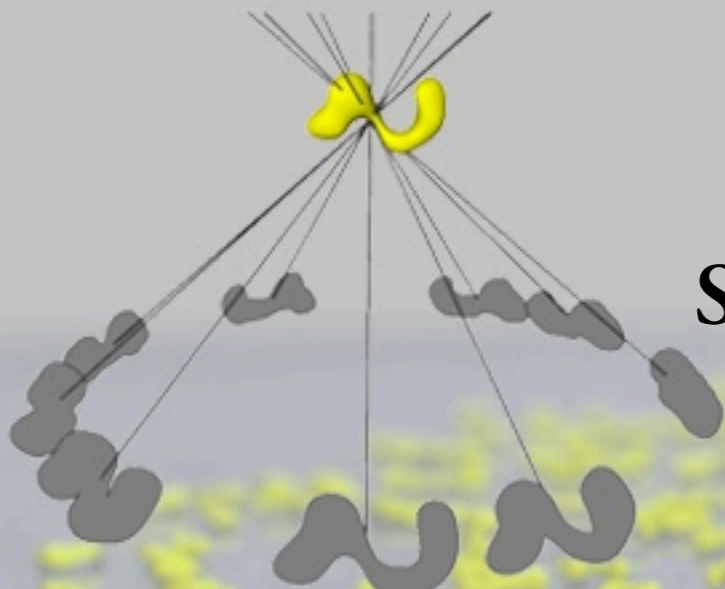


**Dmitry Lyumkis**

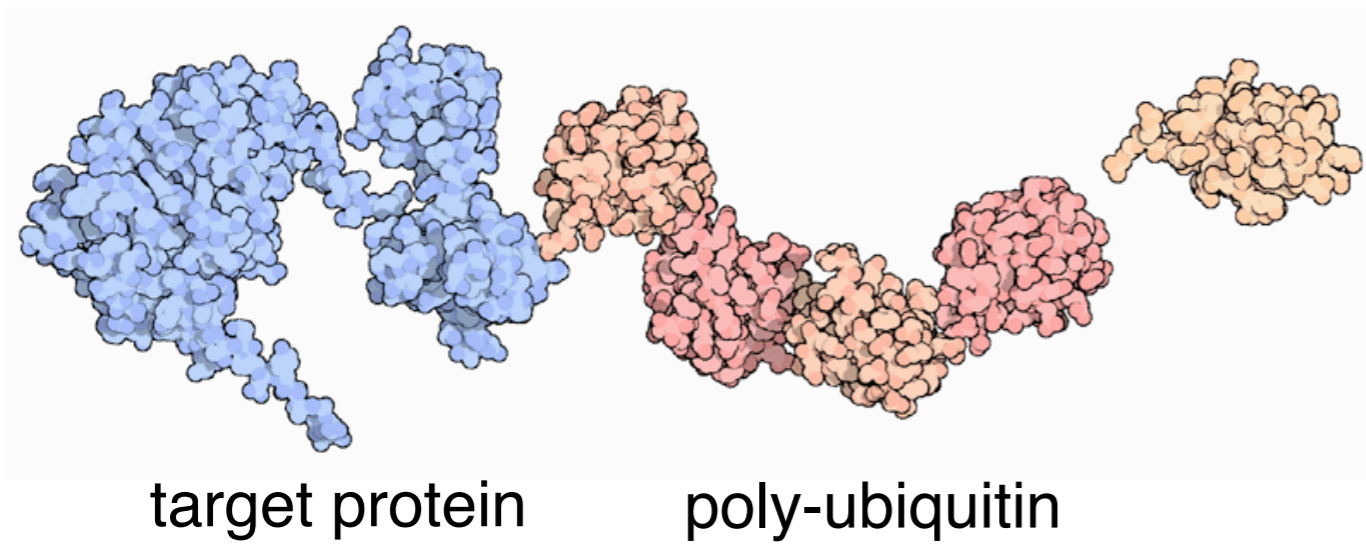
National Resource for Automated Molecular Microscopy

**Single-Particle EM Reveals Large-Scale Conformational  
Variability of the Ltn1 E3 Ligase**

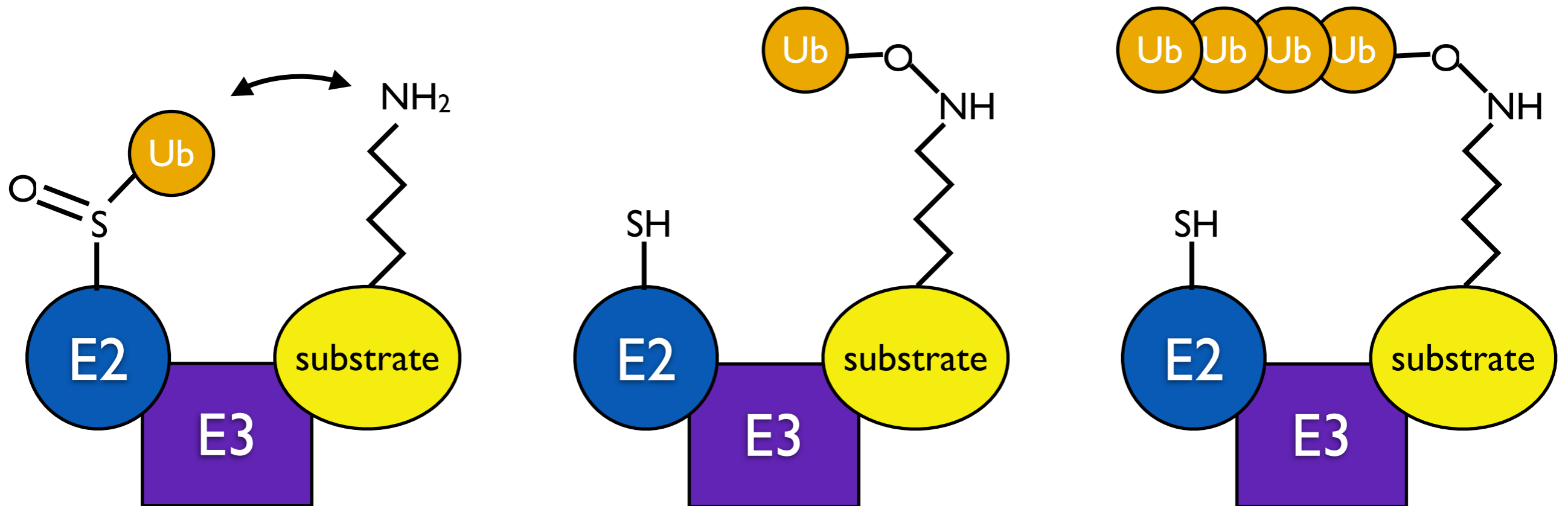


NRAMM cryo-EM workshop  
November 14, 2012

# E3 ubiquitin ligases

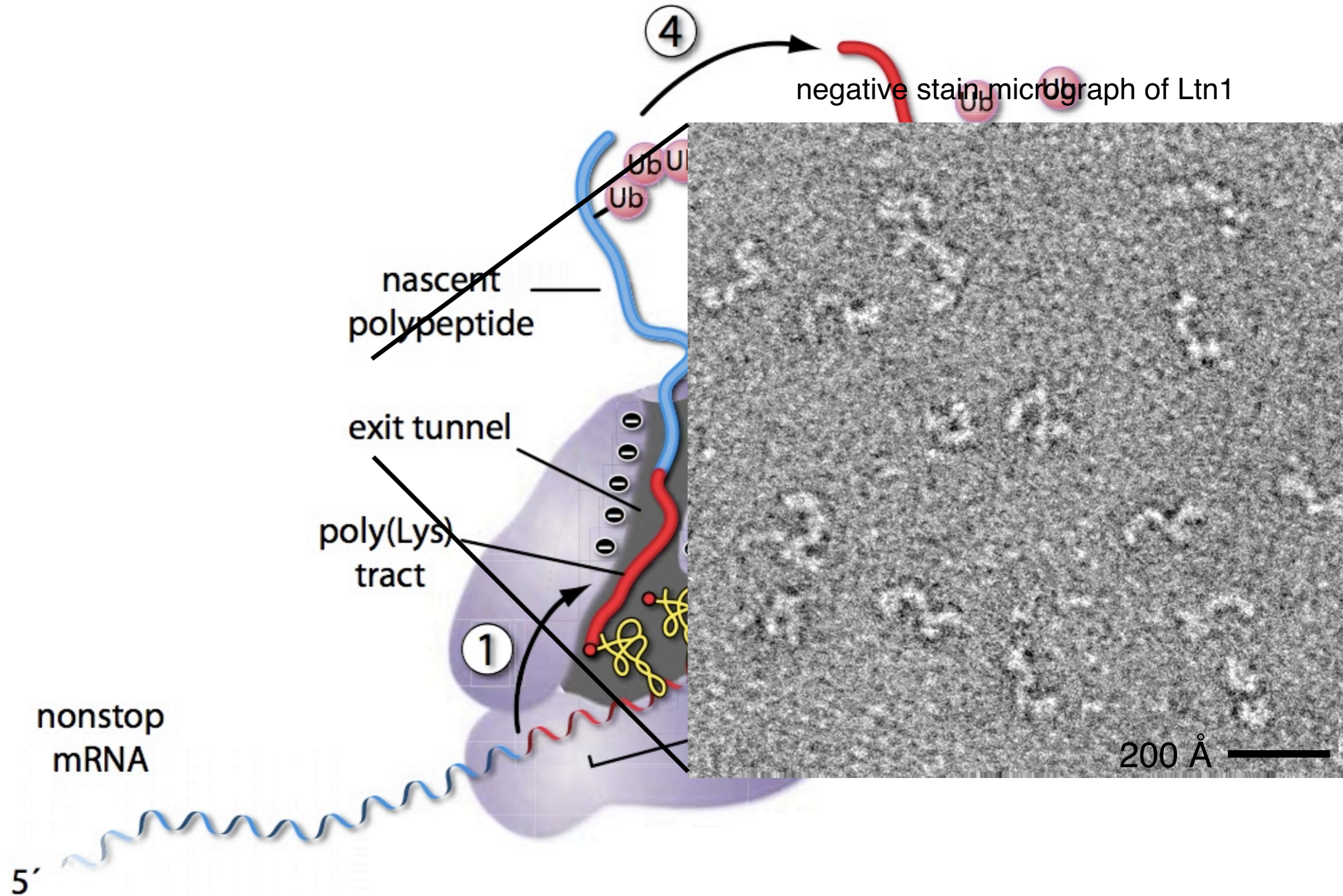


1. proteasome-mediated protein degradation
2. progression of the cell cycle
3. apoptosis
4. DNA transcription and repair
5. ... and virtually every process in the cell





# Ltn1 is a RING-E3 ubiquitin ligase that is responsible for protein quality control in eukaryotes

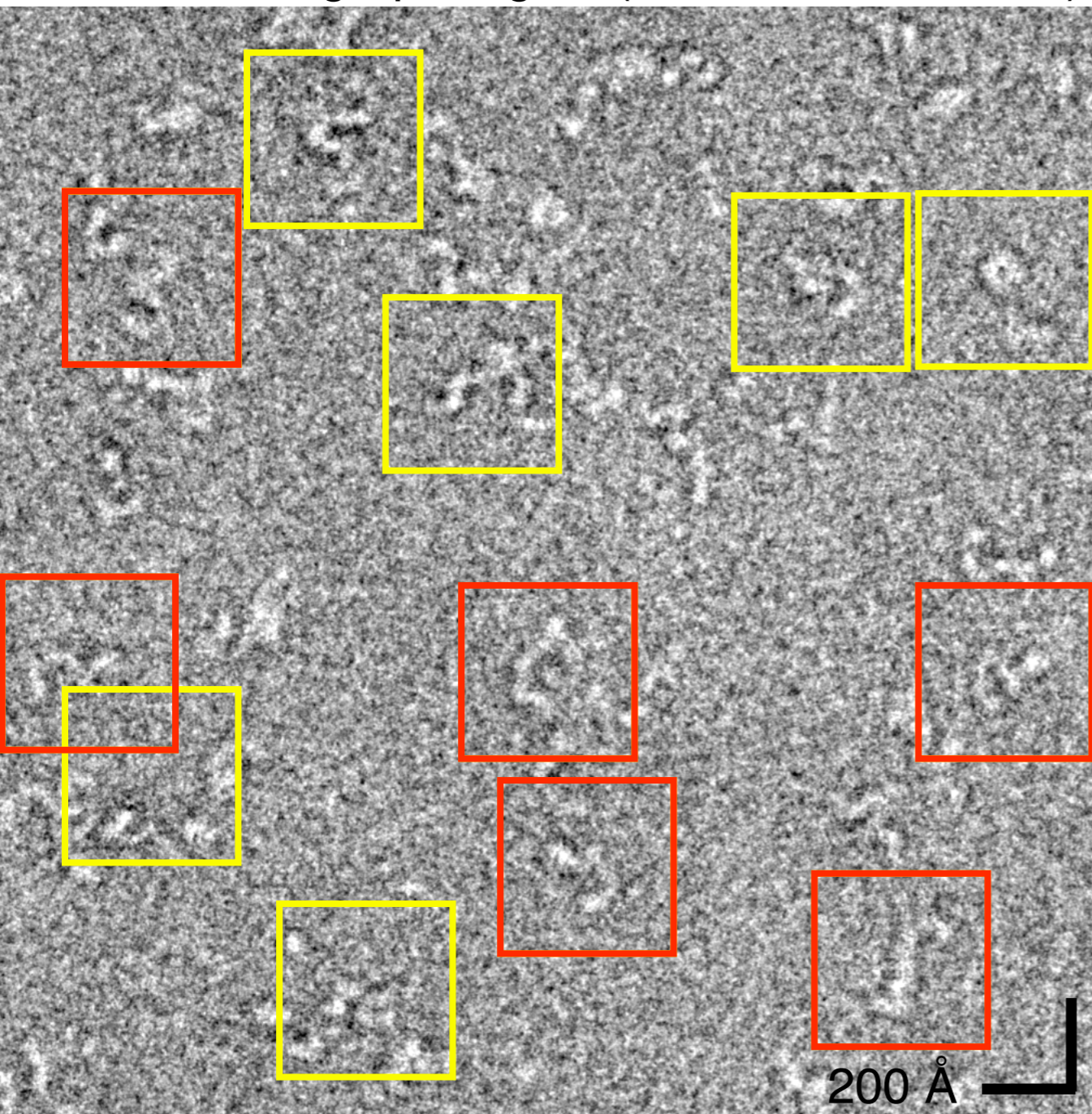


Bengtson, M. H. & Joazeiro, C. A. *Nature* **467**, 470-473 (2010)

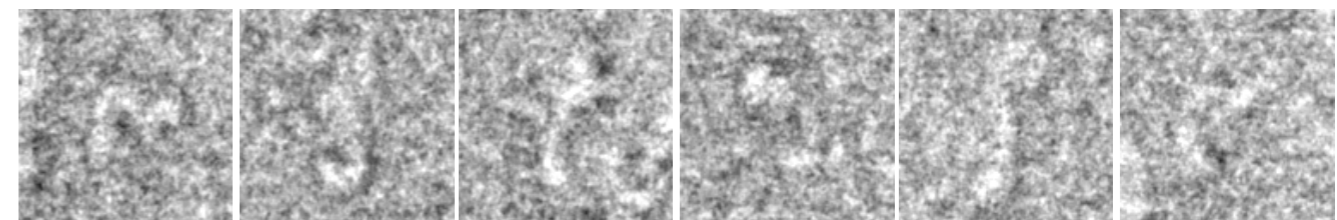
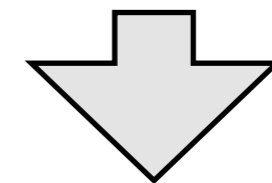
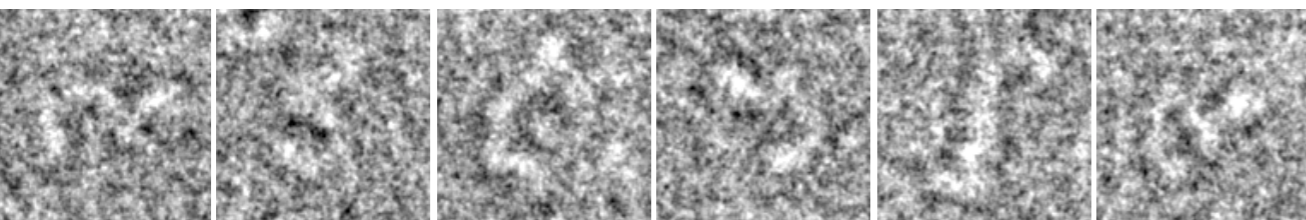
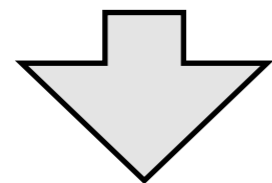
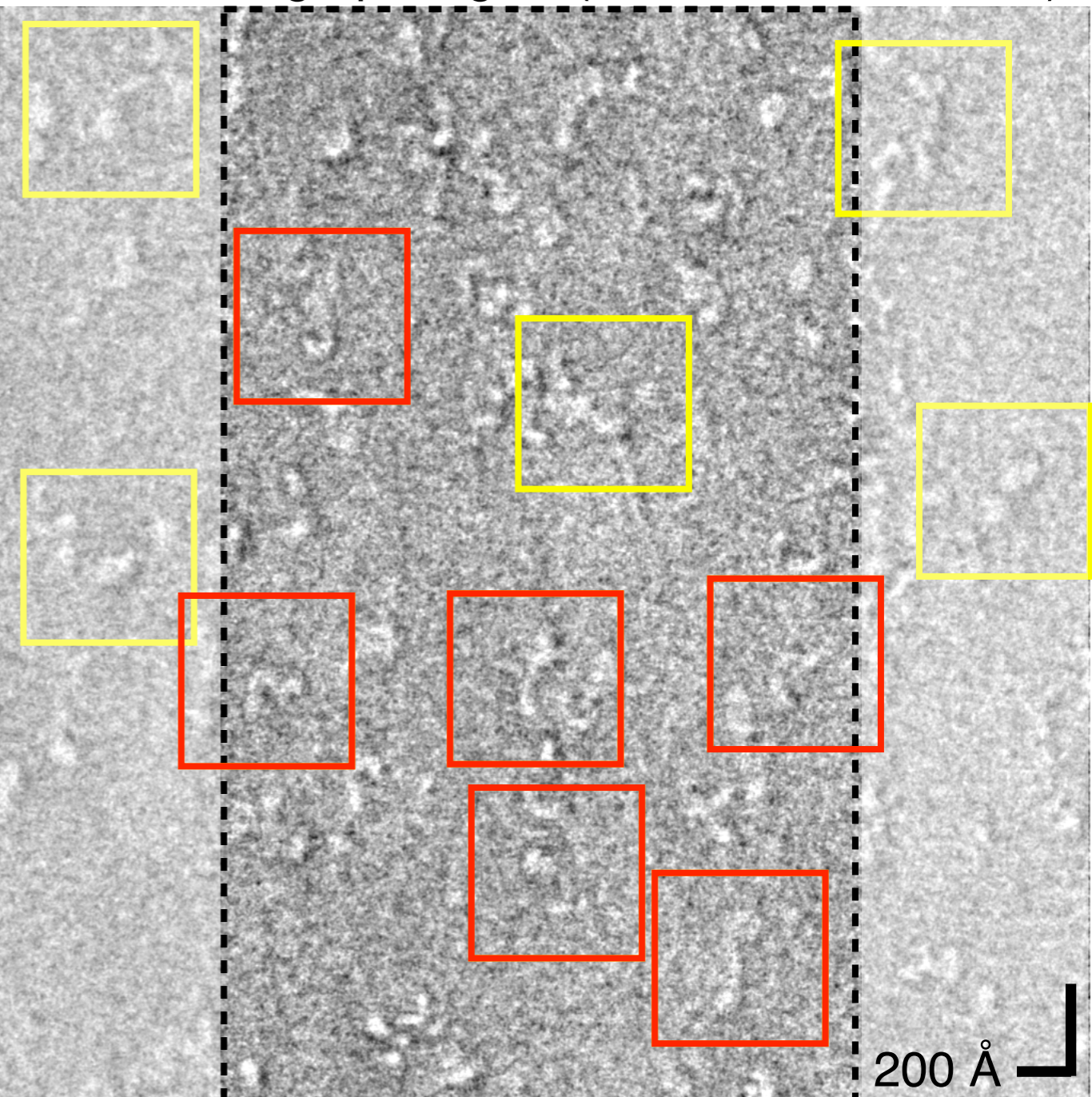


# data collection and particle extraction

untilted micrograph region ( $0^\circ$ , 2 $\mu\text{m}$  def, 2% UF)

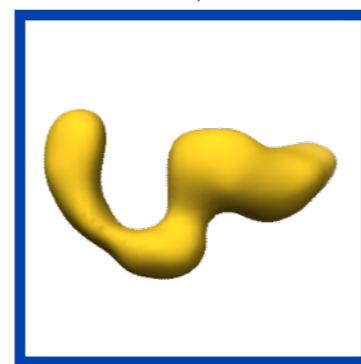
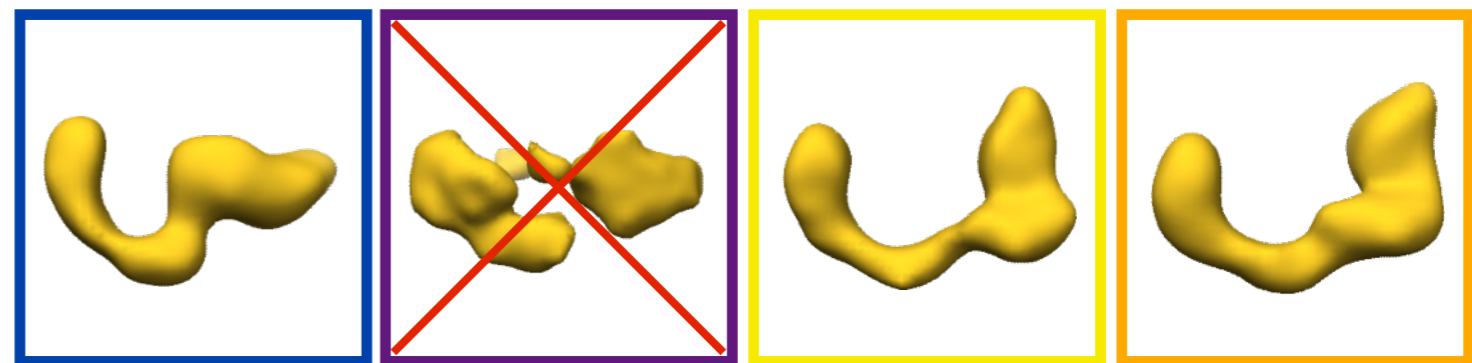
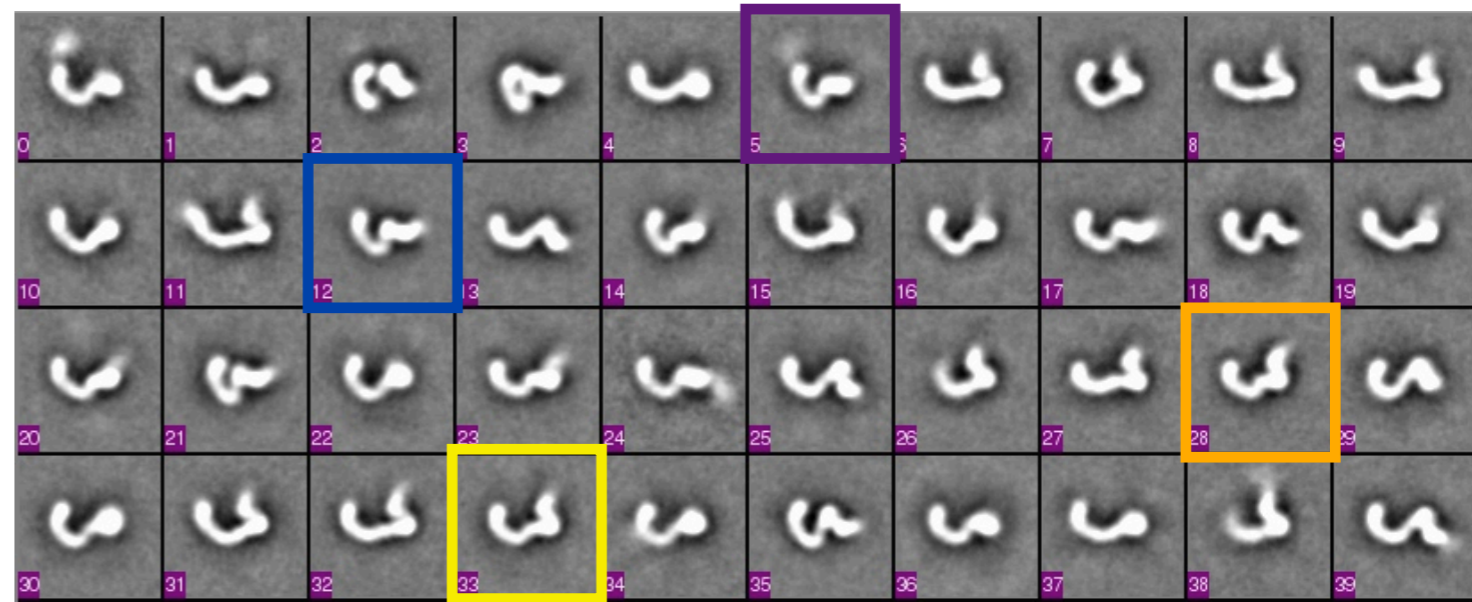
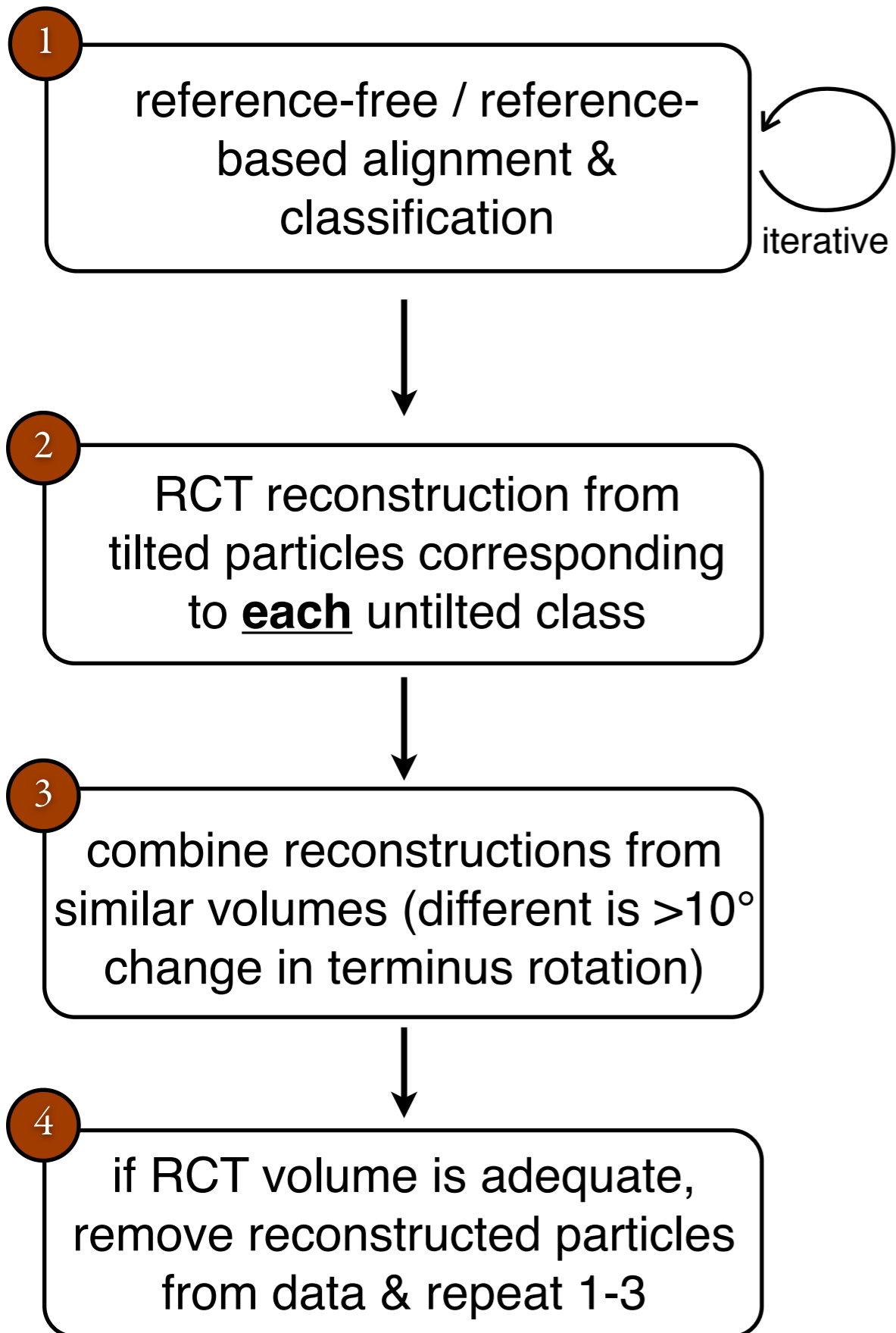


tilted micrograph region ( $55^\circ$ , 2 $\mu\text{m}$  def, 2% UF)

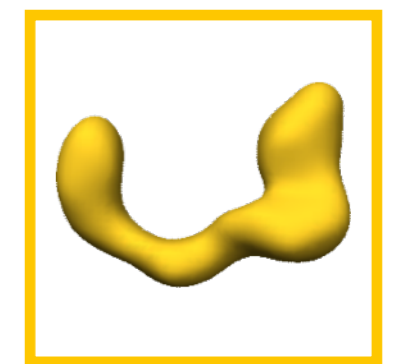




# alignment, classification, & 3D reconstruction



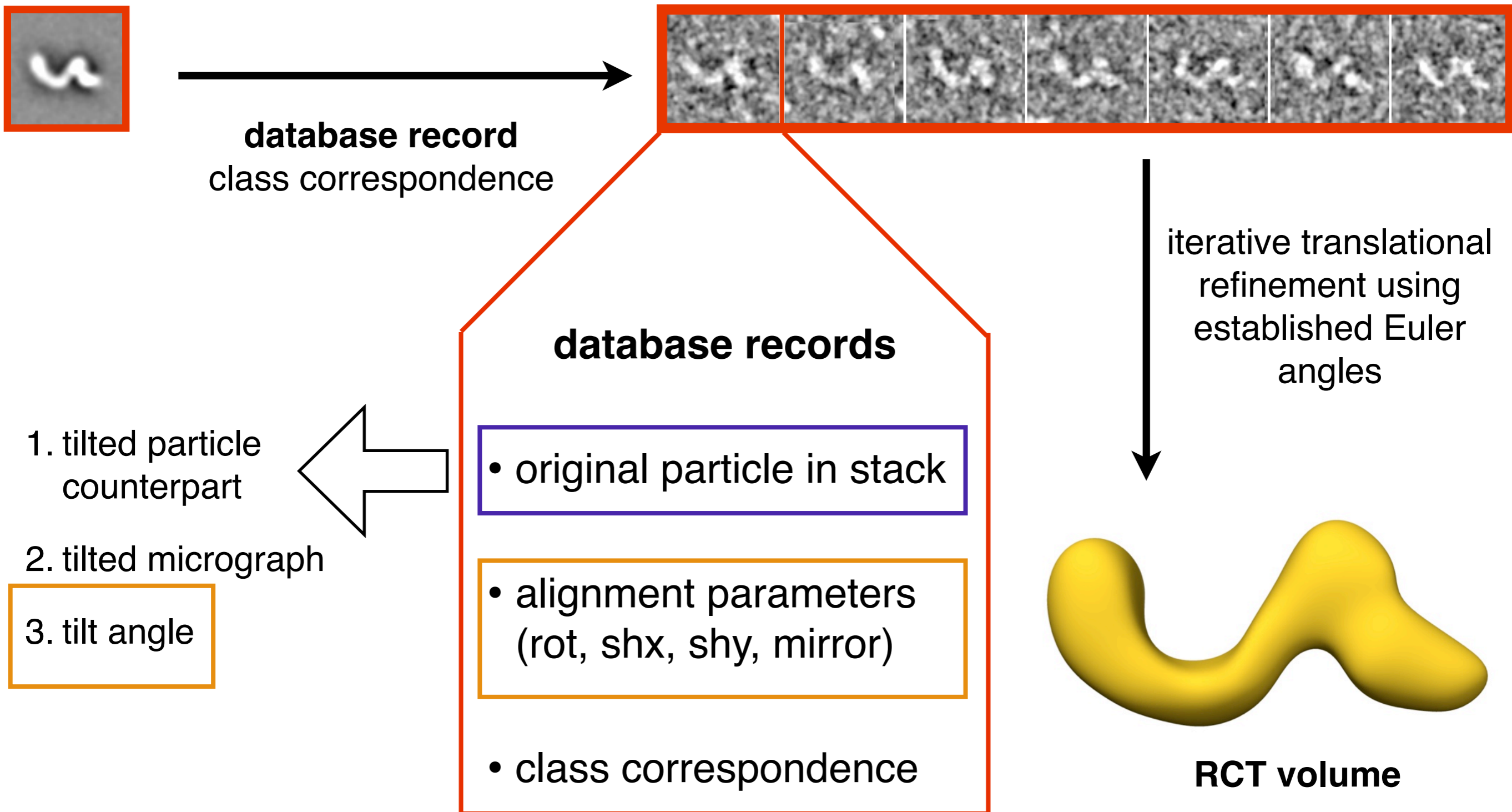
**final volume:  
conformation 1**



**final volume:  
conformation 2**



# automated RCT infrastructure in Appion





# automated RCT results

QuickTime Player File Edit View Share Window Help Stop Recording (84%) Sat 5:50 PM Administrator

RCT Volume Summary

image viewer RCT Volu... RCT Run Re... makeDDRaw... Particle Sele... Hierarchical ... Stack Report Hierarchical ... Hierarchical ... http://...10.hed

cronus3.scripps.edu/betamyamiweb/processing/rctsummary.php?expld=8589 Google

Most Visited Getting Started Latest Headlines database Bookmarks

**RCT Volume Summary Page** logged in as *dlyumkis* [Log Out]


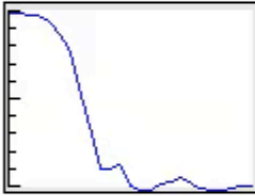
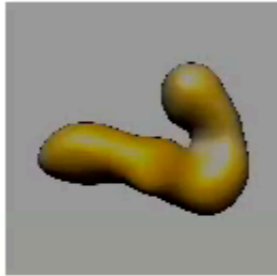
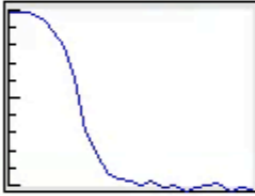
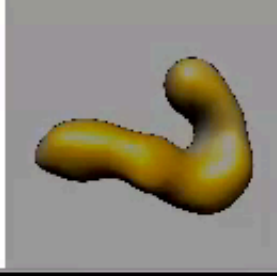
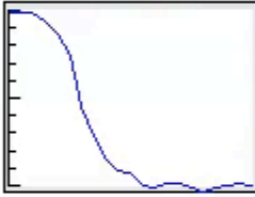
Project: Joazeiro - E3 ligase (224) [Appion Stats]  
 Session: **11mar23d** - data collection: free Listerin from Chris Lima, grid B1 from 11mar10c [User Guide]  
 Image Path: /am/data15/legnon/11mar23d/rawdata

Hide | Expand | Contract

Object Selection : 8

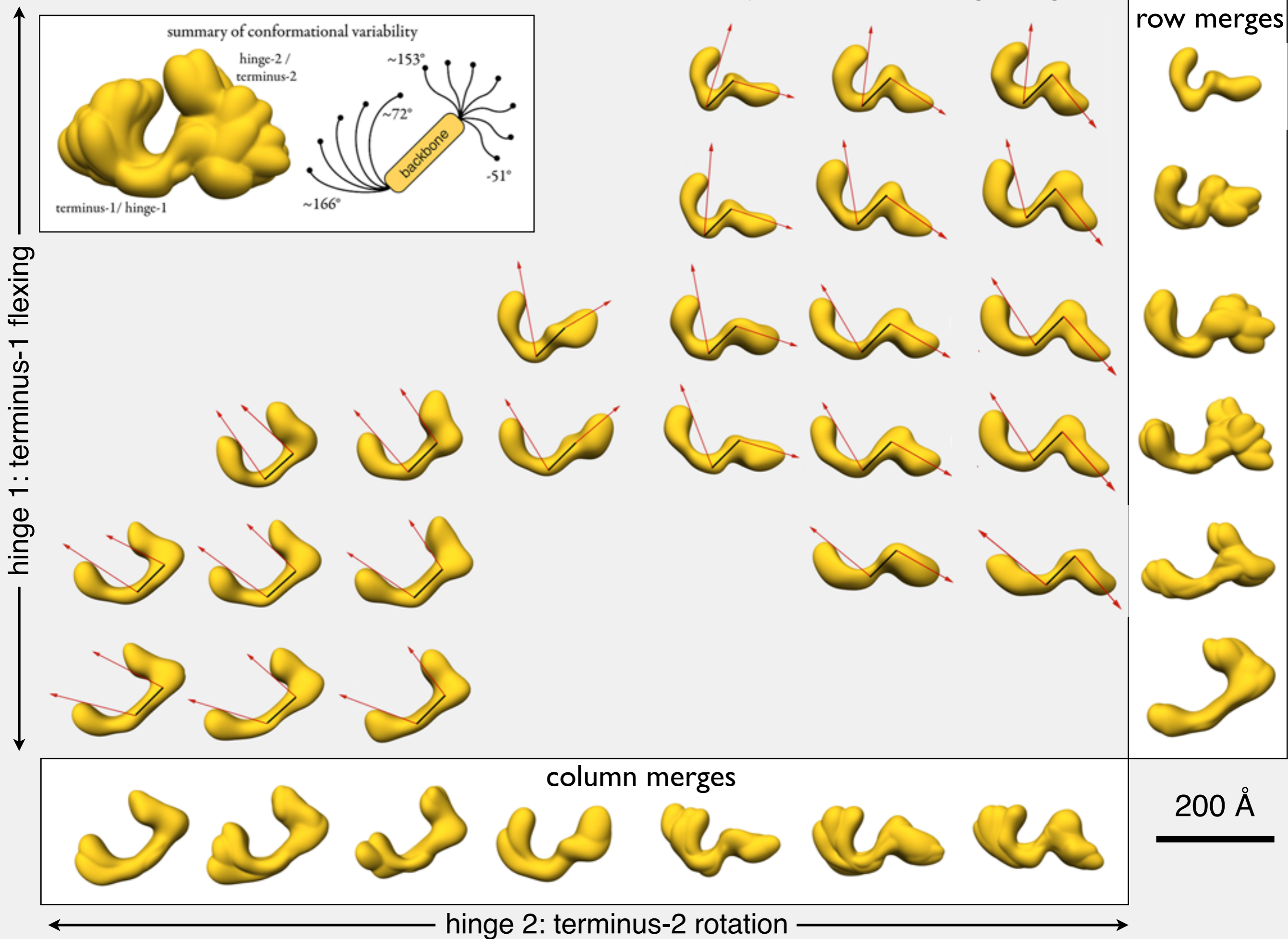
- Template Picking: 1 complete
- DoG Picking: 1 complete
- Signature
- Manual Picking
- Object Tracing
- Repeat from other session
- Align and Edit Tilt Pairs
- Auto Align Tilt Pairs: 4 complete
- CTF Estimation**
- Estimate the CTF: 1 complete
- Repeat from other session
- Stacks : 20**
- Stack creation: 20 complete
- more stack tools
- Particle Alignment**
- Run Alignment: 32 complete
- Run Feature Analysis: 15 complete

[Show 196 hidden rct runs]

defid	name	image	num part	pixel size	box size	fsc res	rmeasure	description
648 <small>hide</small>	<a href="#">rct1clust309class192833</a>		1,028 of 66,718	2.18 Å	160	42.14 Å 	18.59 Å	class refinement: combining classes 19 (rct264), 28 (rct273) & 33 (rct278) for increasing particle numbers in heatmap <small>edit</small>
505 <small>hide</small>	<a href="#">rct500clust463class245</a>		828 of 66,718	2.18 Å	160	47.41 Å 	36.34 Å	sub-classification: tightly folded particles (rebased19), 10 averages, class 2, 4, & 5 <small>edit</small>
504 <small>hide</small>	<a href="#">rct499clust463class24</a>		552 of 66,718	2.18 Å	160	51.01 Å 	16.45 Å	sub-classification: tightly folded particles (rebased19), 10 averages, class 2 & 4 <small>edit</small>



# Ltn1 shows extensive conformational variability about two hinge regions



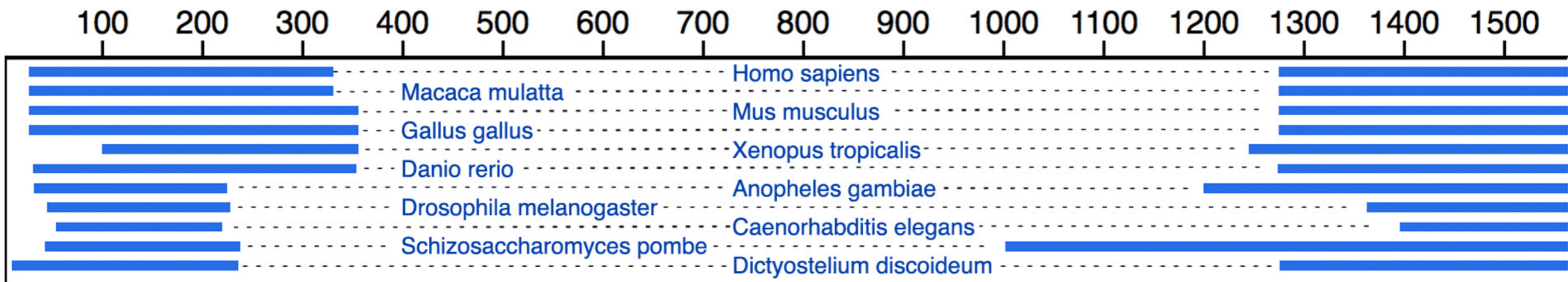


# Ltn1 contains 2 conserved termini

RING E3s are responsible for two functions:

- (1) substrate recognition
- (2) recruitment of E2-Ub conjugate

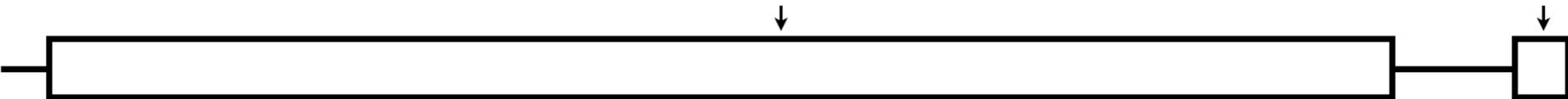
sequence conservation between orthologs



domain analysis

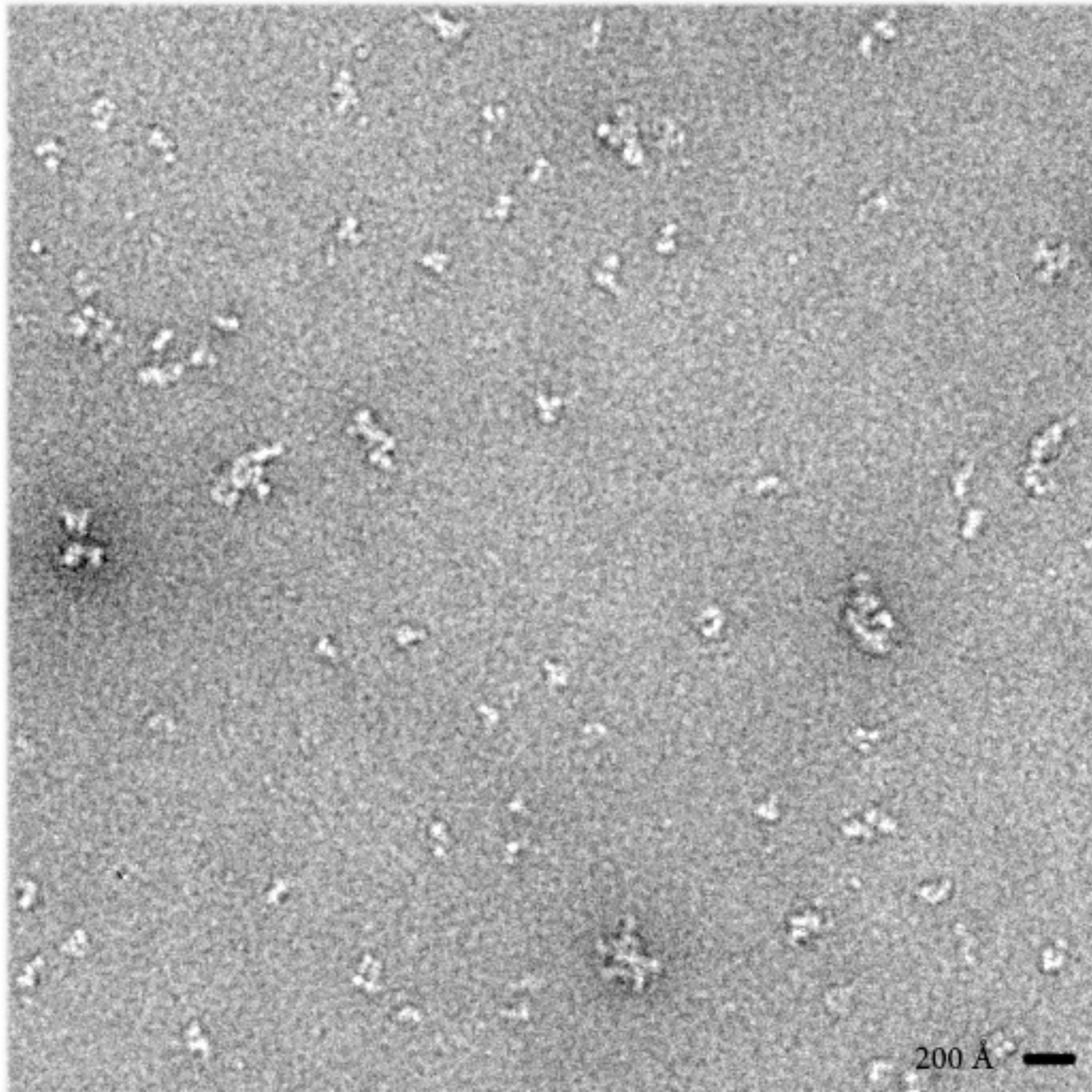
predicted HEAT repeats: aa 44-1390

C-terminal RING domain: aa 1509-1562





... so we made a truncated protein to orient the termini  
micrograph portion of  $\Delta N$ -Ltn1 (amino acids 477-1562)

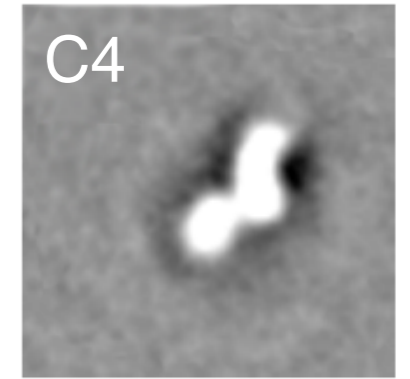
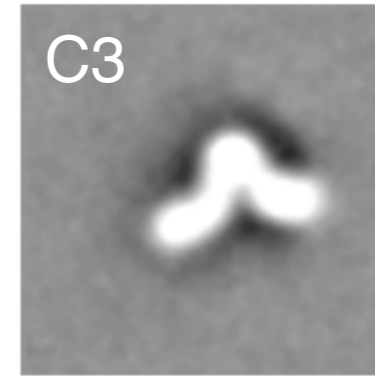
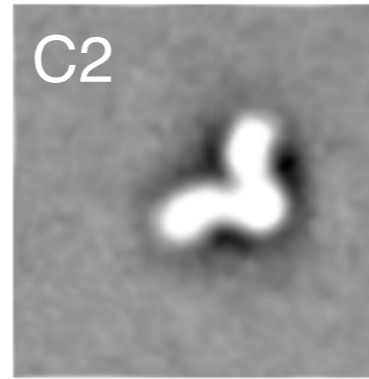
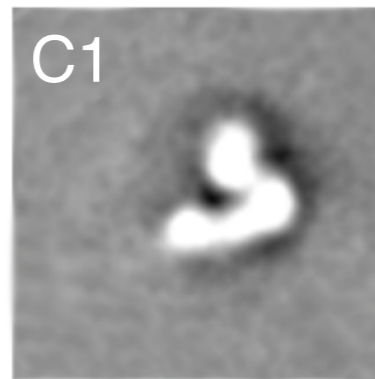




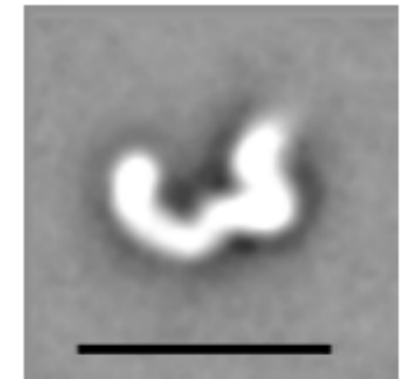
# Orientation of Ltn1's N and C termini

RING E3s are responsible for two functions: (1) substrate recognition, (2) recruitment of E2-Ub conjugate

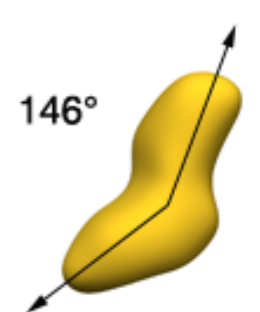
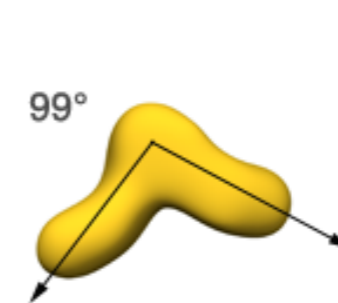
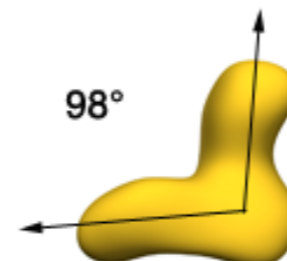
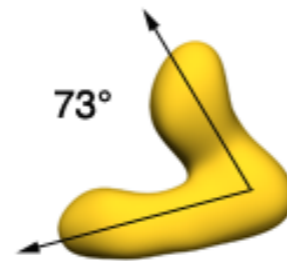
$\Delta$ N-Ltn1 Class Average  
(amino acids 477-1562)



Ltn1 Class Average  
(amino acids 1-1562)



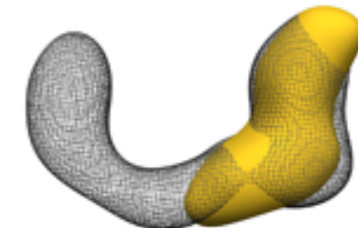
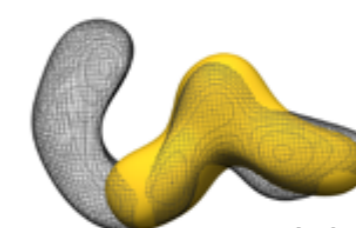
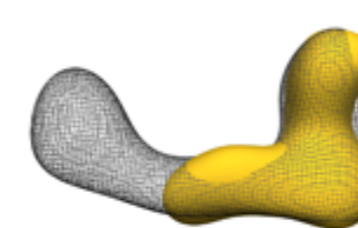
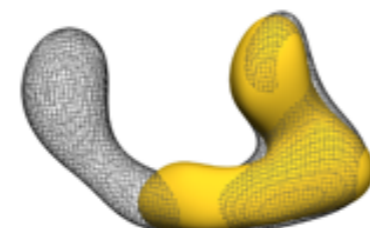
$\Delta$ N-Ltn1 3D reconstruction  
(amino acids 477-1562)



Ltn1 3D reconstruction  
(amino acids 477-1562)



overlay

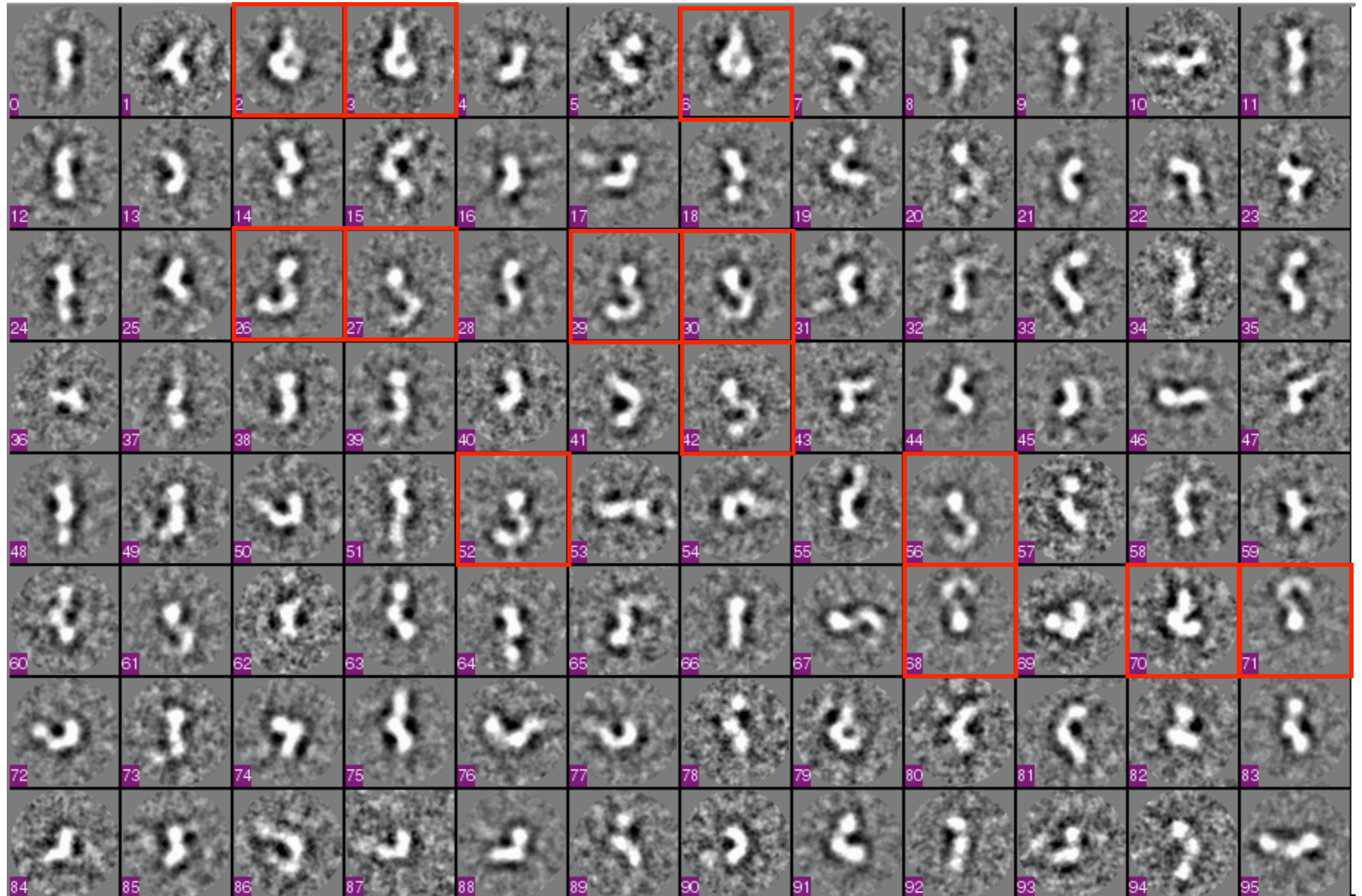


200 Å



# Ltn1 in vitreous ice

Does the same mobility occur in a more native environment?





# Mobility of Ltn1's termini is present in frozen hydrated preparations (cryo-EM)

**primary variance within data  
(intra-model heterogeneity)**

- (1) recalculate 10,000 volumes by bootstrap resampling  
(2) calculate variance

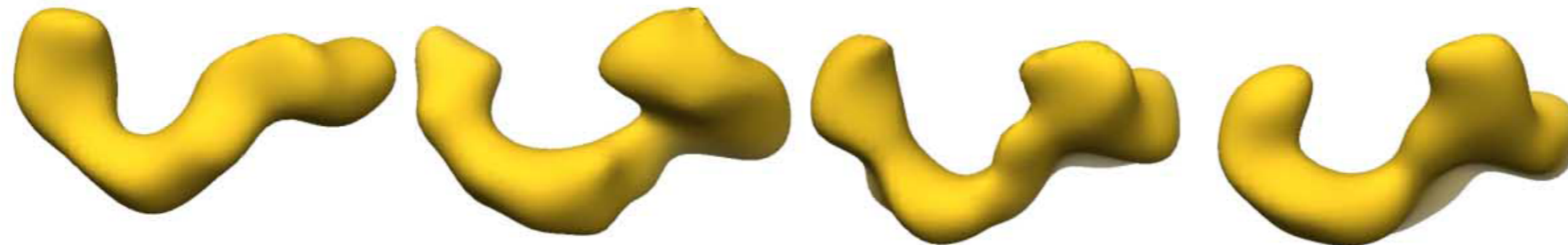
Penczek, P. A., M. Kimmel, et al. (2011). *Structure* **19**(11): 1582-1590



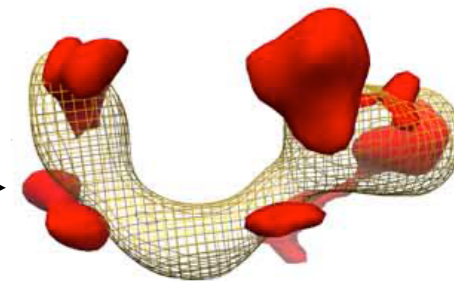
1 model

4 models

**variance between  
reconstructed volumes  
(inter-model heterogeneity)**



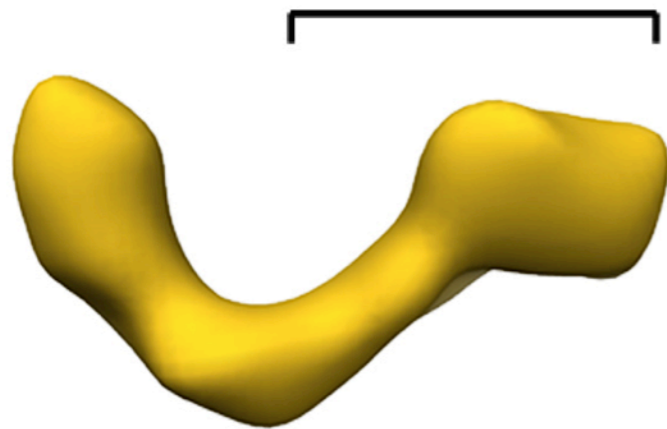
Scheres, S. H. (2012). *J Struct Biol*



- (1) Ltn1 exhibits similar heterogeneity in ice as it does in stain, and its basic architecture is preserved
- (2) predominant heterogeneity within the structure accounts for the mobility of its C-terminus

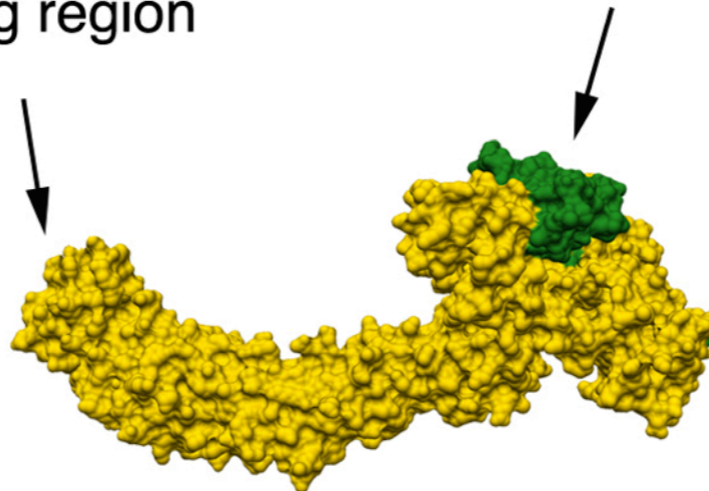
# Ltn1 is similar to CRL E3s in overall shape and size

RING-containing half  
E2 binding region



RCT reconstruction 1

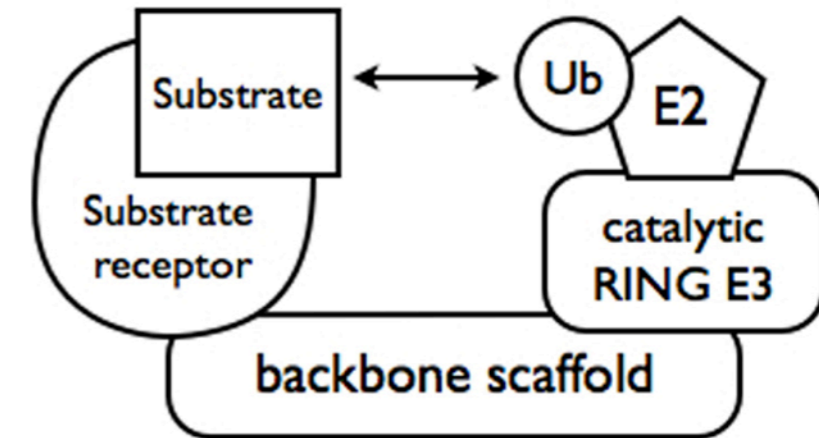
adaptor /  
substrate receptor  
binding region



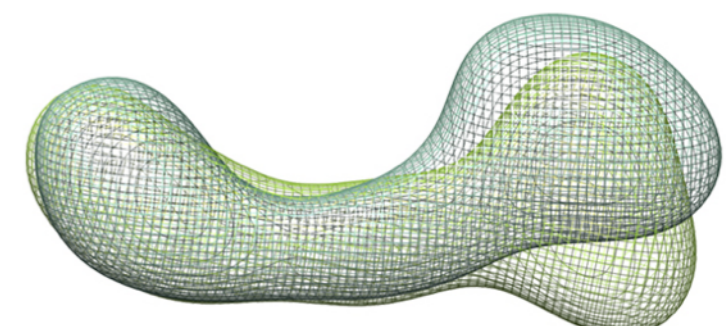
Zheng, N., et al. (2002). *Nature* **416**(6882): 703-709

RCT reconstruction 2

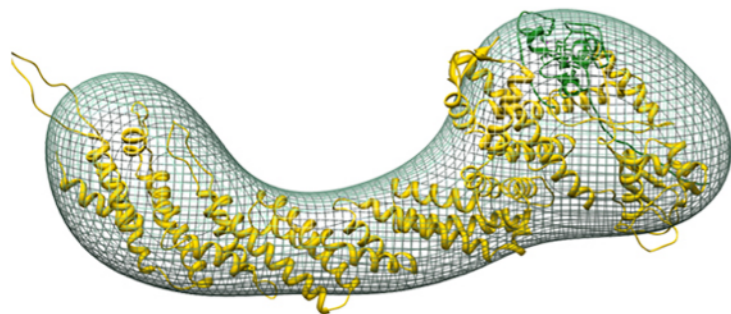
RING-containing subunit  
E2 binding region



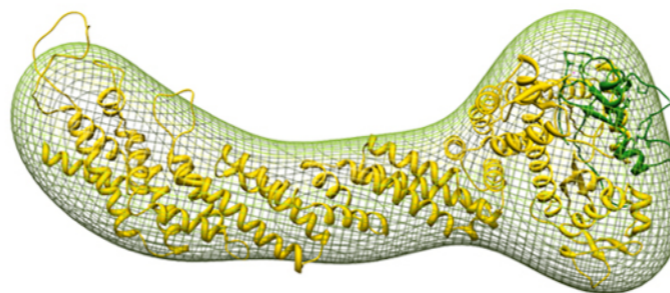
overlay



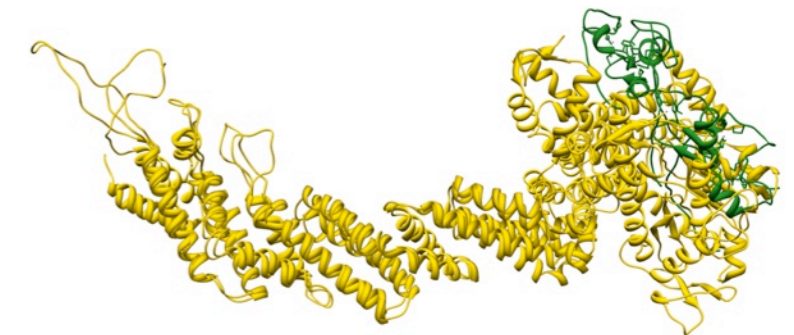
MD fit to 1



MD fit to 2



MD simulations

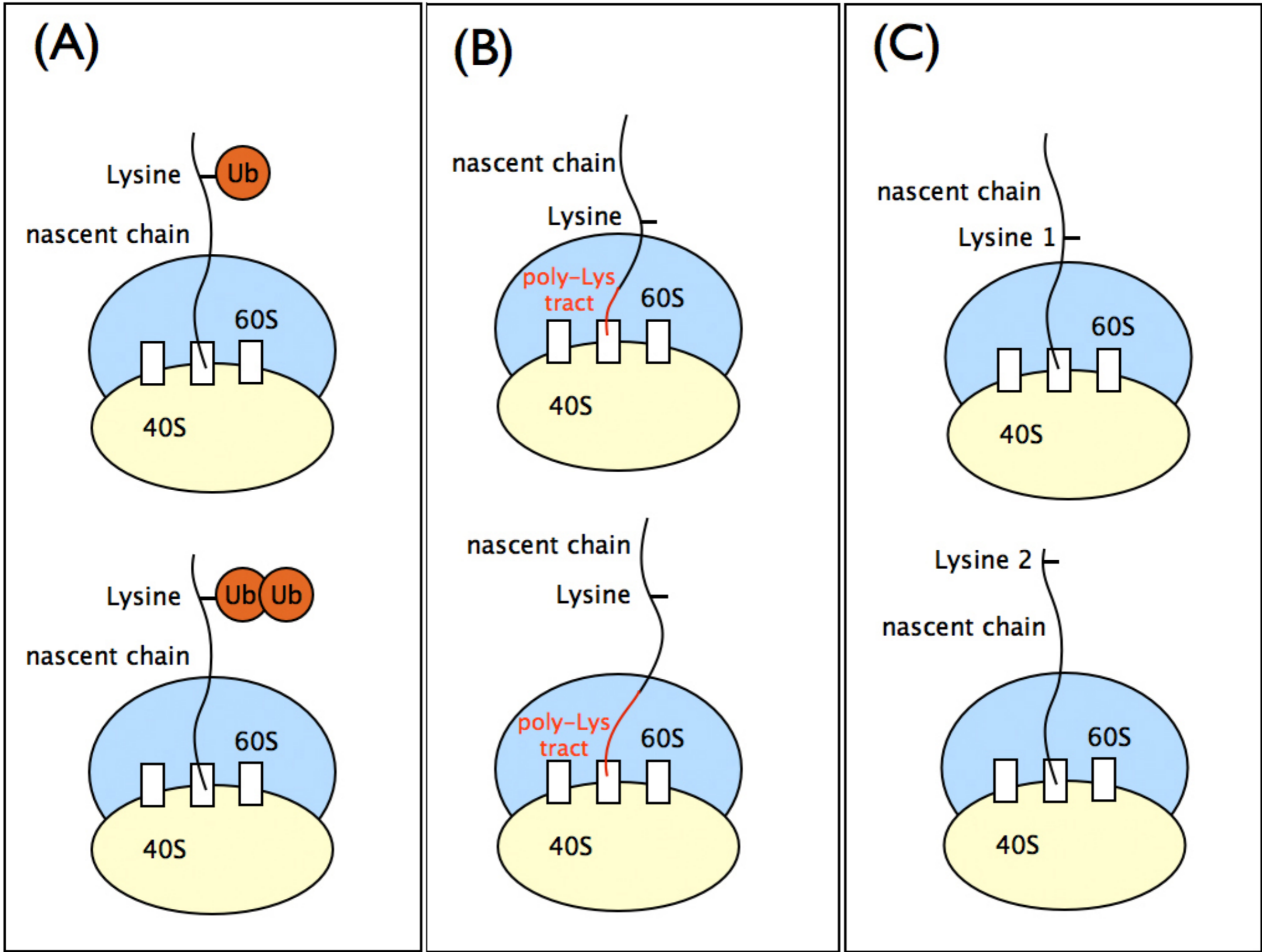


Liu, J. and R. Nussinov (2011). *JBC* **286**(47): 40934-40942

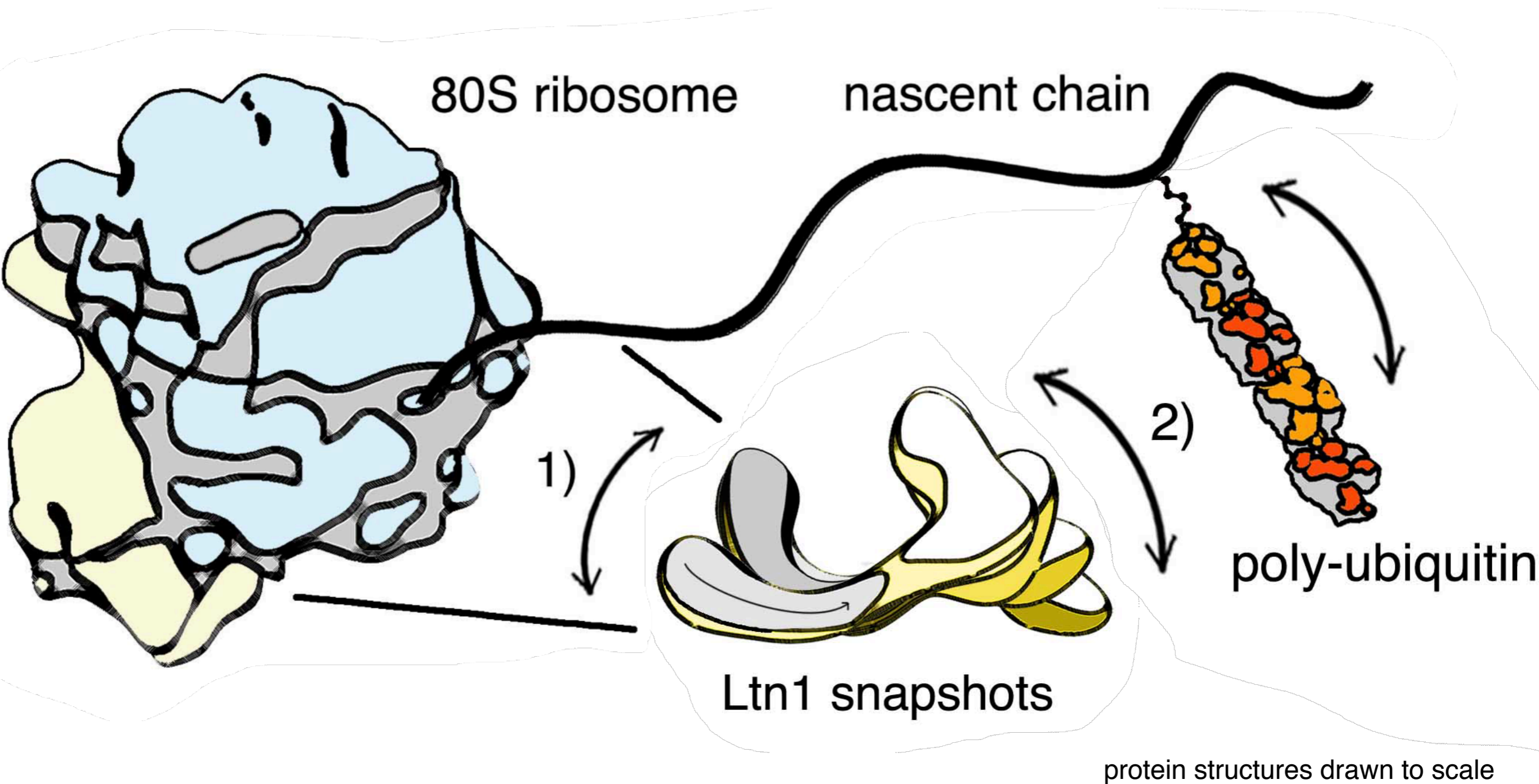
- (1) The employed strategy using negative stain is identifying real mobility within the protein
- (2) Ltn1's mobility cannot be simply explained by its large size and extended shape



# relevance of flexibility within the ribosomal context of protein quality control



# Model of Ltn1 E3 function within the context of protein quality control



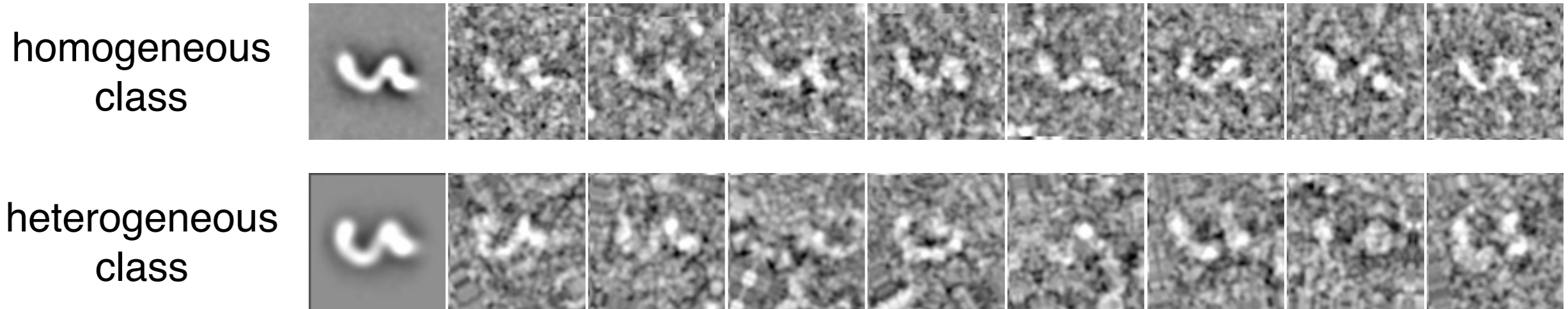
(1) mediate poly-ubiquitylation

(2) recognize **different** target lysines in a heterogeneous and dynamic environment specific to protein quality control

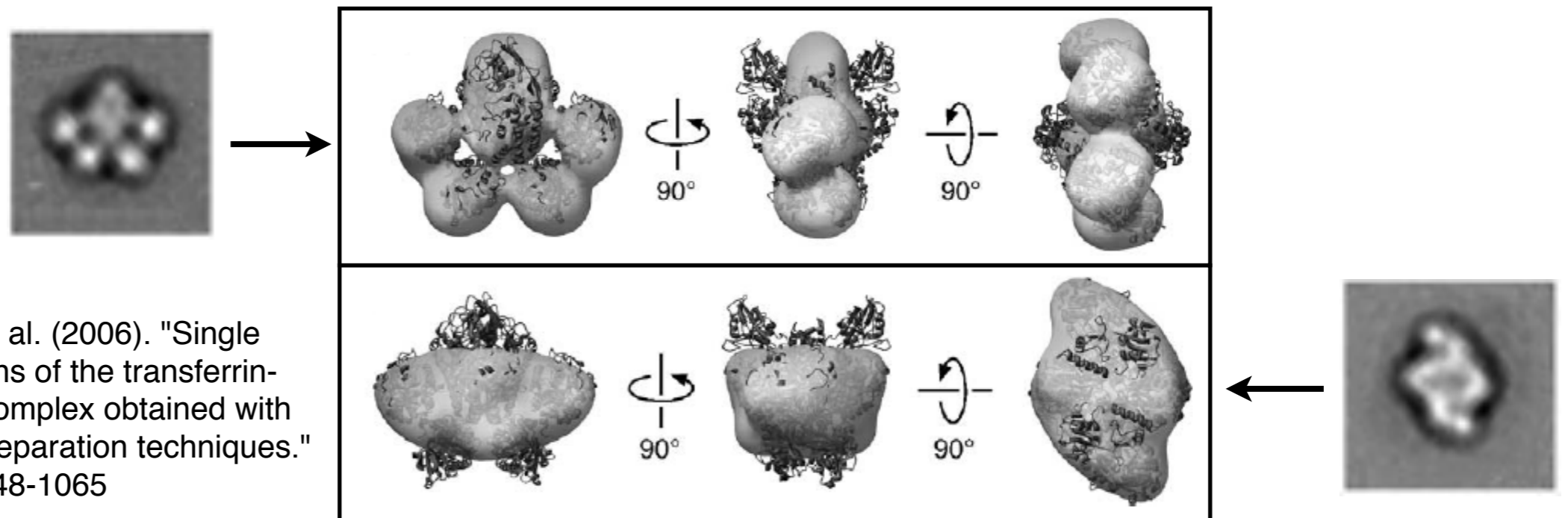


# validation / optimization of RCT results

1. look at the particles within your class



2. minor things - careful tilt-pair matching (especially when it is done automatically), FSC value, curve, etc.
3. effects of specimen flattening: different structure by RCT may not always mean different conformation and/or composition



# conclusions

1. Ltn1 is a highly dynamic protein and this may have functional implications for its role in protein quality control
2. Growing evidence that flexibility plays an important role in E3 ubiquitin ligases
3. RCT as a robust methodology for sorting heterogeneity in single-particle data
  - fair amount of automation (record keeping!)
  - data collection in stain  $\approx$  RCT
4. public availability of the negative stain dataset: <http://maskiton.scripps.edu>



# Thanks to ...

## Principal Investigators

Bridget Carragher  
Clinton Potter  
Ron Milligan  
Claudio Joazeiro

## Collaborators

Claudio Joazeiro  
Mario Bengtson  
Joong-Won Lee  
Chris Lima  
Selom Doamekpor  
Matt Petroski  
Tasha B. Toro

## AMI / Joazeiro groups

see below

