

2014 Workshop on Advanced Topics in
EM Structure Determination
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Discussion Group:

Model building, fitting & validation using high-resolution cryo-EM maps

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Good news: Technological and methodological improvements in signal detection and data processing have made it possible to determine biomolecular structures at (pseudo)atomic resolution.

However: A general perception is that the quality of atomic models derived from cryo-EM reconstructions is typically suboptimal, when compared to crystal structures obtained from X-ray diffraction at similar resolution.

“XPLOR-NIH (Maki-Yonekura et al., 2010), CNS (Cheng et al., 2011) and Phenix.refine (Baker et al., 2013) have previously been used for refinement of models into cryo-EM data by adopting a pseudo-crystallographic approach. However, many structures deposited alongside high-resolution (4 Å or better) cryo-EM reconstructions have not been refined and consequently have worse stereochemistry than crystal structures solved at similar resolutions.” (Brown et al., accepted)

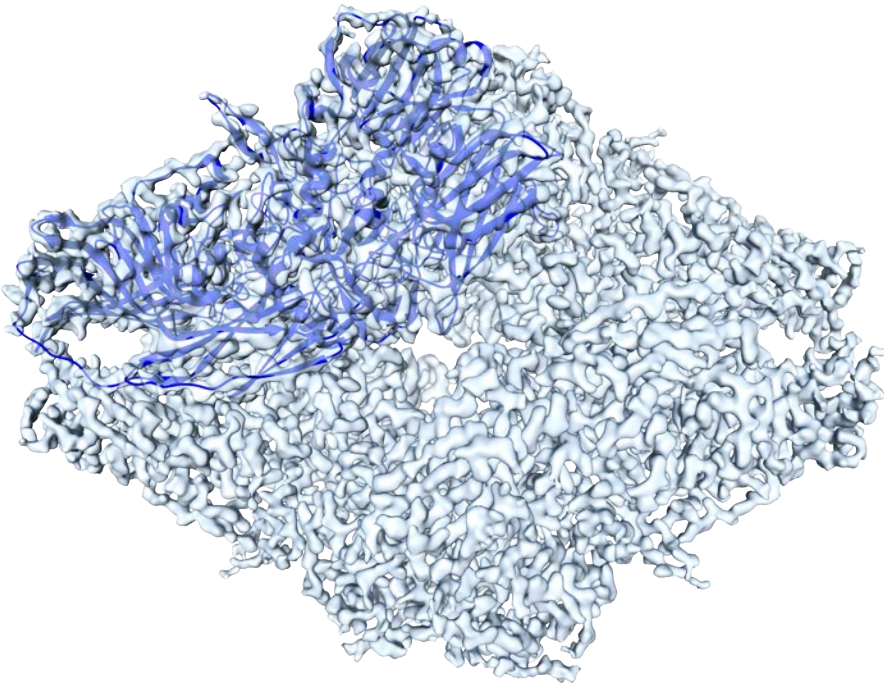
Are the existing tools for model building and validation adequate?

Have these tools been optimized for EM density maps?

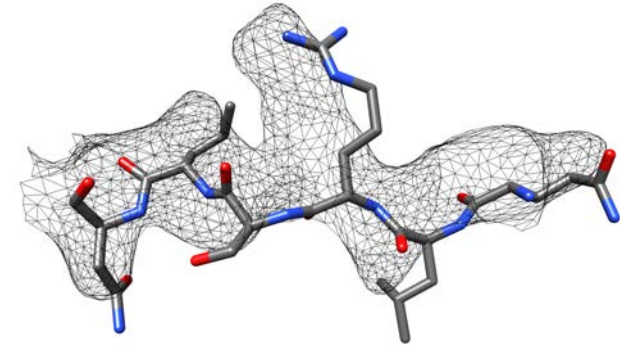
Is there a lack of awareness of what tools are already available?

Would it be desirable to create/adopt a standardized description of model quality?

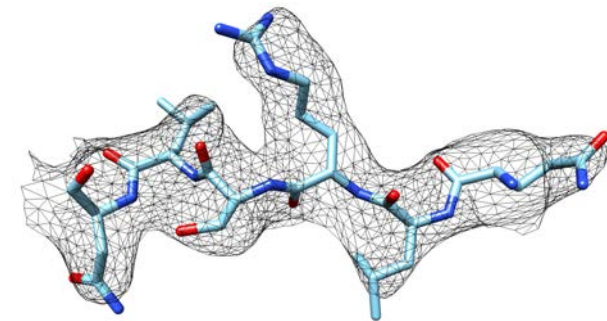
Reference	Target	Resolution	Fitting	Model building	Refinement	Validation
Miyazawa et al., 2003	Acetyl-choline receptor pore (2D)	4.0 Å	Program O	Program O	Program O	PROCHECK
Ludtke et al., 2008	GroEL	4.2 Å	COOT	SSEHunter COOT	COOT	Comparison to X-ray structure
Cong et al., 2010	Mammalian chaperonin TRiC/CCT	4.0 Å	Chimera COOT	MODELLER	COOT	COOT?
Yu et al., 2011	Cytoplasmic polyhedrosis virus (CPV)	3.1 Å	COOT	COOT REMO	CNS	(CNS force field)
Li et al., 2013	20S proteasome	3.3 Å	Chimera	-	MDFF	(MDFF force field)
Liao & Cao et al., 2013	Rat TRPV1 channel	3.3 Å		COOT	COOT	COOT?
Allegretti et al., 2014	F ₄₂₀ reducing hydrogenase	3.4 Å	Chimera COOT	COOT	COOT	COOT?
Amunts & Brown & Bai et al., 2014	Yeast mitochondrial large ribosomal Su	3.2 Å	Chimera? MOLREP COOT	RCrane I-TASSER	COOT REFMAC v.5.8 ERRASER-PHENIX	MolProbity FSC _{work} & FSC _{test}
Wong & Bai et al., 2014	Plasmodium falciparum 80S ribosome	3.2 Å	Chimera COOT	I-TASSER	REFMAC v.5.8 ERRASER-PHENIX	MolProbity, FSC _{work} & FSC _{test}
Voorhees & Fernandez et al., 2014	Mammalian ribosome in complex with Sec61	3.4 Å	Chimera COOT	COOT	REFMAC v.5.8	FSC _{work} & FSC _{test}
Bartesaghi & Matthies et al., 2014	β-galactosidase	3.2 Å	Chimera COOT	COOT	COOT	COOT MolProbity
Lu & Bai et al., 2014	Human γ-secretase	4.5 Å	COOT	COOT	REFMAC v.5.8	FSC _{work} & FSC _{test}

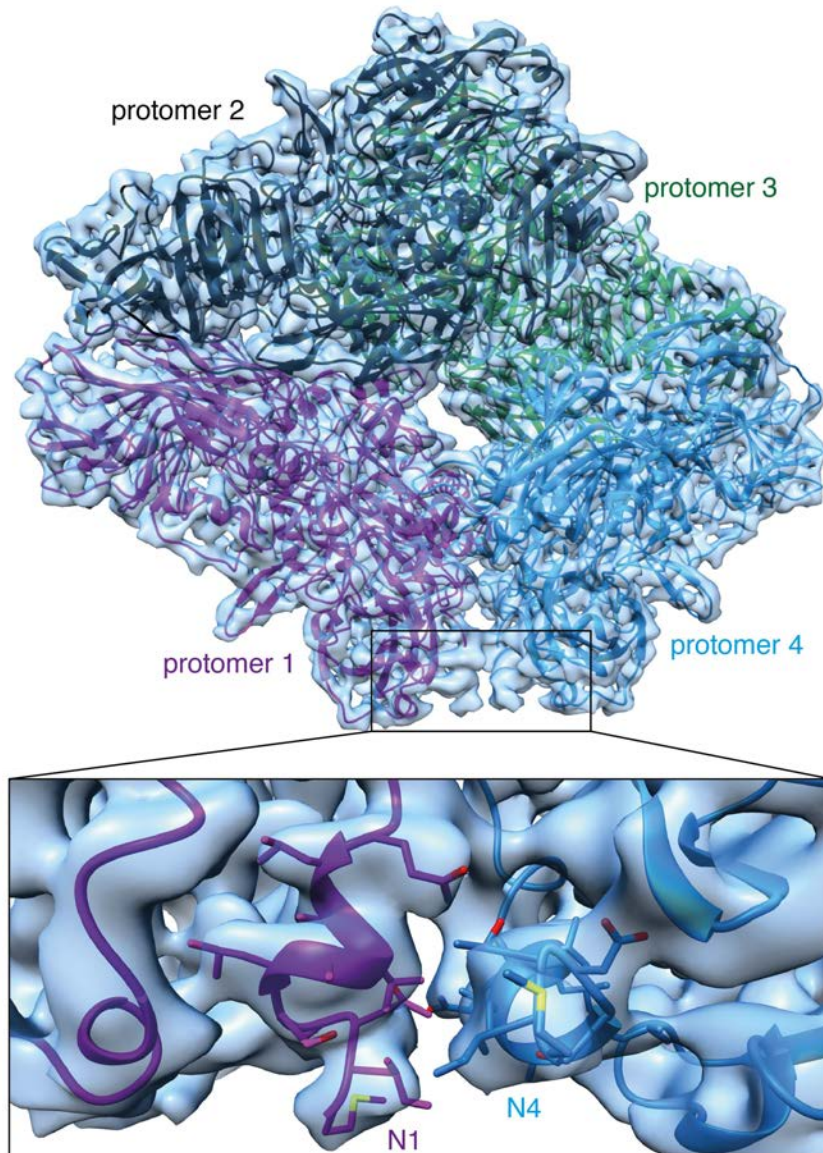


Rigid-body fitting of a single subunit of an X-ray structure using UCSF Chimera

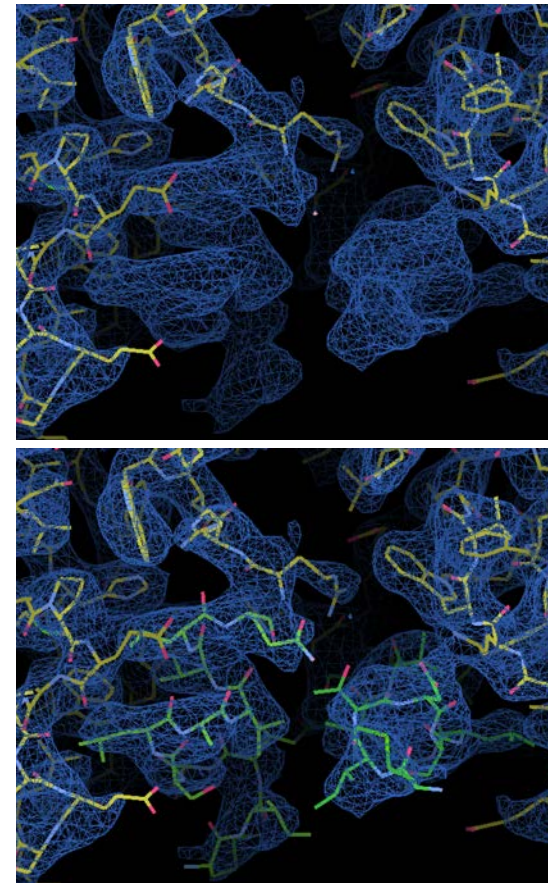


Flexible fitting and real space refinement using COOT

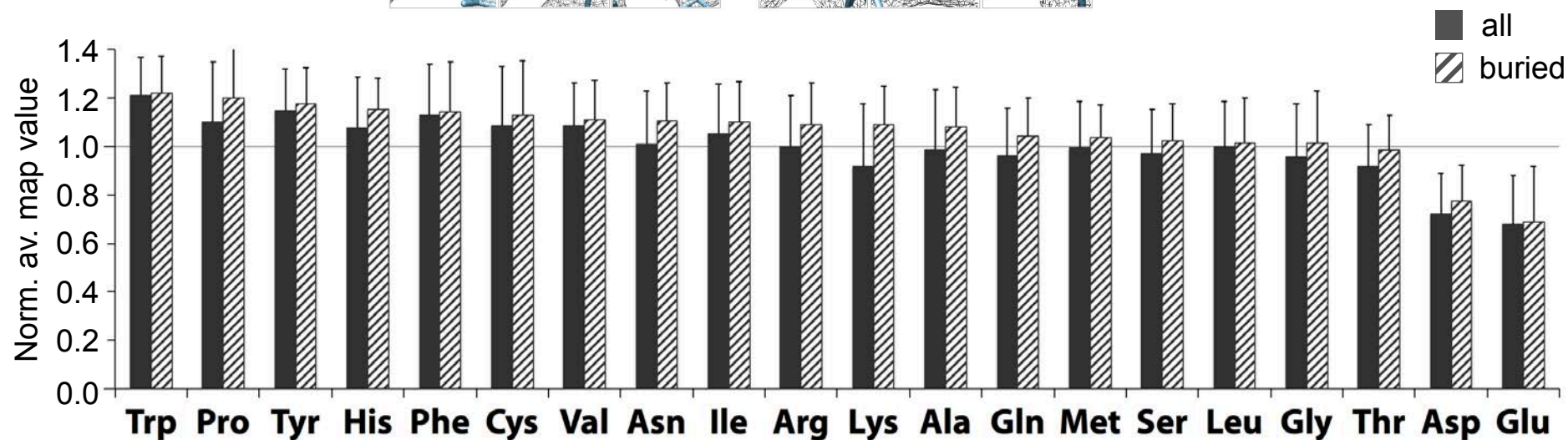
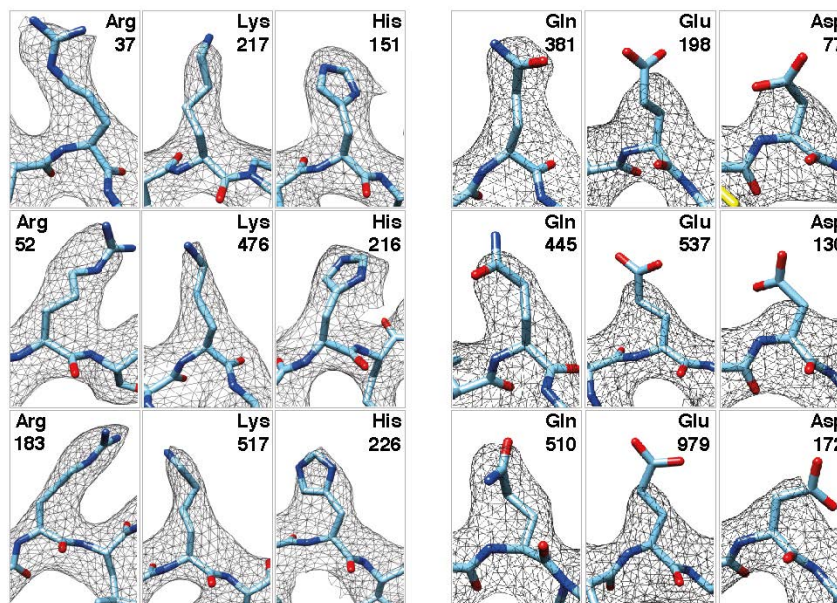




- N-terminal domain of unknown structure
 - areas with low correlation were deleted and rebuilt
- > addition of N- or C-terminal residues to the model one by one in COOT followed by refinement

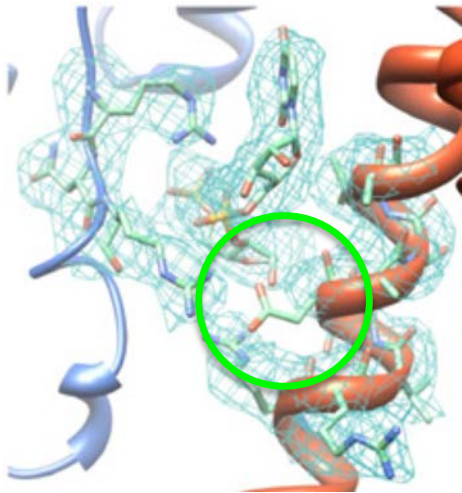


Bartesaghi & Matthies *et al.*, 2014

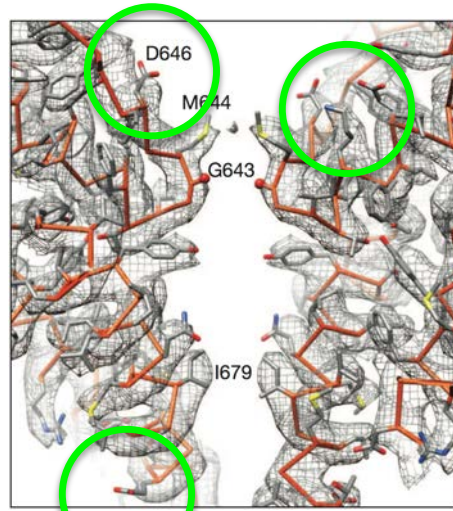


Bartesaghi & Matthies *et al.*, 2014

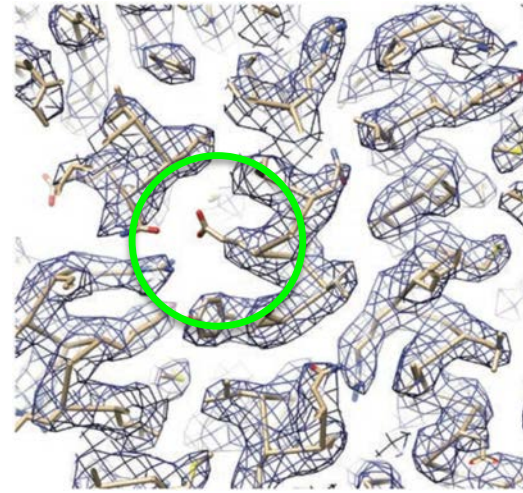
Lower density for glutamates and aspartates



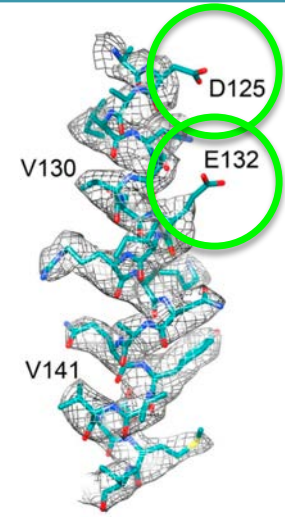
Ge & Zhou, 2011 (3.3 Å virus)



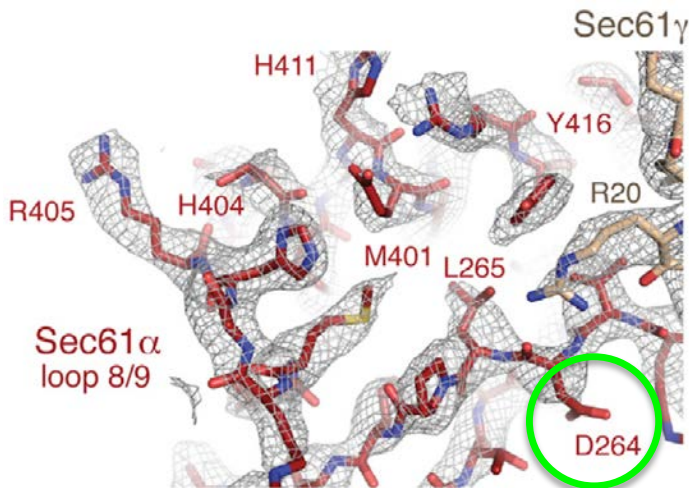
Liao & Cao *et al.*, 2013 (3.4 Å TRPV1)



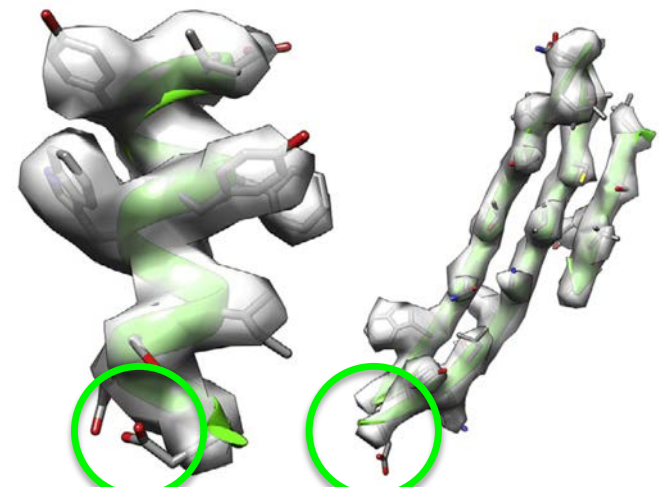
Li *et al.*, 2013 (3.3 Å 20S proteasome)



Allegretti *et al.*, 2014 (3.36 Å FRH)

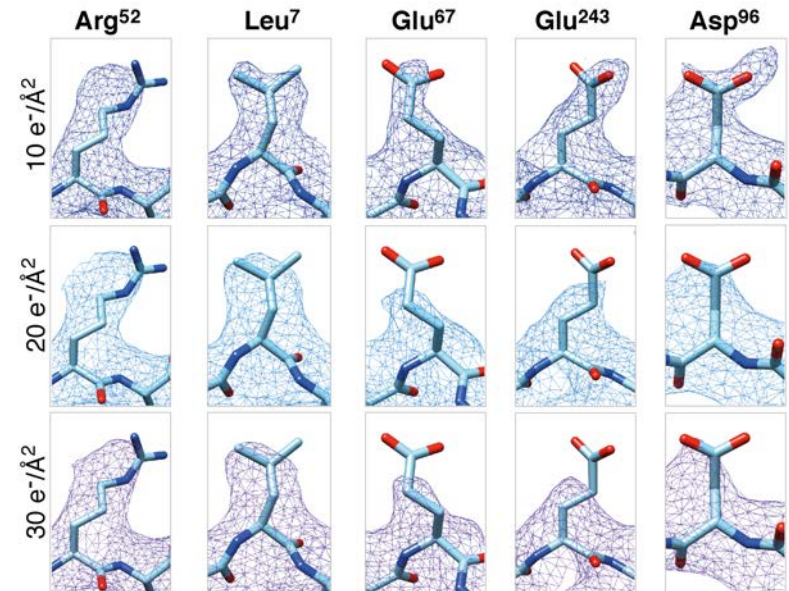
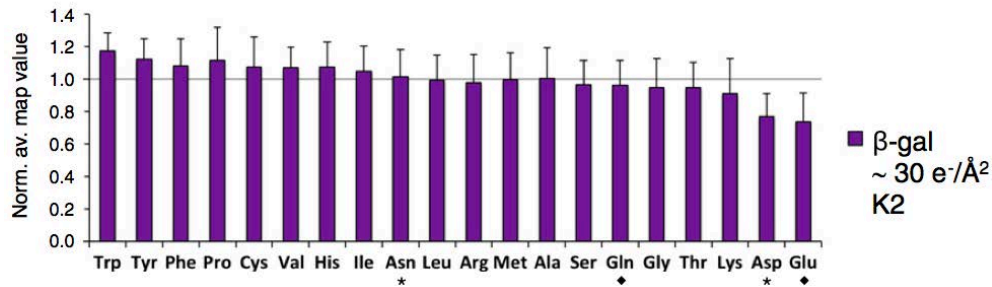
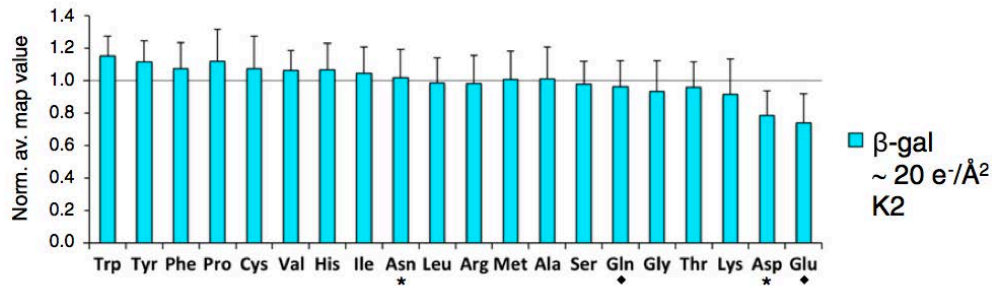
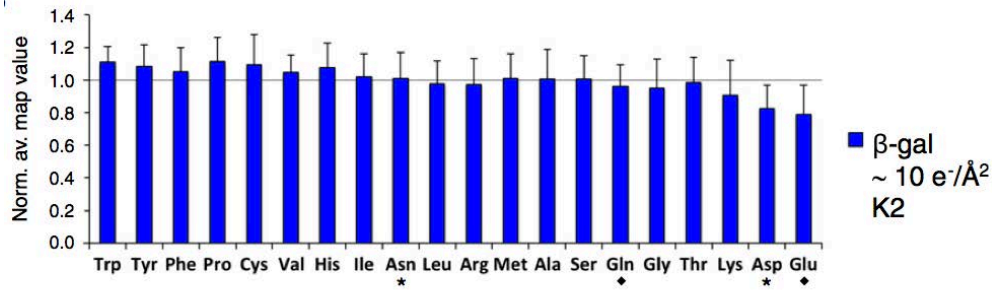
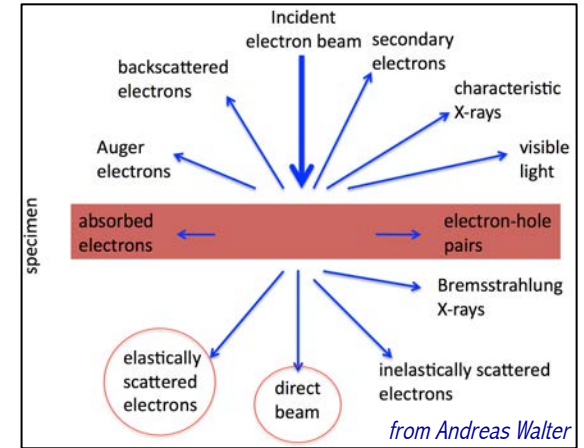


Vorhees *et al.*, 2014 (3.4 Å ribosome-Sec61)

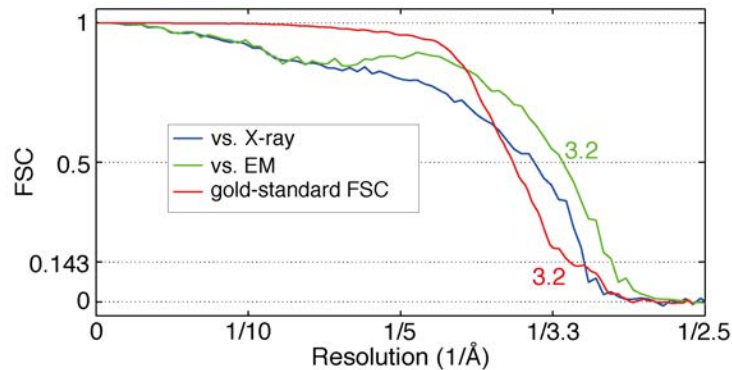


Campbell & Kearney *et al.*, 2014 (3.7 Å virus)

- Radiation damage or effect of electron scattering, which is influenced by local electric charges and ionization states, or both, or is it something completely different?
- How do we model the side chains if we do not have enough signal?
- How many maps should we make available?
- Which maps/data should be used for validation & b-factor calculations?



Bartesaghi & Matthies *et al.*, 2014



afternoon Matthies.. Welcome to Coot

Stereochemistry/Geometry:
 Peptide bonds
 Phi/Psi angles (Ramachandran)
 side chain rotamers
 clashes
 correct distances for hydrogen-
 bonds and salt bridges

MolProbity4 structure validation now provides many of its validation metrics through CCTBX, the open-source component of the Phenix crystallographic package. CCTBX allows for consistent validation results with Phenix, as well as added functionality, such as geometry regularization of NQH flips. Read more about this change [here](#).

We have updated Reduce to add hydrogens at a length more consistent with electron-cloud positions, and accordingly adjusted the Van der Waals radii in Probe to compensate for the change. This will affect comparison of results calculated with older versions of MolProbity, but generally results in lower clashscores. For analyses using nuclear-position hydrogens, you have the option of selecting nuclear x-H positions when adding hydrogens. Read more about this change [here](#).

Ramachandran scoring has also been updated to use new six-category distributions, derived from a larger Top8000 dataset of high quality PDB files.

Please don't hesitate to report any [bugs](#) you may encounter.

If for some reason you need to use MolProbity4 version 4.02, which is now a retired legacy version, please go to <http://rutile.biochem.duke.edu>.

Rigid-body fitting

- 3SOM
- ADPEM
- Attract-EM
- BCL::EM-Fit
- CoAn/CoFi
- EMatch
- Emfit
- EMLZerD
- GMFit
- IMP
- IQP
- MultiFit
- Situs
- UCSF Chimera
- X-PLOR
- ...

Flexible fitting & Refinement

- CNS-DEN
- COOT
- DireX
- Emap (CHARMM)
- EM-IMO
- Flex-EM / RIBFIND
- FRODA
- iMODFIT
- IMP
- MapSGLD (CHARMM)
- MDFF (NAMD)
- MDfit
- NMFF
- NORMA
- Phenix.refine
- REFMAC
- ROSETTA
- RSRef
- S-flexfit
- YUP.SCX
- XPLOR-NIH
- ...

Integrative Modeling

- IMP
- ROSETTA

Model building

- COOT
- EM-fold
- MODELLER /Mod-EM
- ROSETTA
- TASSER
- Assemble2
- Rcrane
- ...

Villa & Lasker, 2014; ...

Model validation

- ADP-EM
- COOT
- MolProbity
- ProQM
- Situs
- ...

Cross-validation methods

- (Amunts & Brown & Bai & Llacer *et al.*, 2014)
- (Wong & Bai & Brown *et al.*, 2014)
- (DiMaio *et al.*, 2013)
- (Shaikh *et al.*, 2003)
- (Falkner & Schröder, 2013)

- Homology modeling vs. *de novo* building:
When is the sequence identity too low?
 - *uncertainties in sequence alignments*
 - *validity of homology-modeling premise*
- Are automated *de novo* methods ready to replace manual approaches?
 - *sufficiently fast for large systems?*
 - *easy to use? (installation, documentation, usage)*
 - *specialized high-performance computing equipment?*
- What are the users' experiences?

Some software tools

- Homology modeling
 - MODELLER
 - TASSER
- *De novo* modeling
 - COOT
 - Rosetta
 - EM-fold (helices)
 - Assemble2 (RNA)
 - COOT/Rcrane (RNA)

FF used to alter atomic model to conform to the EM map

Challenge: maps contain errors and uncertainties difficult to quantify and are transferred into the final model;

FF algorithms can also introduce errors from limitations in sampling and scoring, particularly at mid-resolutions

- Manual vs. automated fitting, or both?
- Real-space vs. reciprocal-space refinement, or both?
- What are the advantages and disadvantages of
 - traditional methods used for X-ray data
 - methods based on MD/Monte-Carlo simulations
 - molecular-modeling methods
- Is the choice of method dependent on resolution?
- What are the users' experiences?

Some software tools

- Traditional X-ray tools
 - CNS
 - XPLOR
 - COOT
 - Phenix.refine
 - REFMAC
 - RSTref (CNS)
- Molecular modeling tools
 - Rosetta
 - DireX
 - Flex-EM (MODELLER)
 - IMP
 - EM-IMO
 - S-flexfit
- MD/MC Simulation-based methods
 - MDFF (NAMD)
 - EMAP (CHARMM)
 - MDfit
 - FRODA
- Normal-modes/elastic-network
 - iMODFIT
 - NMFF
 - NORMA
 - YUP.SCX

I. Stereochemistry & geometry

Peptide bonds, Phi/Psi angles (Ramachandran), side chain rotamers & clashes, correct distances for hydrogen-bonds and salt bridges, etc.

II. Structural model vs. data, i.e. cross-validation

Need for generally accepted cross-validation method, e.g. FSC_{work} vs. FSC_{test} , as well as a local descriptor of confidence in coordinate assignments (such as B-factors in X-ray crystallography)

III. Independent experimental validation

Crosslinking, Cys-accessibility measurements, EPR/FRET, mutants, ...

- What are the users' experience and opinions?

Some software tools

- ADP-EM
- COOT
- MolProbity
- ProQM (membrane proteins)
- Situs

Cross-validation

- FSC_{work} vs. FSC_{test} (Amunts & Brown & Bai & Llacer *et al.*, 2014; Wong & Bai & Brown *et al.*, 2014; Fernandez *et al.*, 2014)
- Splitting data in half and only use one half for model building and refinement (DiMaio *et al.*, 2013)
- Omitting data from the high spatial frequency range (Falkner & Schröder, 2013)
- Exclusion of resolution shells in reciprocal space (Shaikh *et al.*, 2003)