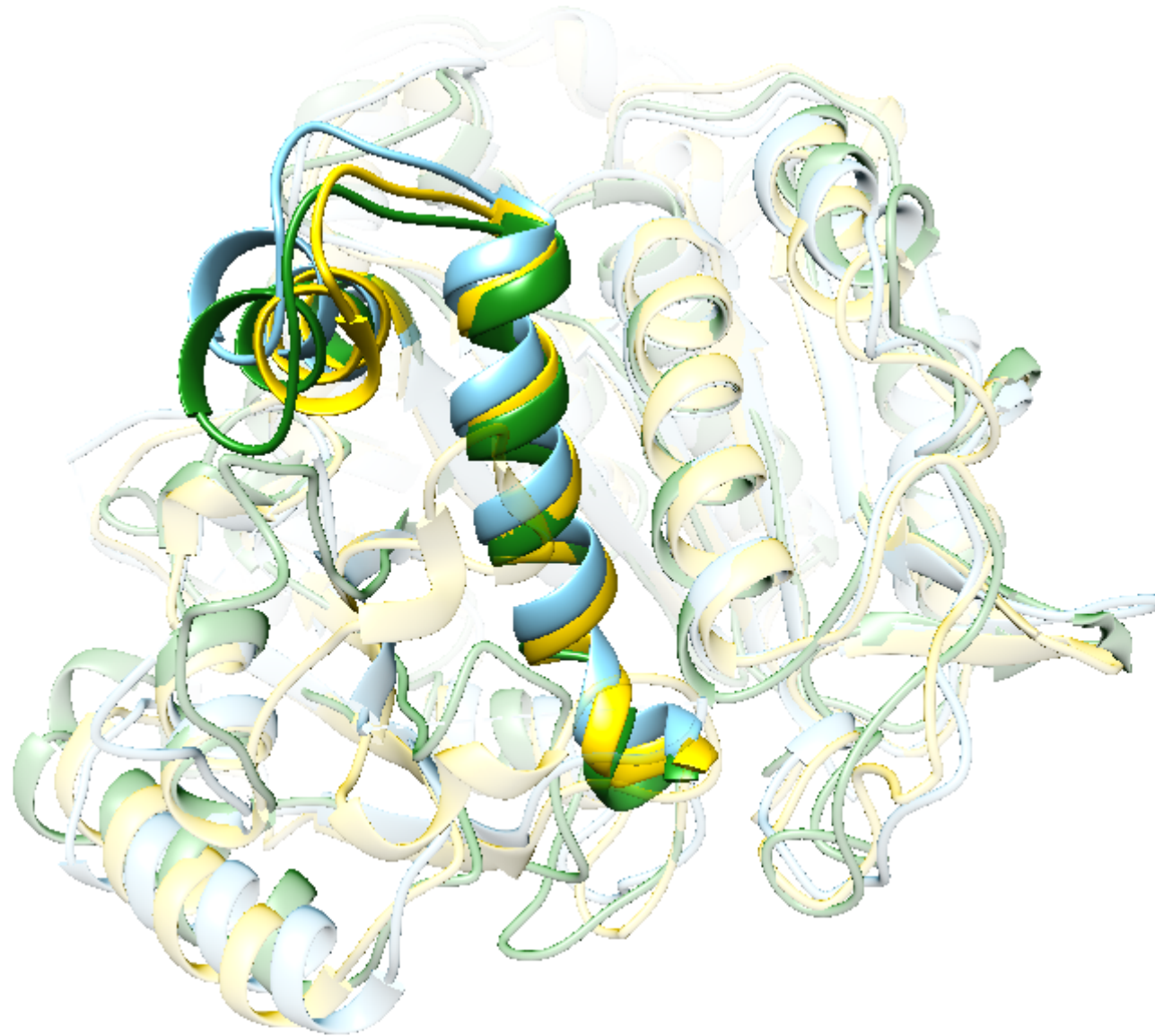


GCP3 N-terminus

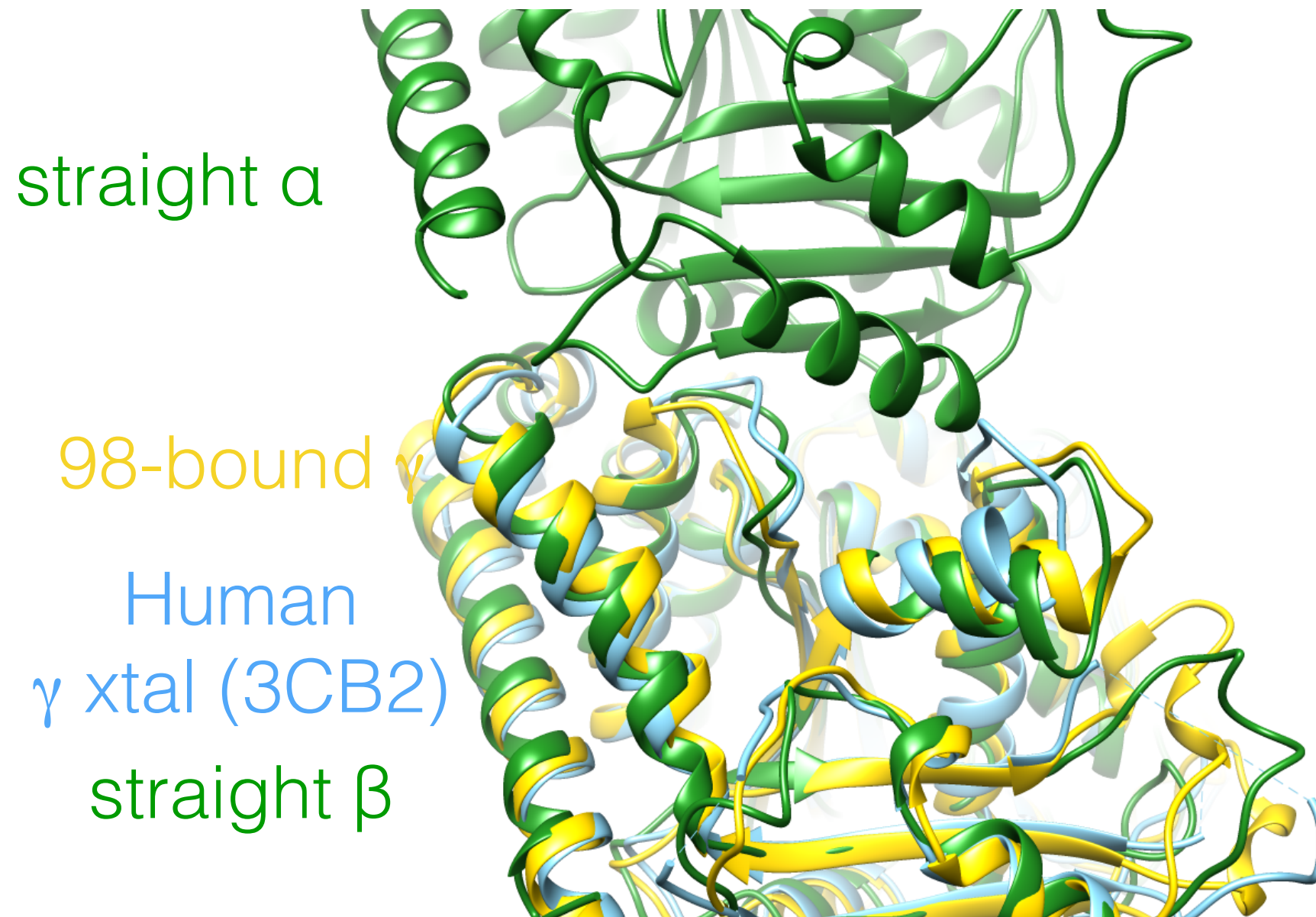
Twist of the conserved GCP domains is the major re-arrangement

What is the conformation of γ -tubulin on the γ -TuSC

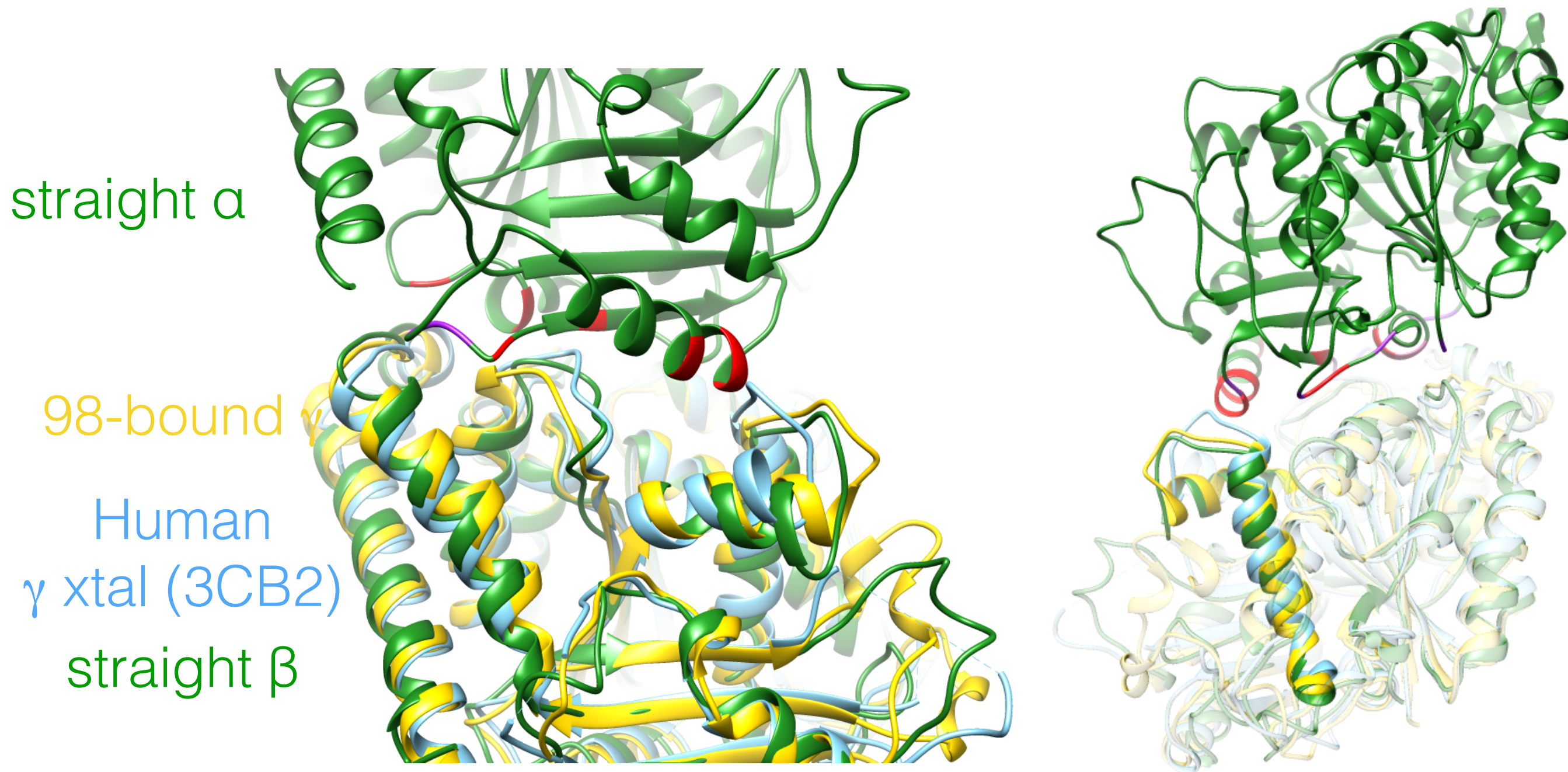
98-bound γ
Human
 γ xtal (3CB2)
straight β



What is the conformation of γ -tubulin on the γ -TuSC



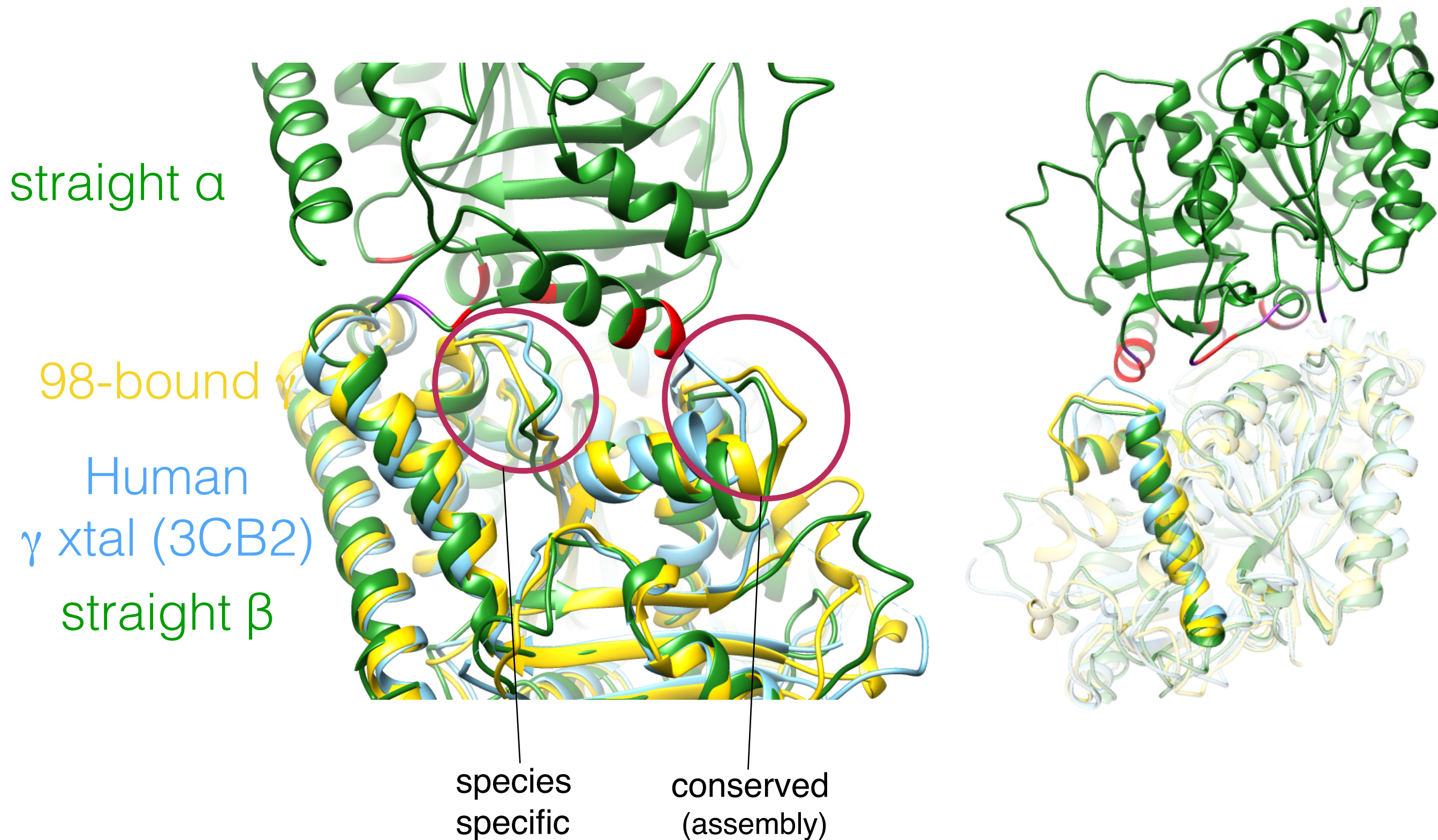
What is the conformation of γ -tubulin on the γ -TuSC



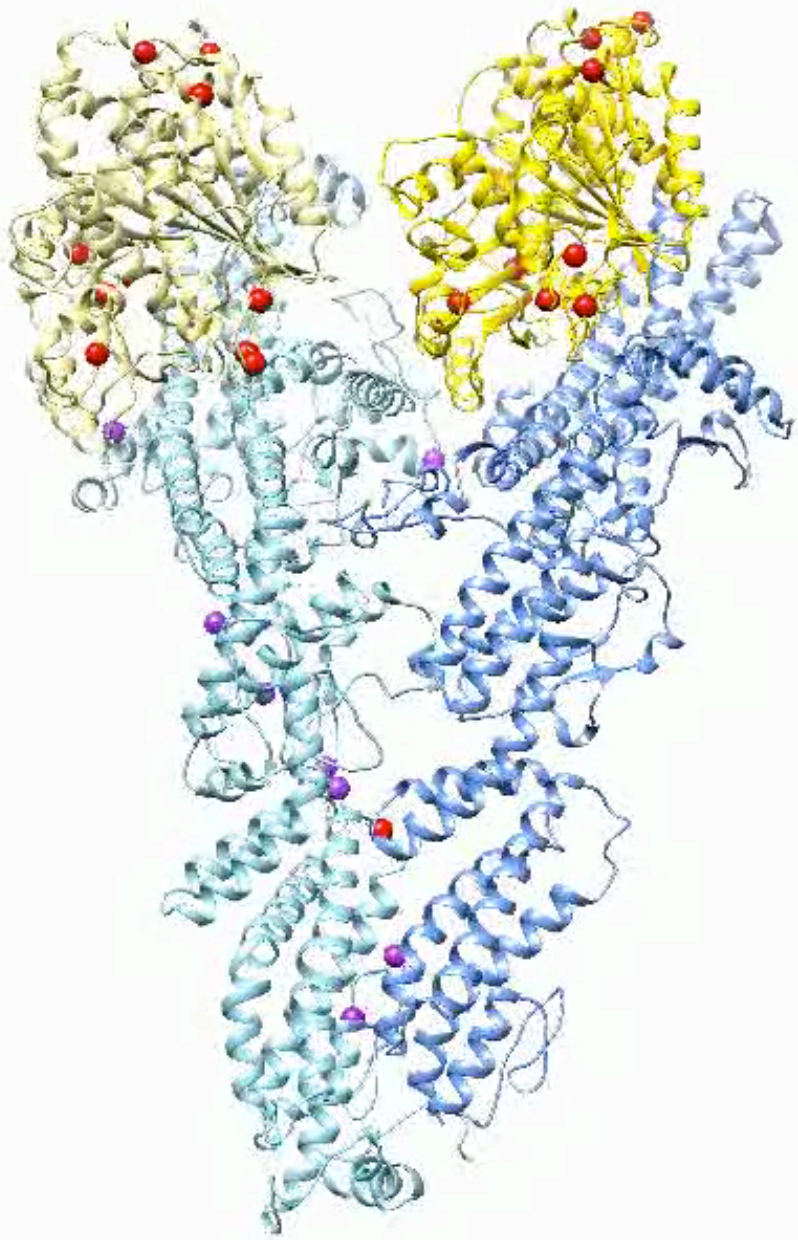
clashes w/human γ -tubulin

clashes with human and yeast γ -tubulin

What is the conformation of γ -tubulin on the γ -TuSC



GCP2 phospho sites suggests functional roles



GCP2

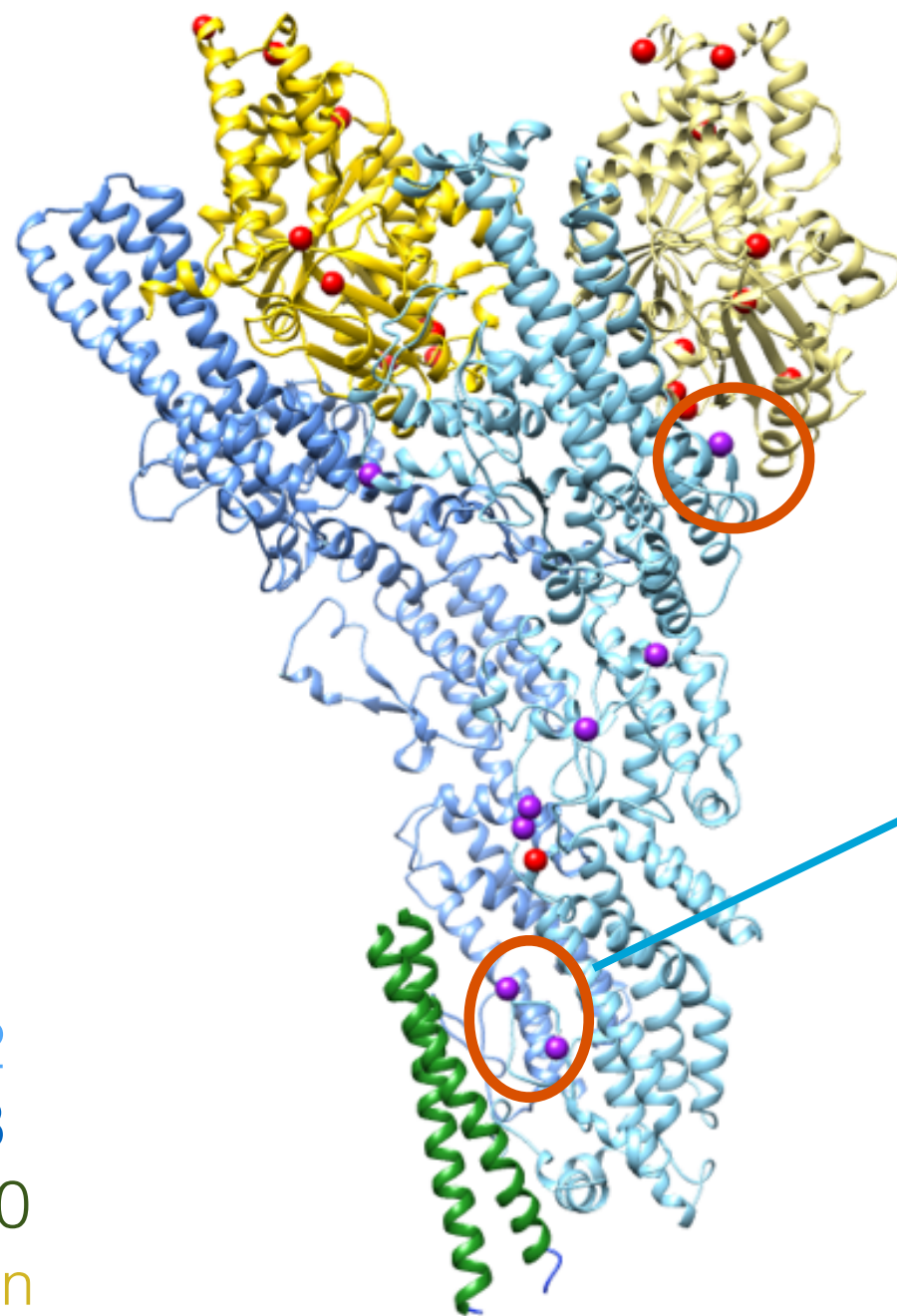
GCP3

Spc110

γ -tubulin

new phos

GCP2 phospho sites suggests functional roles



GCP2

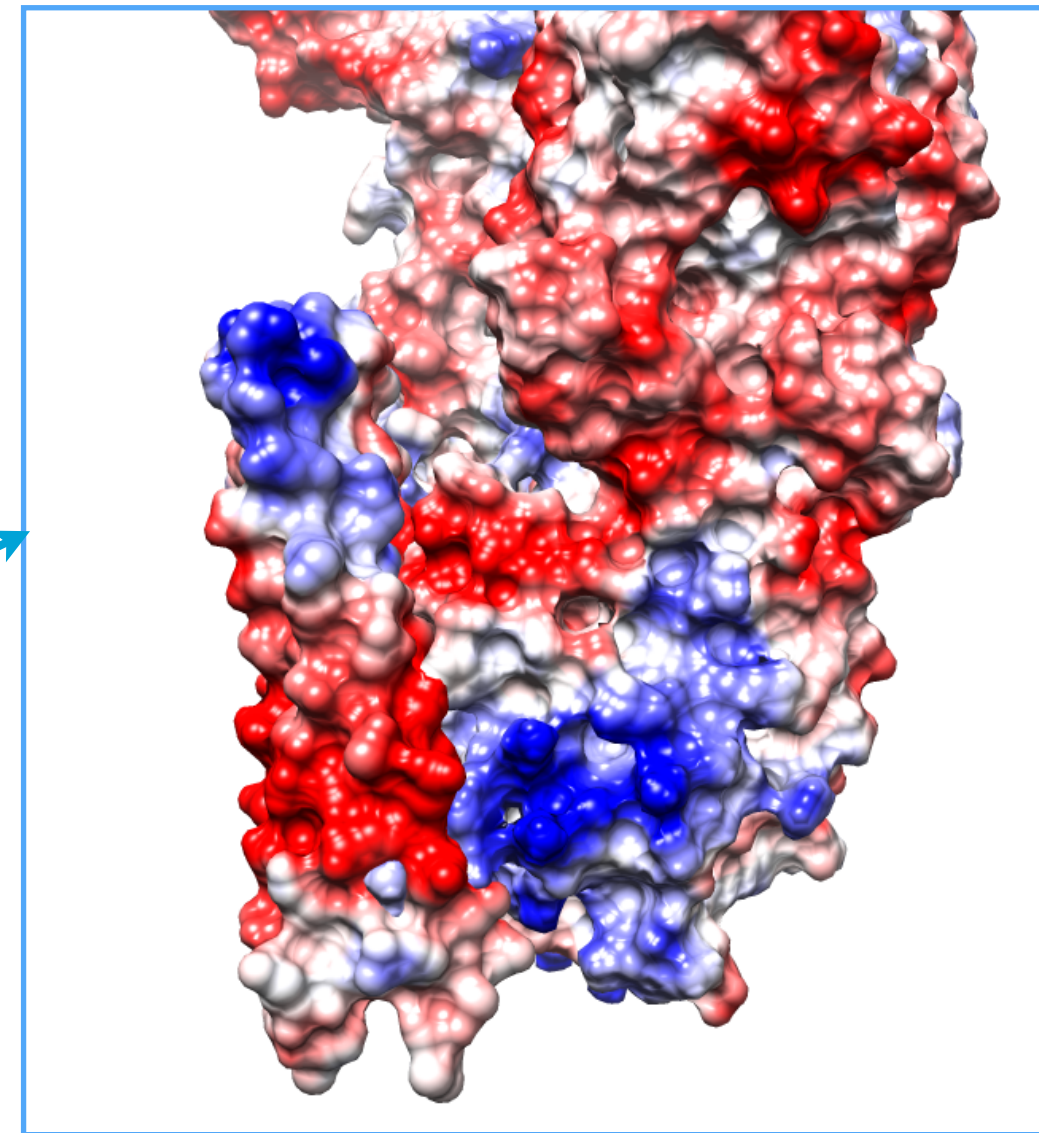
GCP3

Spc110

γ -tubulin

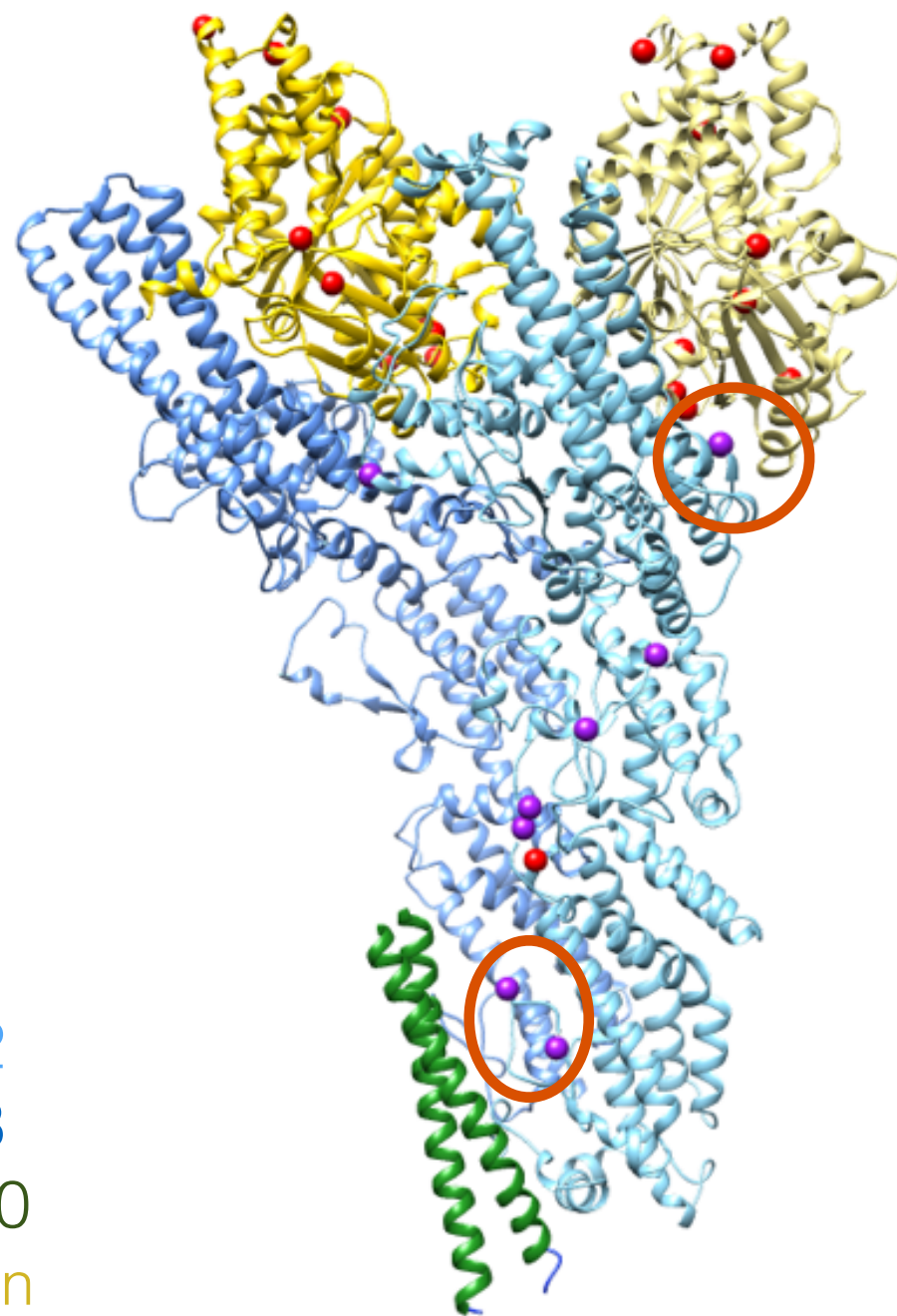
new phos

Spc110 binding



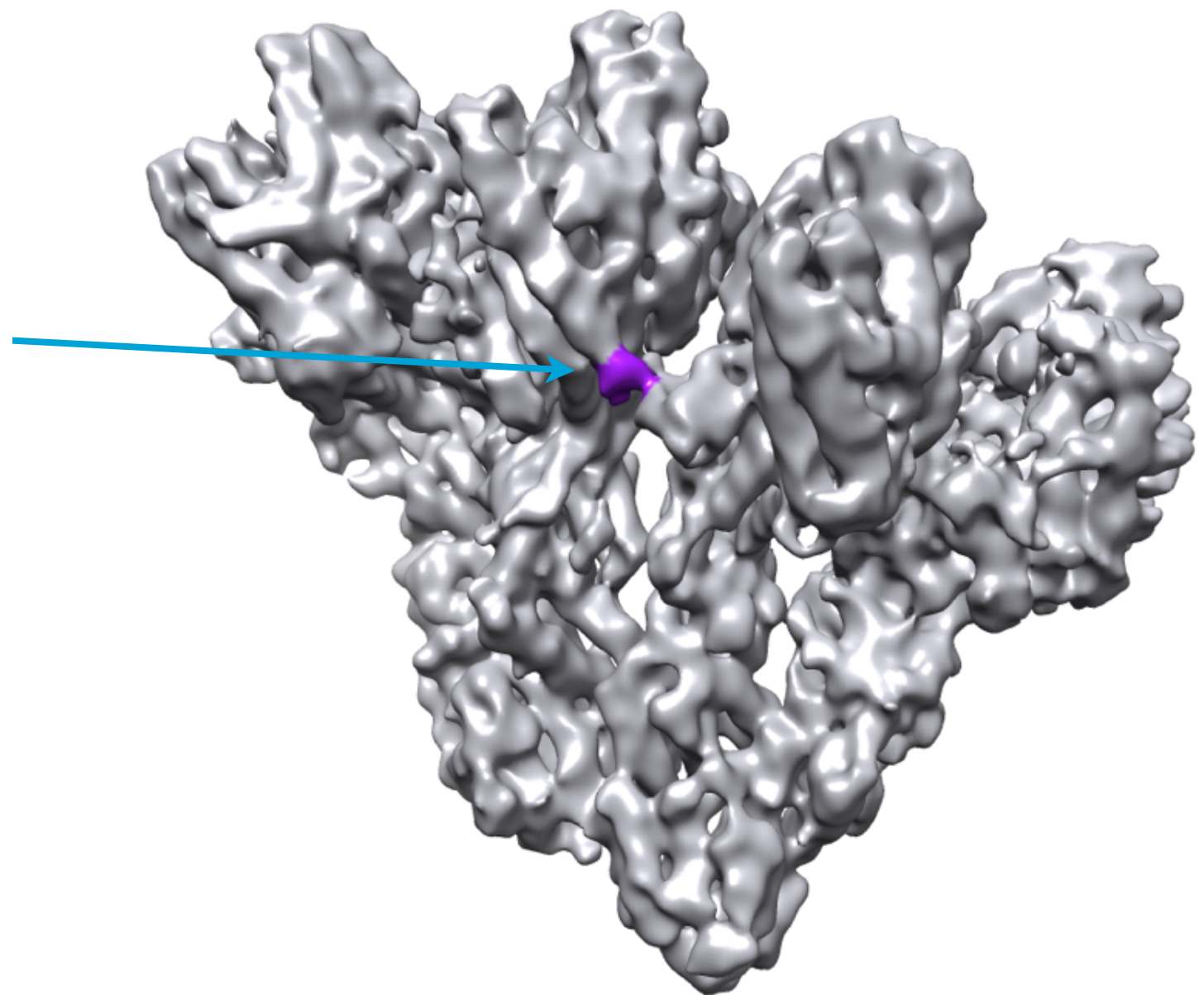
γ TuSC recruitment

GCP2 phospho sites suggests functional roles



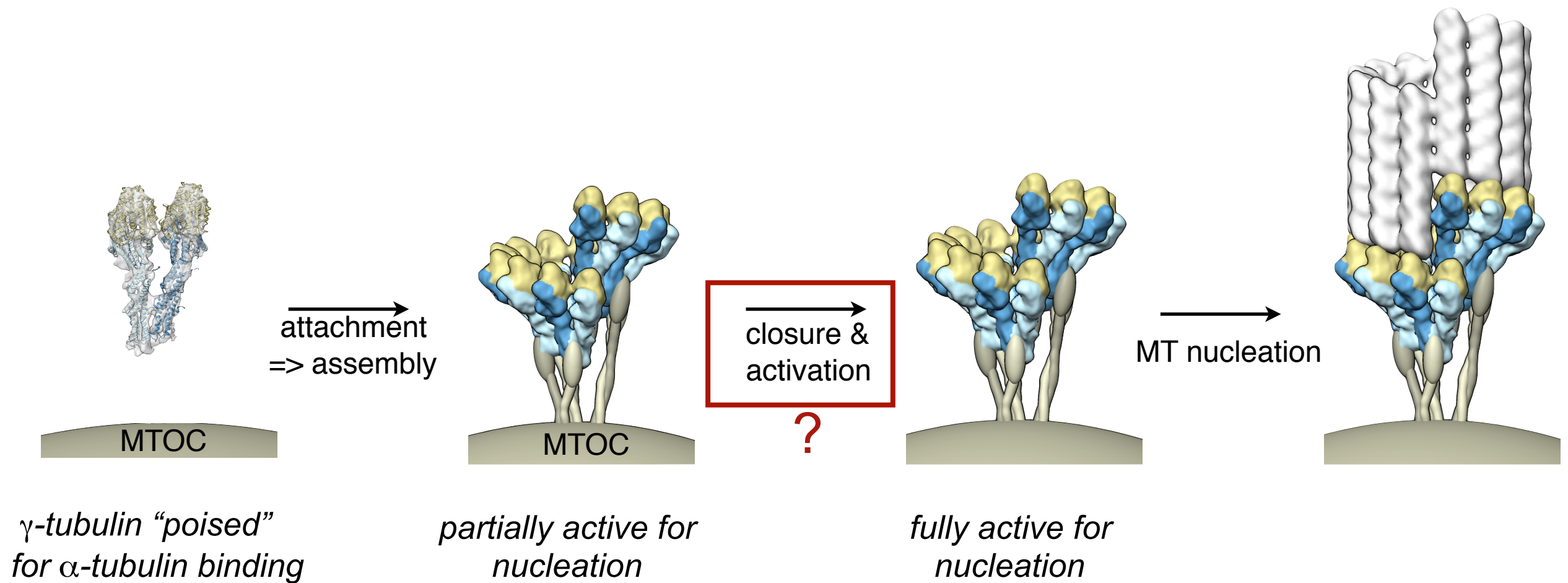
GCP2
GCP3
Spc110
 γ -tubulin
new phos

Spc110 binding



γ TuSC assembly

Models for γ -complex mediated attachment and nucleation



Acknowledgements

UCSF

Agard lab Centrosome/MT team

David Agard
Rose Citron
Andrew Lyon
Michelle Moritz
Sam Li
Ray Wang
Mariano Tabios

EM Core

Michael Braunfeld
Alex Myasnikov
David Bulkley
Cameron Kennedy
Matthew Harrington

Andrej Sali Lab

Charles Greenberg
Shruthi Viswanath

Beyond

Davis Lab, Univ. of Washington

Trisha Davis
Eric Muller
Tamira Vojnar
Genevieve Morin
King Yabut
Kim Fong
Alex Zelter
Richard Johnson
Connie Peng/David Drubin UCB

SPB PO1 Group

Mark Winey
Trisha Davis
Chip Asbury
Ivan Rayment
Andrej Sali
Sue Jasperson