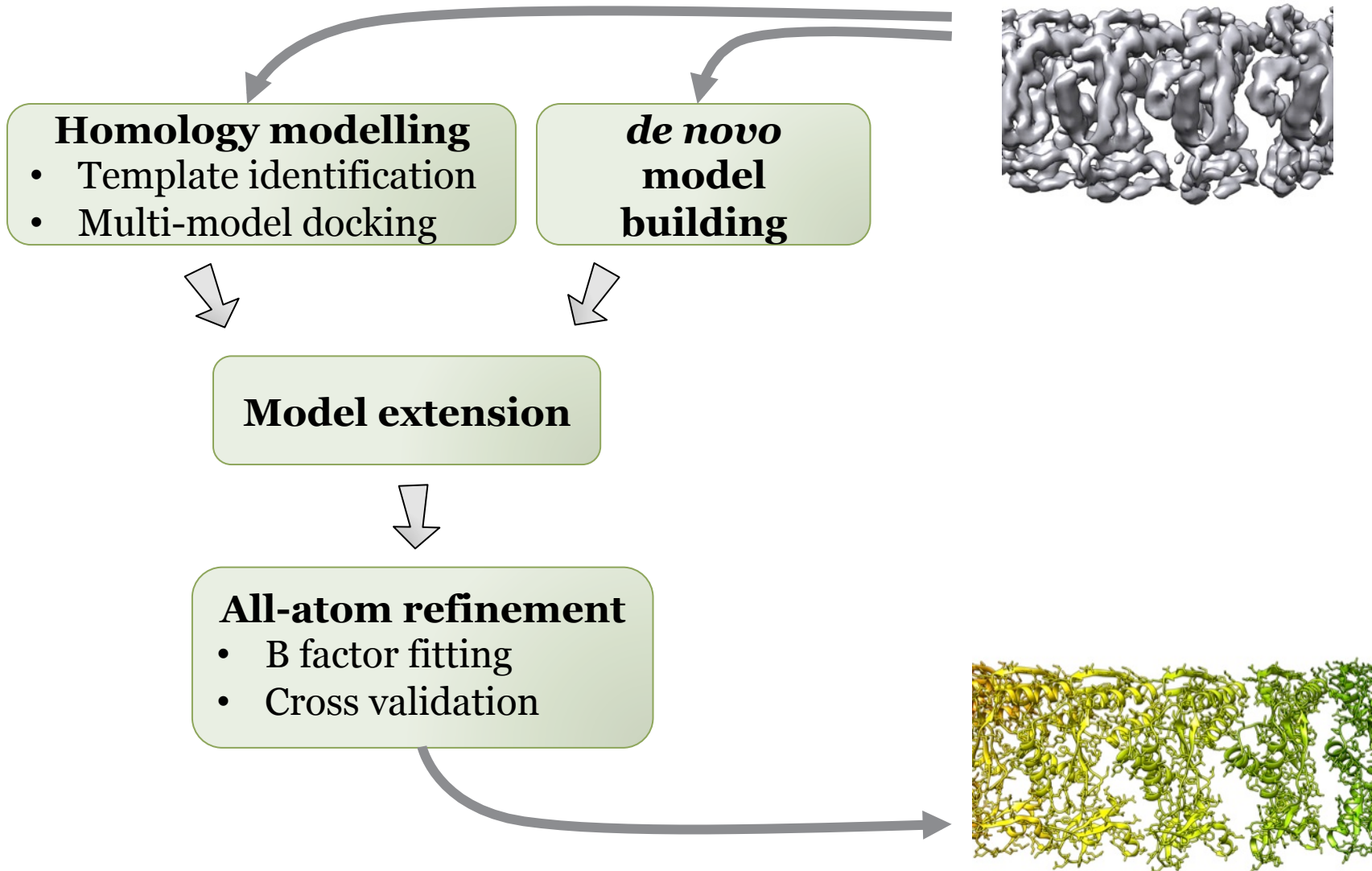


Toward automated structure determination from near-atomic resolution data

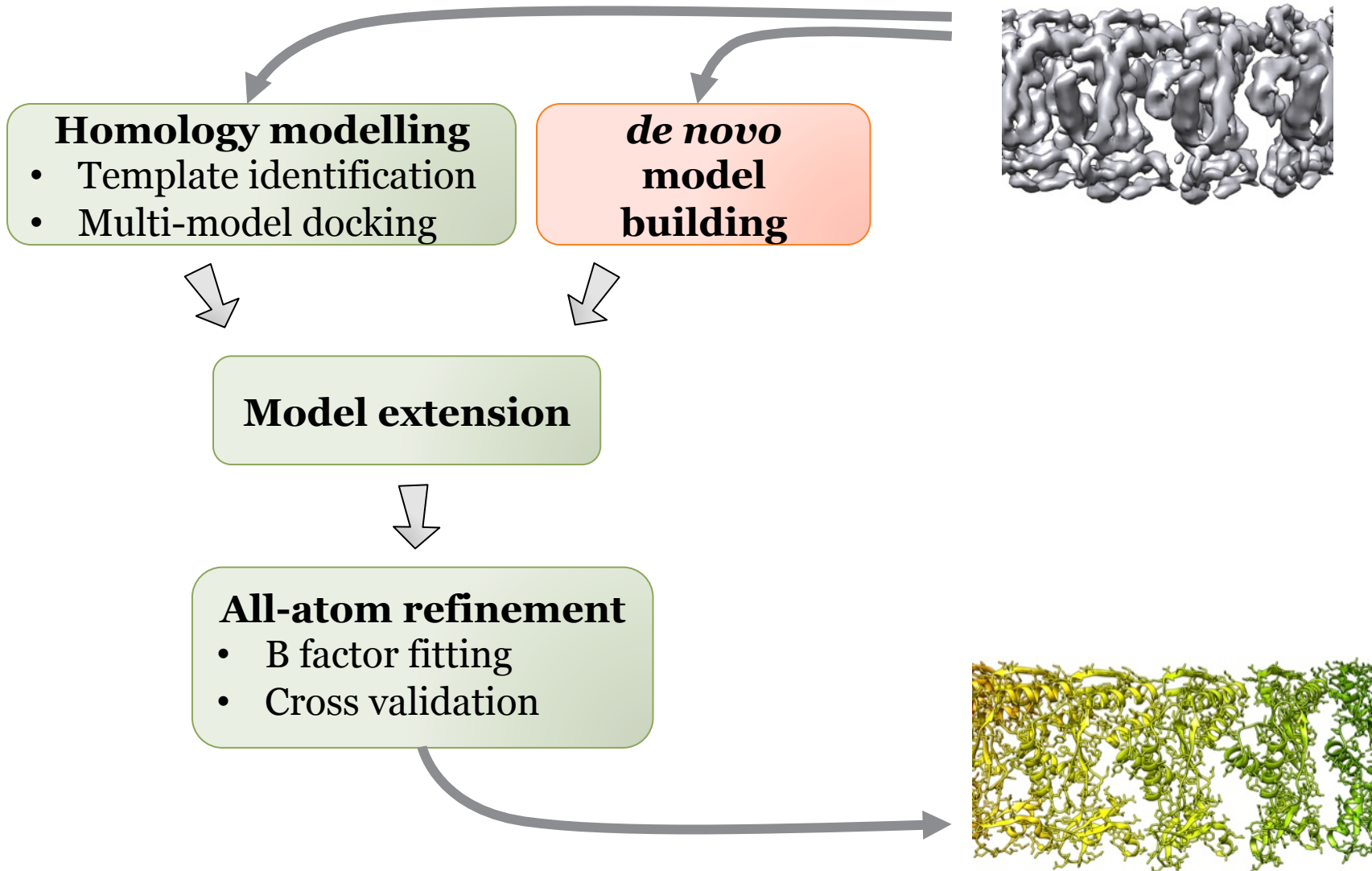
Frank DiMaio
University of Washington
Institute for Protein Design

November 2014

Accurate structure determination with RosettaEM



Accurate structure determination with RosettaEM



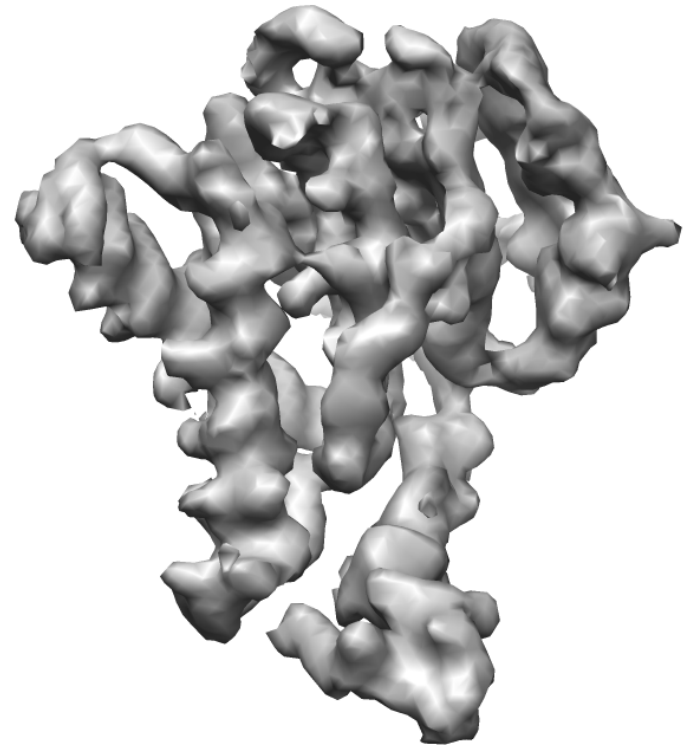
Lack of sidechain detail makes identifying sequence difficult

Crystallographic “autotracing”:

**Backbone
tracing**



**Sequence
registration**

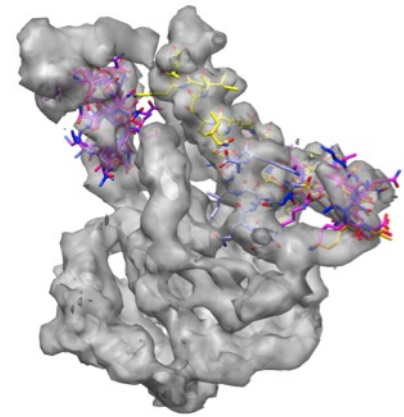
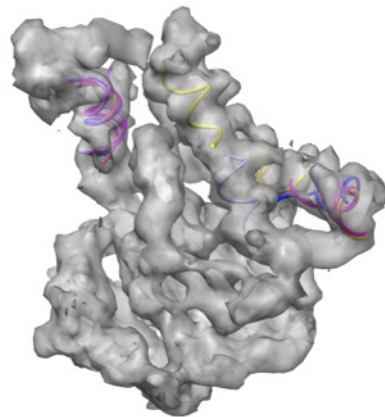


4.8Å reconstruction
20S proteasome
(courtesy Yifan Cheng & Xueming Li)

Searching density for local backbone conformations

Local sequence restricts local structure

...CVK**VTKPL**VARAKL...



6-dimensional
search

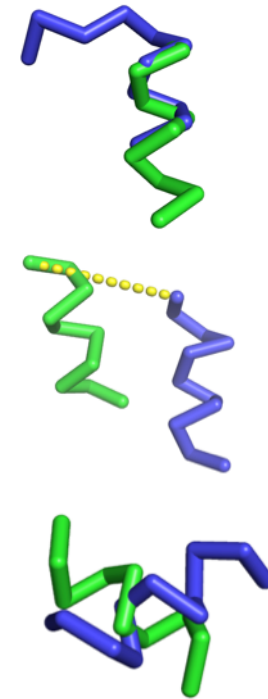
sidechain building
& refinement

Ray Wang (in review)

Selecting a maximally consistent set of fragments

Idea: The correct placements must all be consistent

- adjacent fragments must assign the same residue to the same location
- residues close in sequence must be close in space
- no two residues can occupy the same space

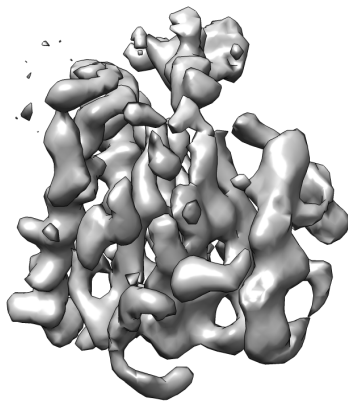


$score(\mathbf{F}) =$

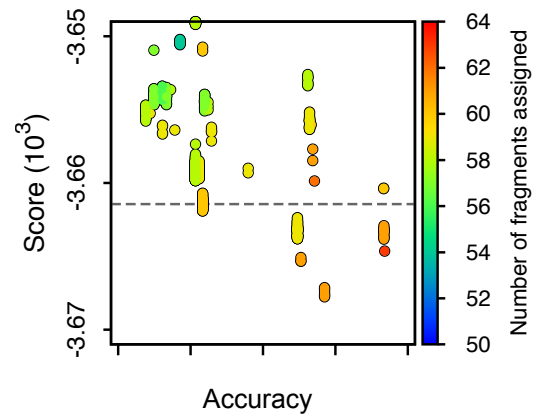
$$\sum_{f_i \in \mathbf{F}} sC_{dens}(f_i) + \sum_{f_i, f_j \in \mathbf{F}} sC_{overlap}(f_i, f_j) + \sum_{f_i, f_j \in \mathbf{F}} sC_{close}(f_i, f_j) + \sum_{f_i, f_j \in \mathbf{F}} sC_{clash}(f_i, f_j)$$

Monte Carlo sampling correctly identifies sequence

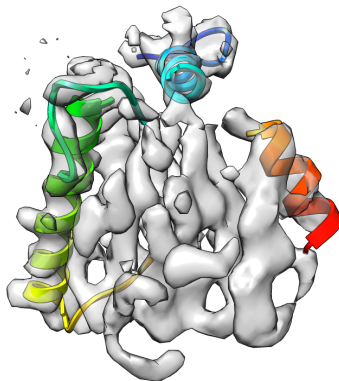
Density Map



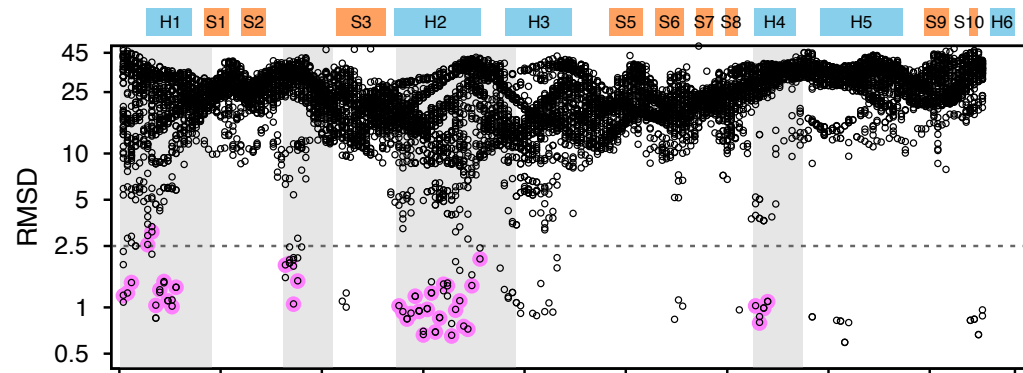
Monte Carlo Sampling



Partial Model



Fragment Placement



Multiple rounds of sampling completes model

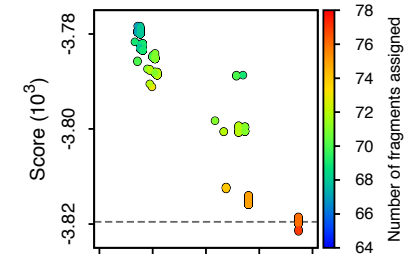
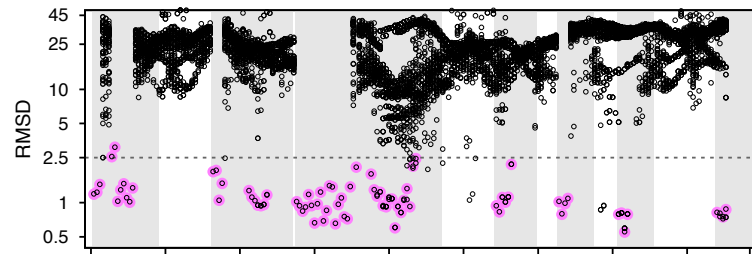
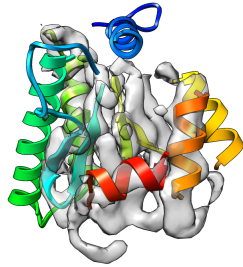
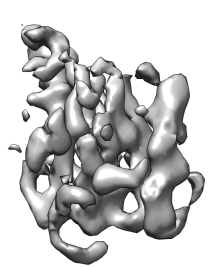
Density Map

Partial Model

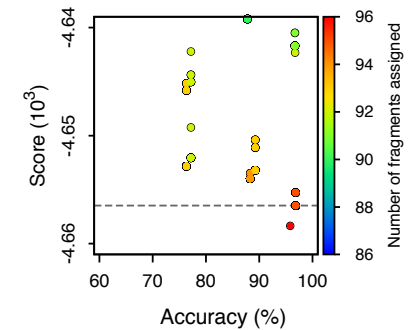
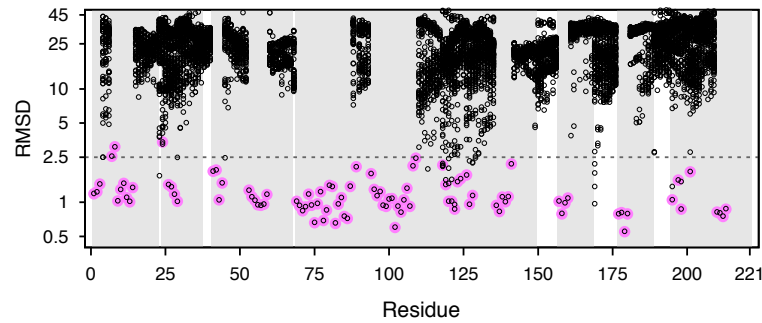
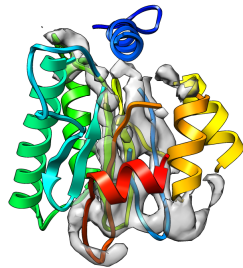
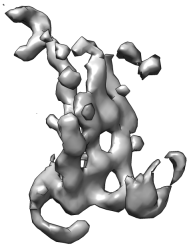
Fragment Placement

Monte Carlo

Round 2

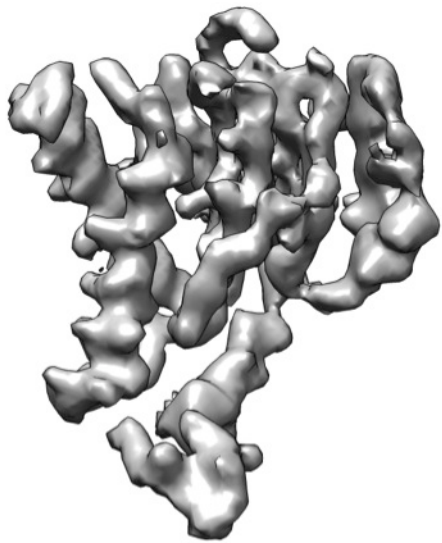


Round 3

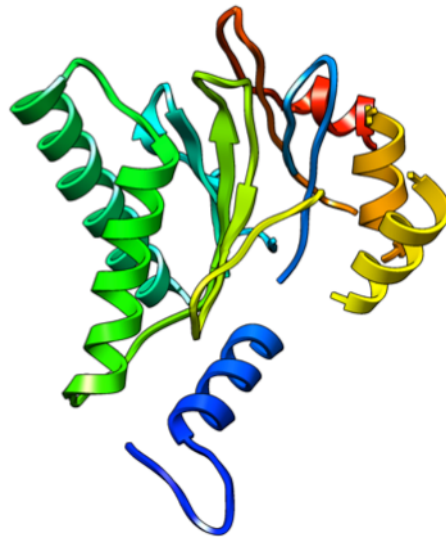


20S proteasome α -subunit at 4.8 Å

Density

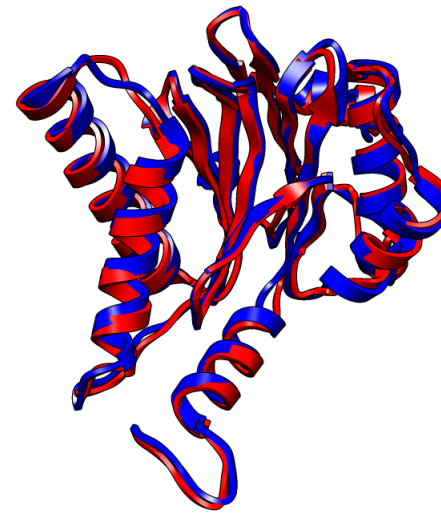


Final Partial Model

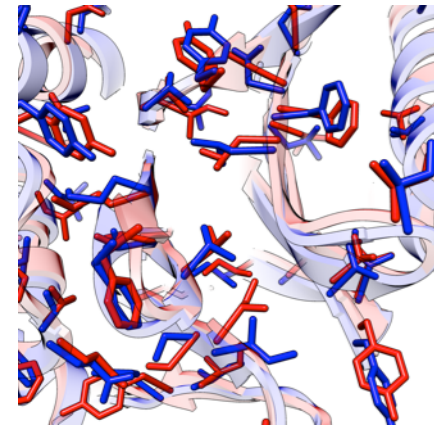


1.28 Å
196/213 rlds

Overlay of the
fulllength model (red)
to the native (blue)



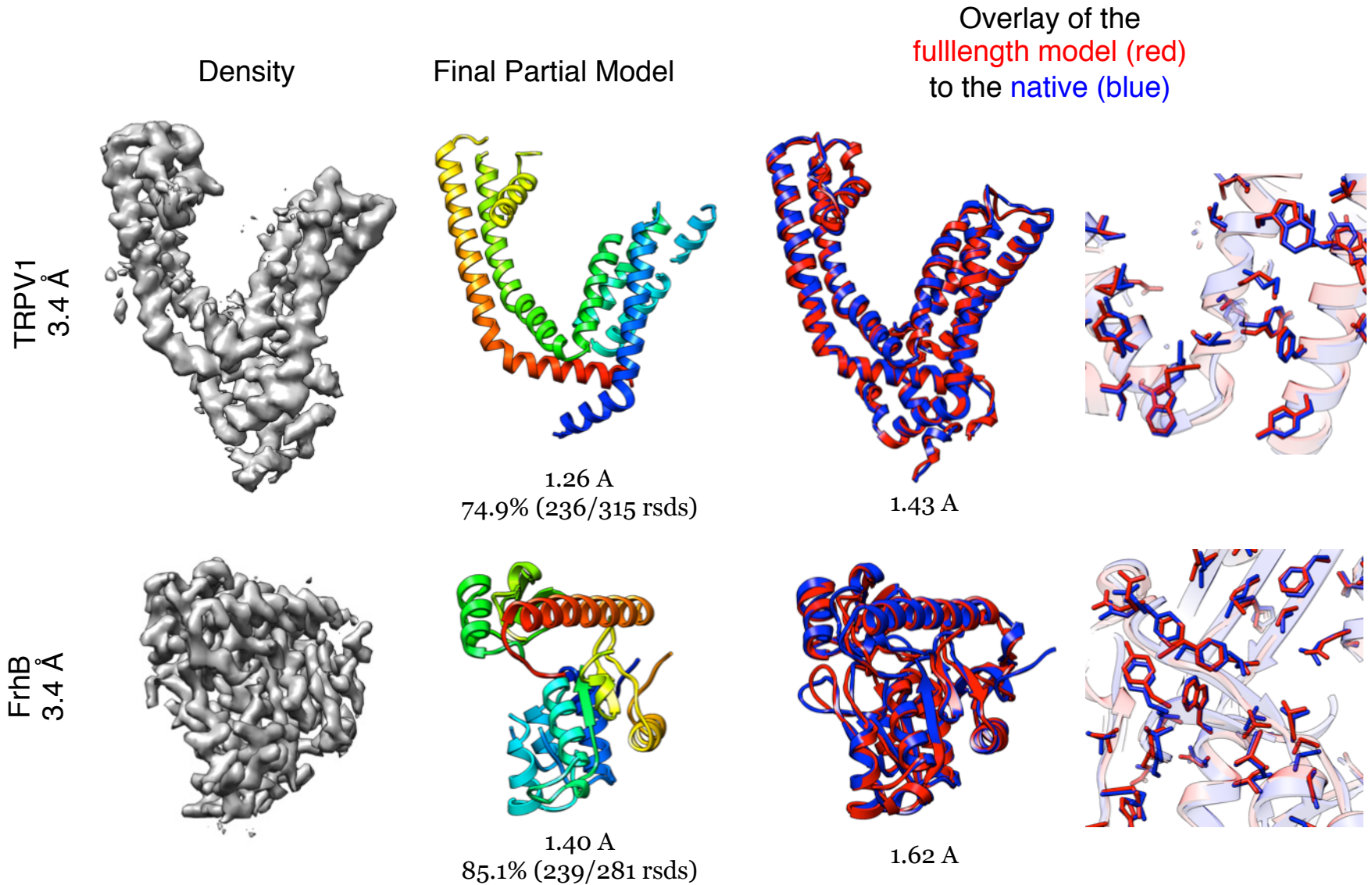
1.19 Å



Automatic structure determination is accurate in 6 of 9 cases

| Target | PDB ID (chain) | EMDB ID | Reported resolution (Å) | Length (aa) | Partial model C α RMSd [Å] (%) | C α RMSd [Å] |
|---------------|----------------|---------|-------------------------|-------------|---------------------------------------|---------------------|
| TMV | 3j06 (A) | 5185 | 3.3 | 155 | 1.3 (81) | 1.7 |
| TRPV1 | 3j5q (A) | 5778 | 3.4 | 310 | 1.1 (76) | 1.4 |
| FrhA | 4ci0 (A) | 2513 | 3.4 | 385 | 2.3 (91) | 1.3 |
| FrhB | 4ci0 (C) | 2513 | 3.4 | 280 | 1.4 (85) | 1.7 |
| FrhG | 4ci0 (B) | 2513 | 3.4 | 228 | 1.6 (73) | 2.2 |
| BPP1 | 3j4u (A) | 5764 | 3.5 | 327 | 17.2 (42) | - |
| VP6 | 1qhd (A) | 1461 | 3.8 | 397 | 1.6 (52) | - |
| 20S- α | 1pma (A) | TBD | 4.8 | 221 | 1.3 (88) | 1.2 |
| STIV | 3j31 (A) | 5584 | 3.9 | 344 | 21.9 (26) | - |

Automatic structure determination is accurate in 6 of 9 cases

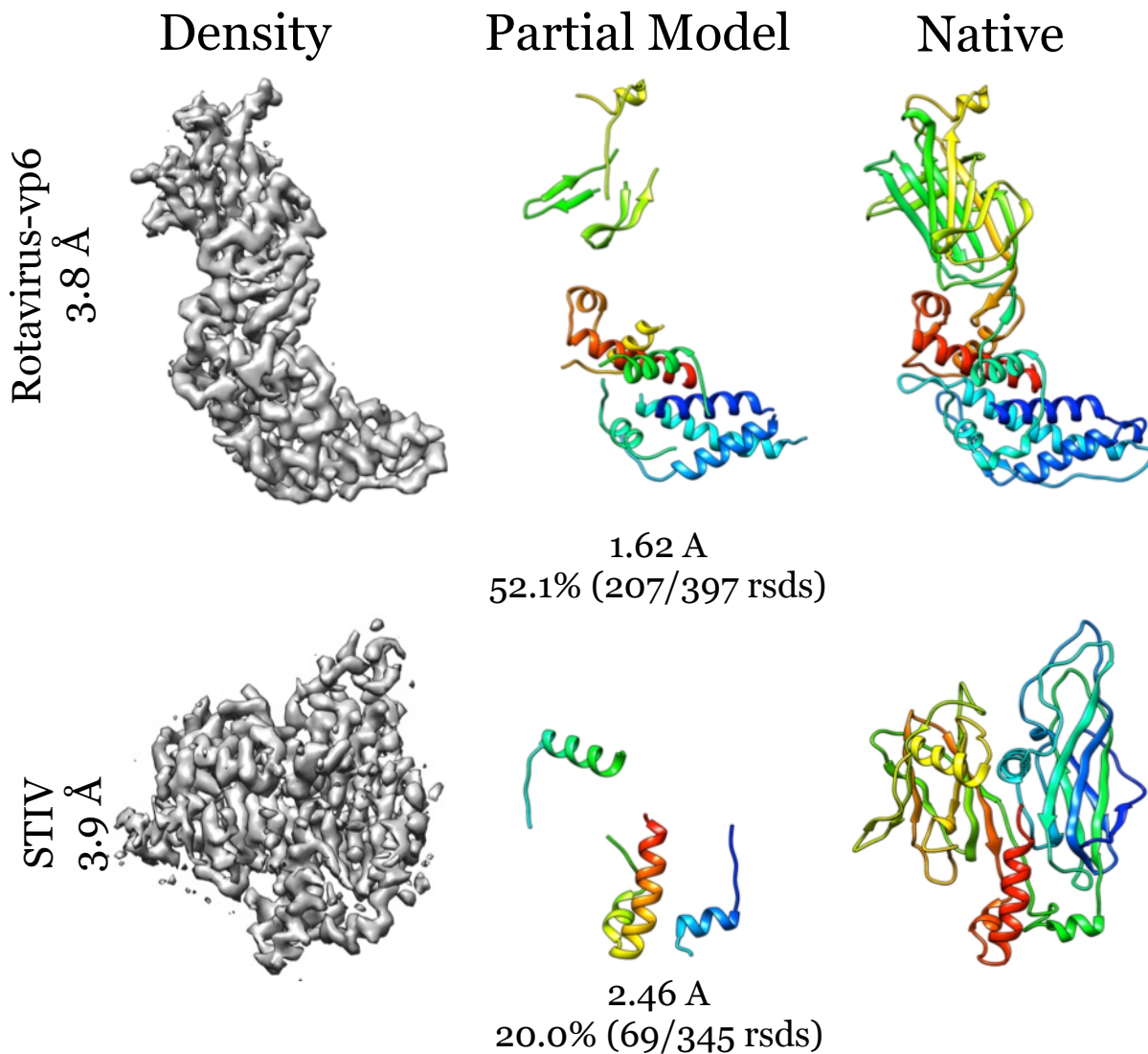


Crystallographic chain tracing is generally unable to register sequence

Using Buccaneer:

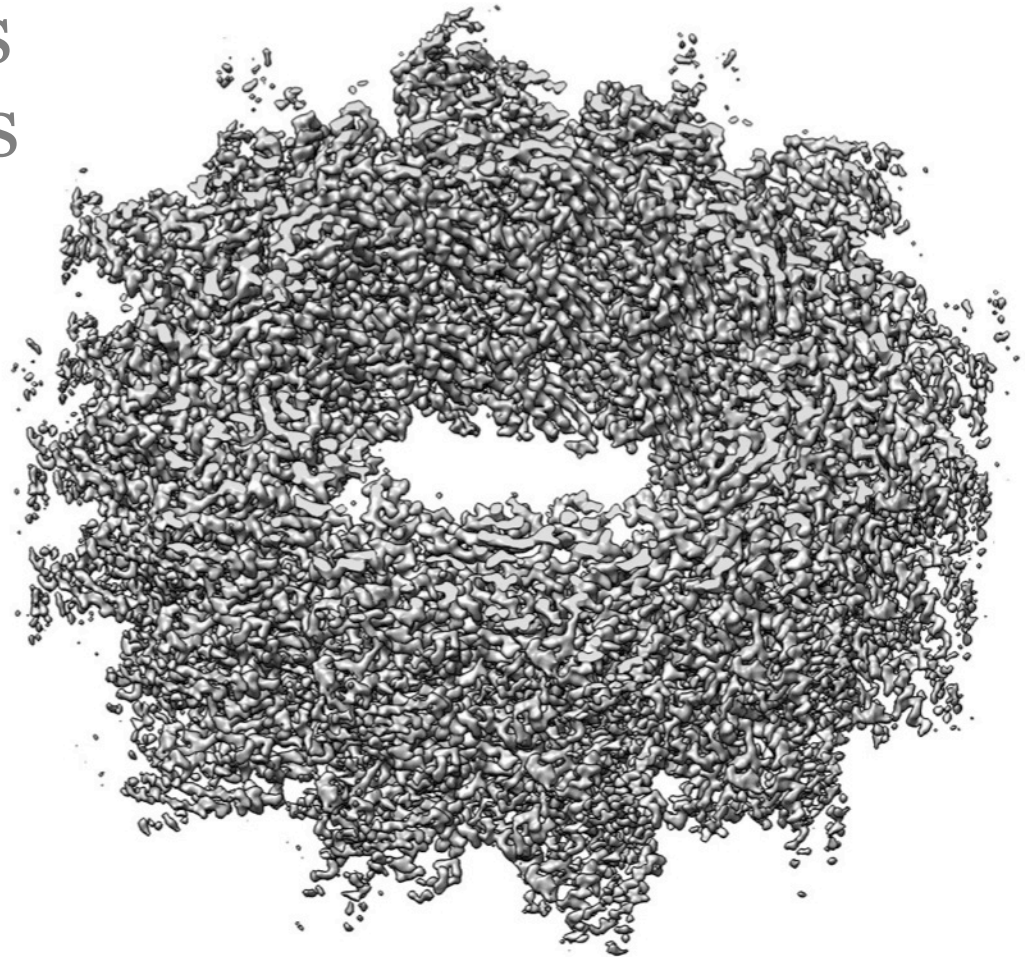
| Target | PDB ID (chain) | Length (aa) | C α atom placed | Sequence registered | Correctly registered |
|---------------|----------------|-------------|------------------------|---------------------|----------------------|
| TMV | 3j06 (A) | 155 | 145 | 56 | 0 |
| TRPV1 | 3j5q (A) | 315 | 257 | 190 | 0 |
| FrhA | 4ci0 (A) | 386 | 382 | 367 | 185 (48%) |
| FrhB | 4ci0 (C) | 281 | 192 | 186 | 126 (45%) |
| FrhG | 4ci0 (B) | 228 | 242 | 190 | 63 (27%) |
| BPP1 | 3j4u (A) | 327 | 339 | 162 | 0 |
| VP6 | 1qhd (A) | 397 | 405 | 155 | 0 |
| 20S- α | 1pma (A) | 221 | 224 | 135 | 7 (3%) |
| STIV | 3j31 (A) | 345 | 553 | 259 | 0 |

Failures are primarily in sheets



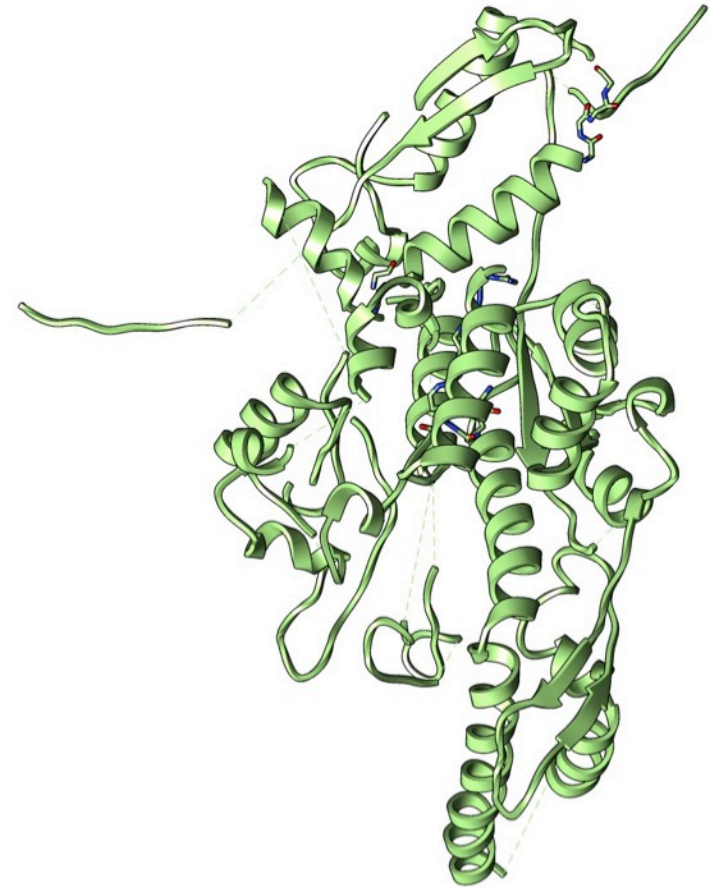
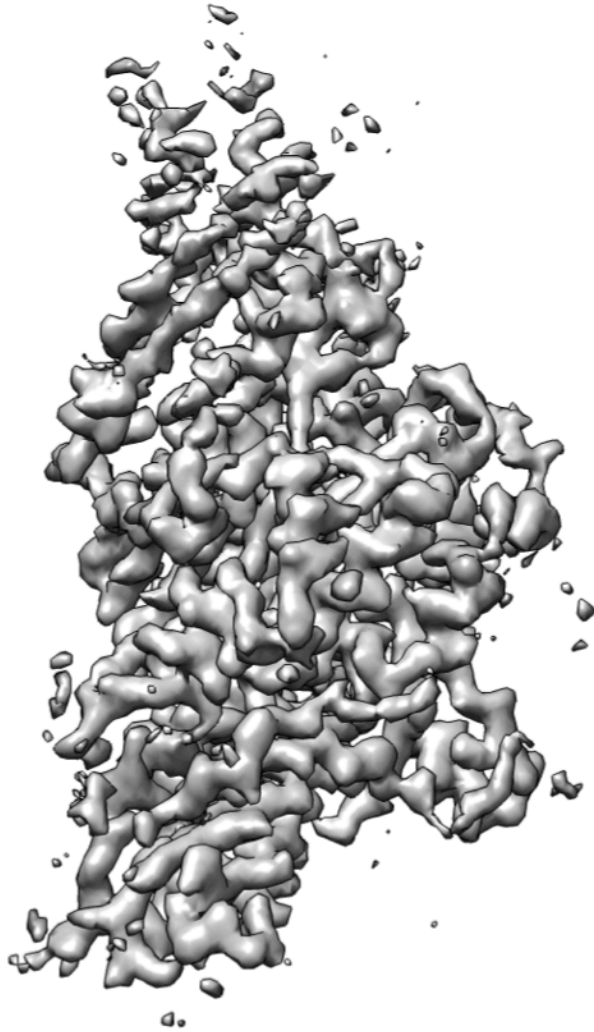
VipAB structure determination

VipA: 168 residues
VipB: 492 residues



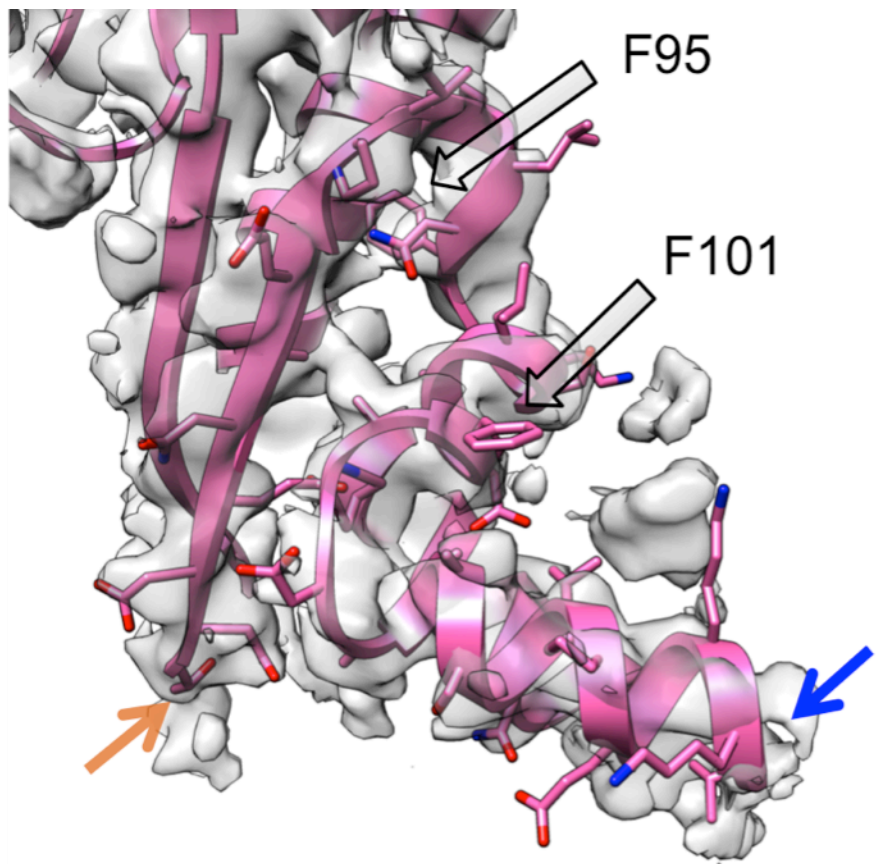
with Misha Kudryashev, Marek Basler, Ed Egelman (*in review*)

VipAB structure determination

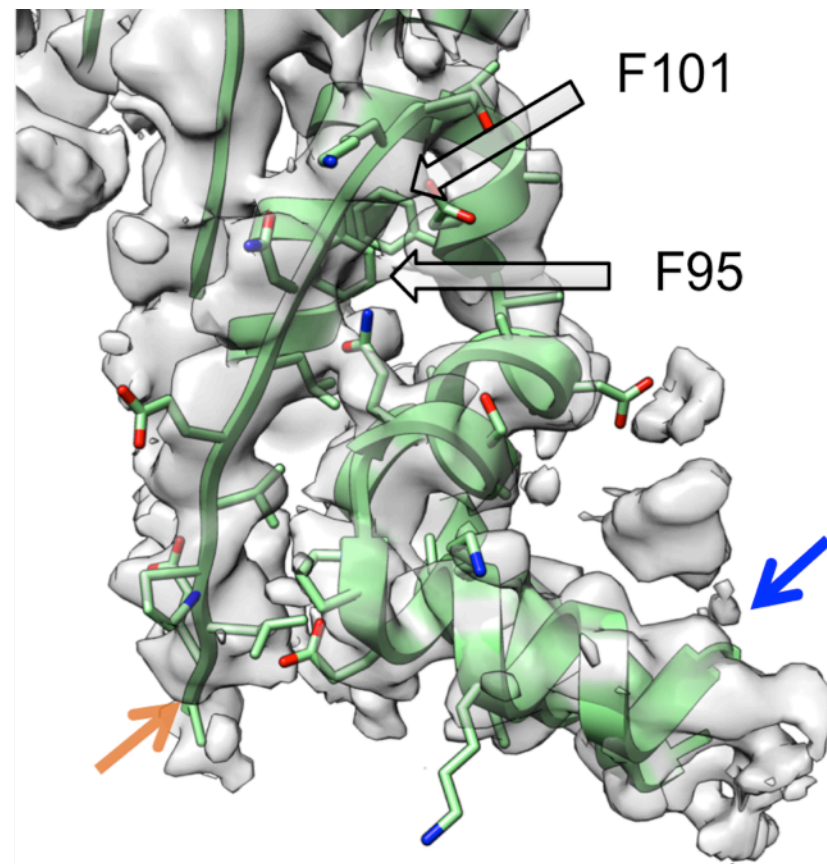


446/660 residues

Our method corrects errors from the manually traced model

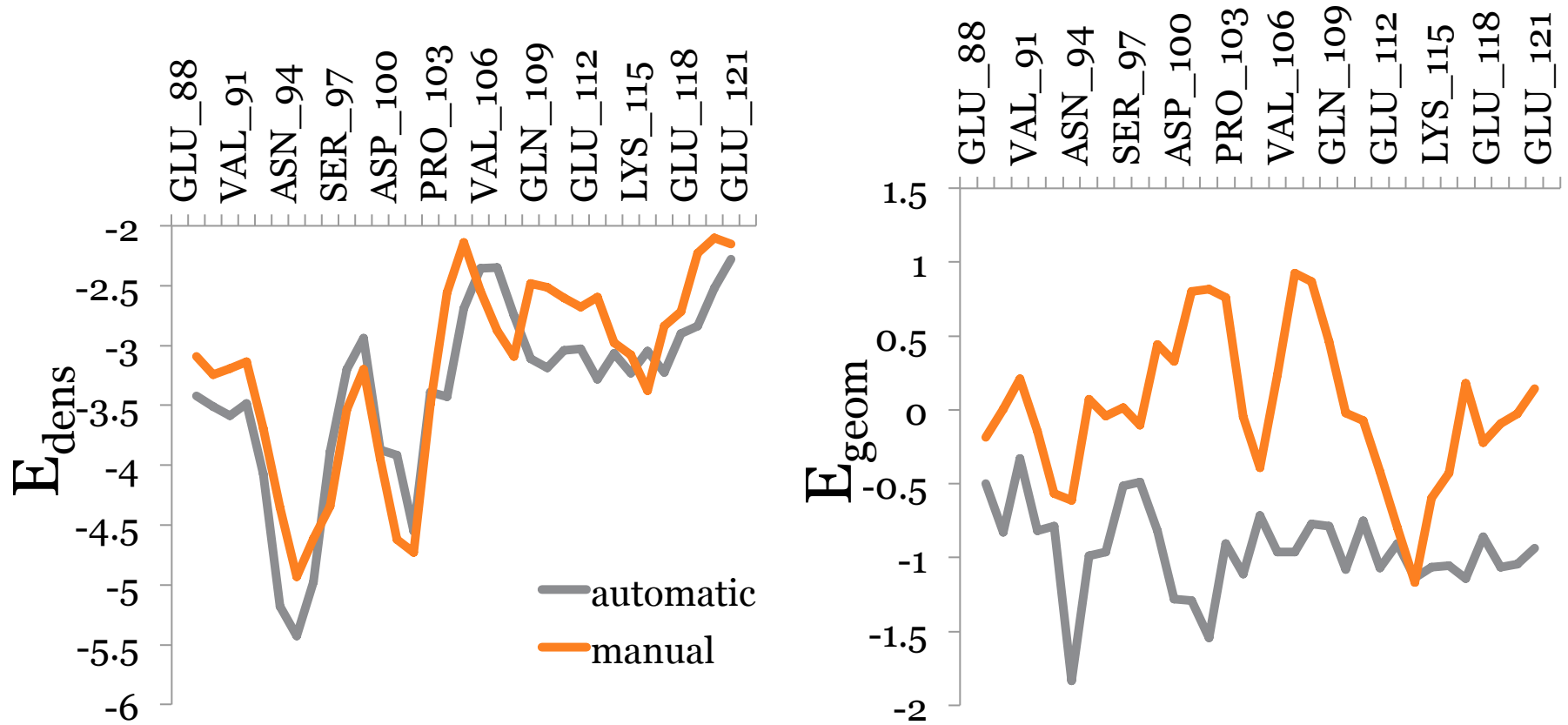


manual model

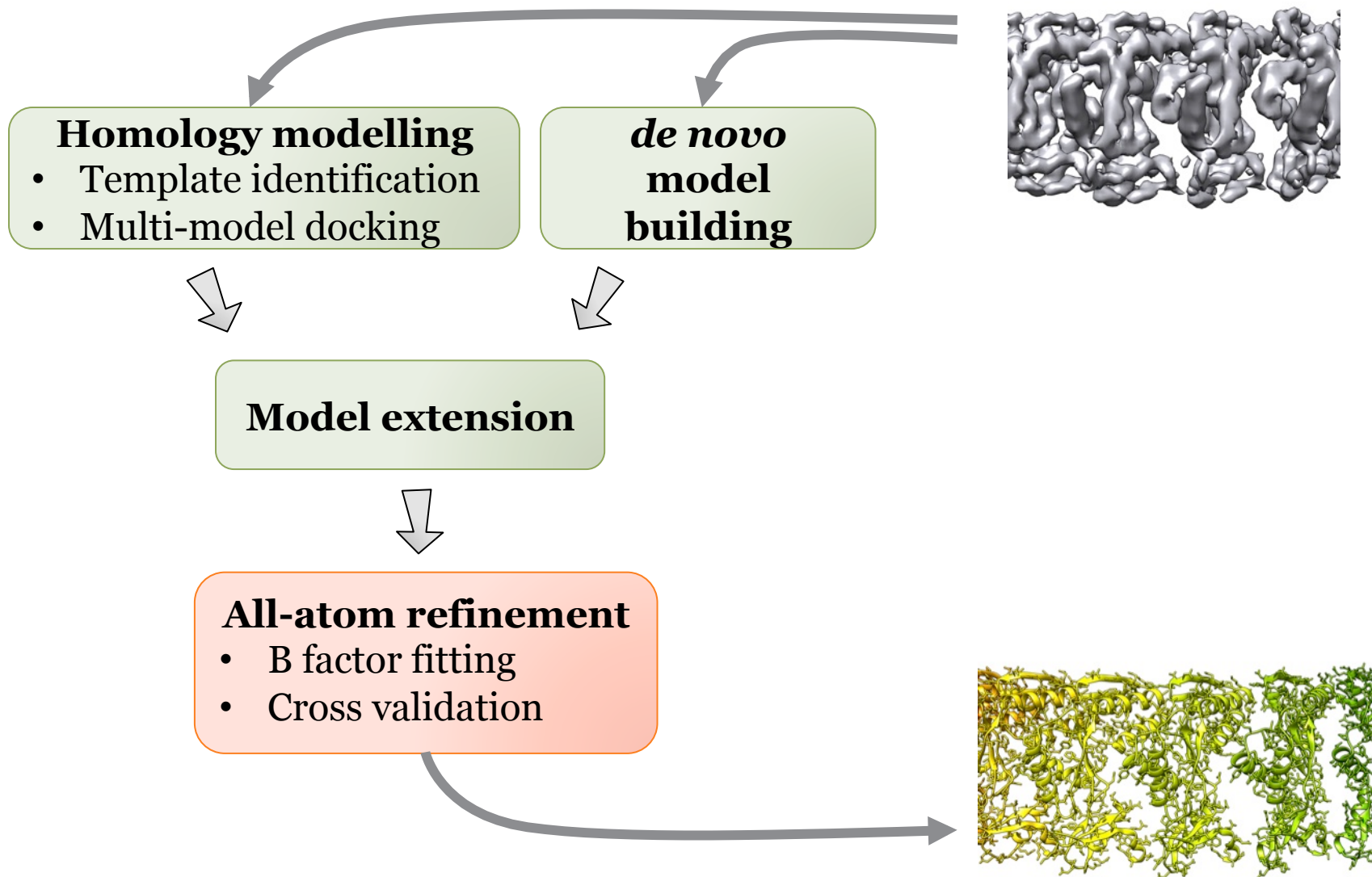


Automated model

Our method corrects errors from the manually traced model

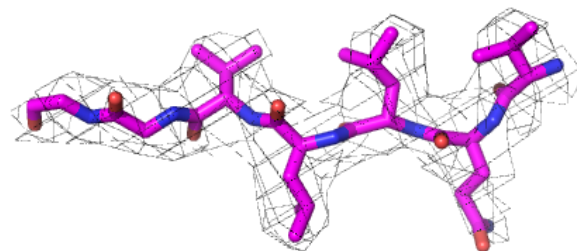
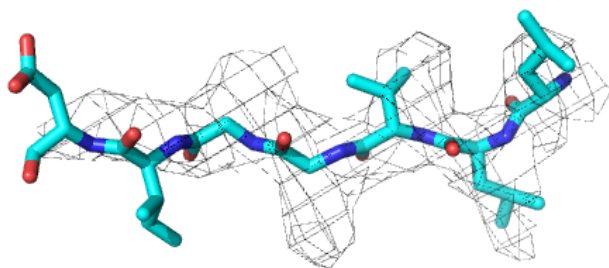


Accurate structure determination with RosettaEM



Refinement against EM density

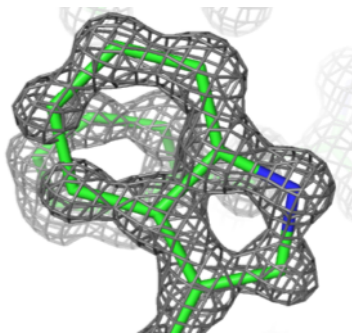
- Refinement
 - identify (and correct) errors in the initial model
 - improve fit to data
 - improve model geometry



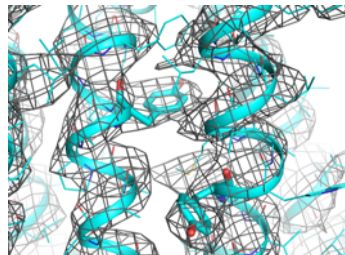
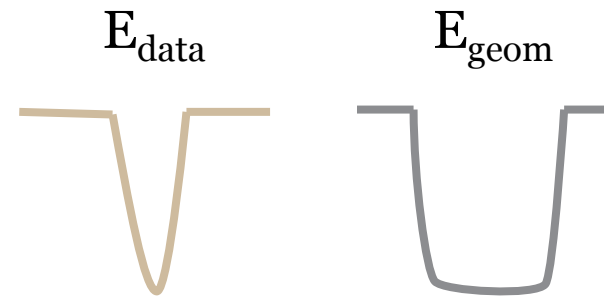
Refinement at low resolution requires a better geometry potential

Refinement: find atom positions optimizing:

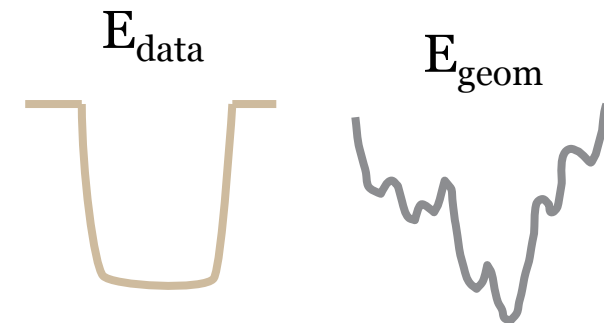
$$E = E_{geom} + w \cdot E_{data}$$



High-resolution



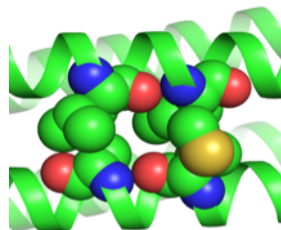
Low-resolution



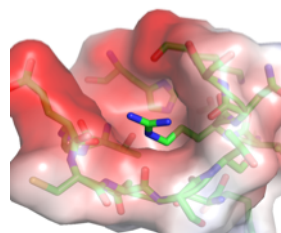
Rosetta forcefield disambiguates low-resolution solutions

Information from known structures reduces conformational space

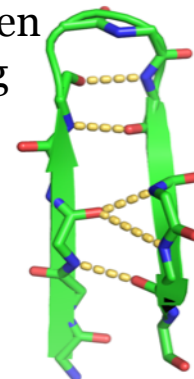
Core packing



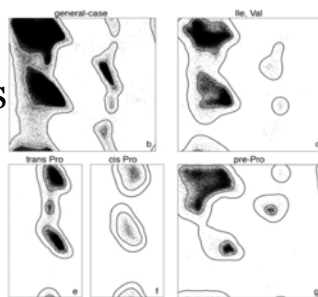
Electrostatics



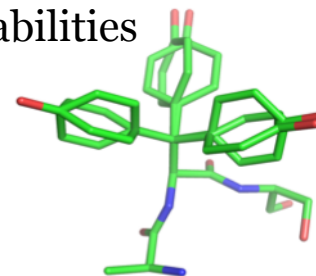
Hydrogen bonding



Torsional probabilities

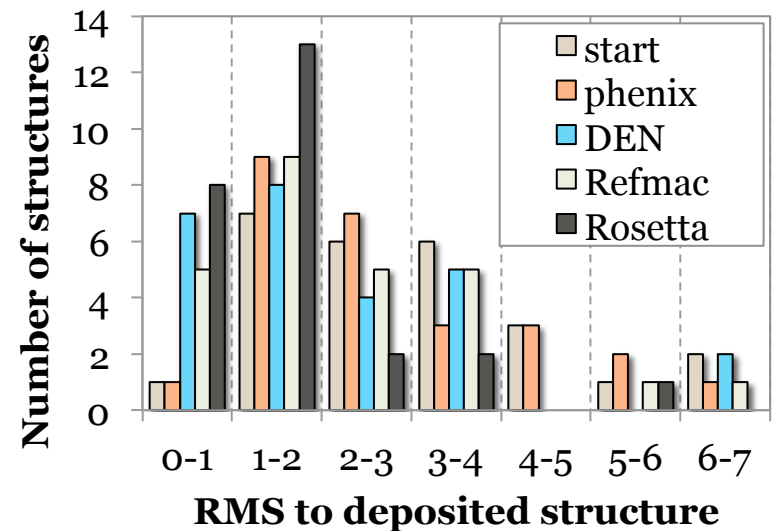
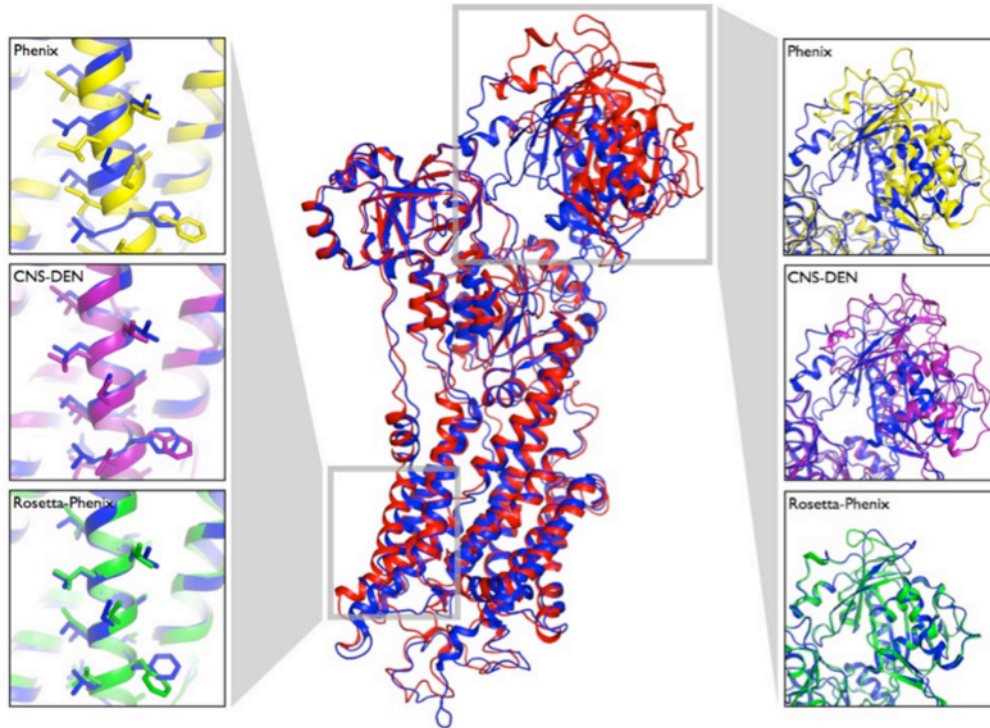


Rotamer probabilities



+ tools for improved optimization
(discrete sidechain optimization,
torsion and Cartesian space minimization, dynamics)

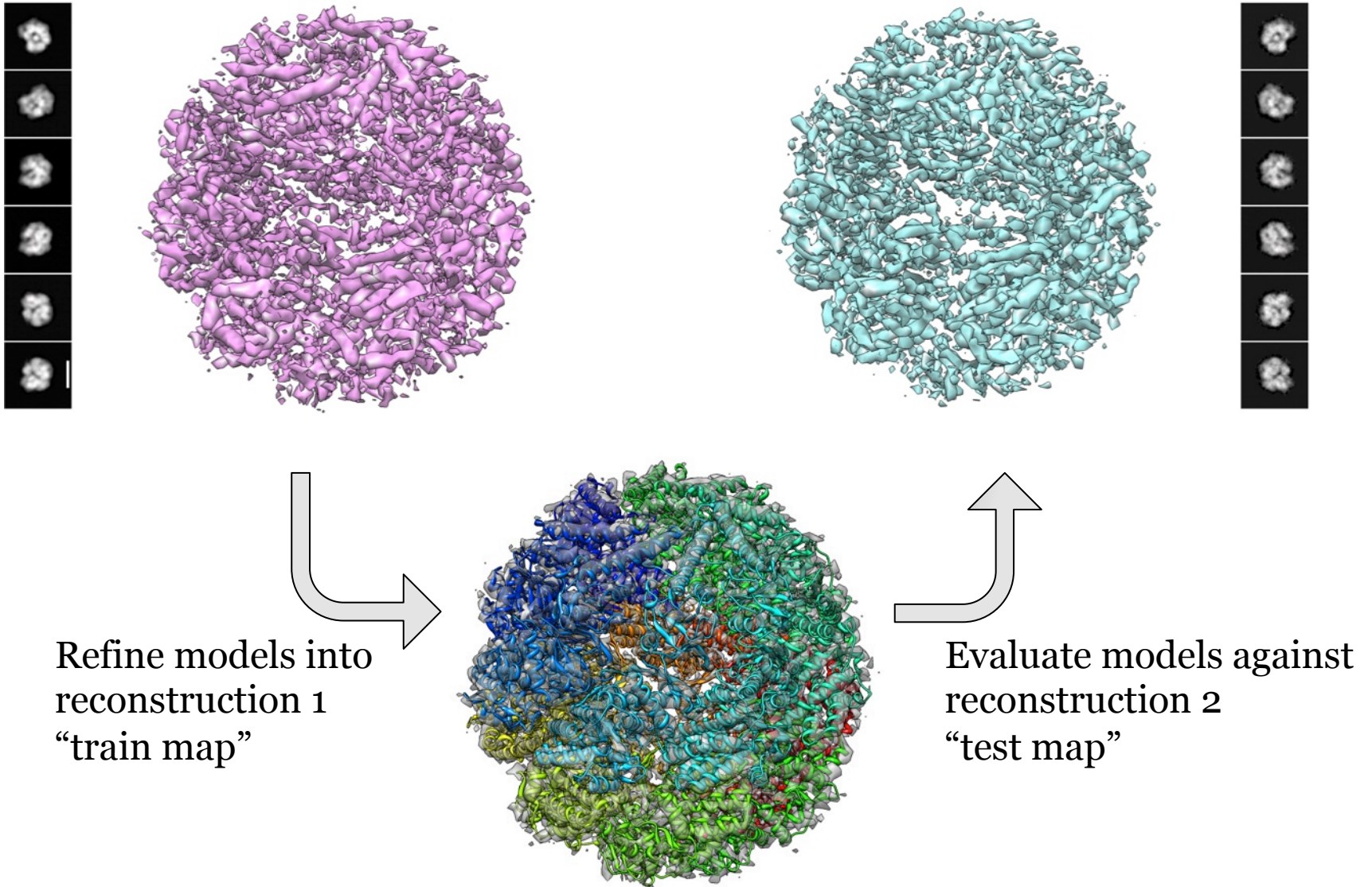
Our approach improves refinement against low-resolution crystallographic data



Key components for refinement against cryoEM

- Model validation
 - Independent map agreement over high-resolution shells
- Variations in local resolution
 - Atomic B factors describing how spread the density is around each atom
- Small radius of convergence
 - Discrete backbone optimization in refinement

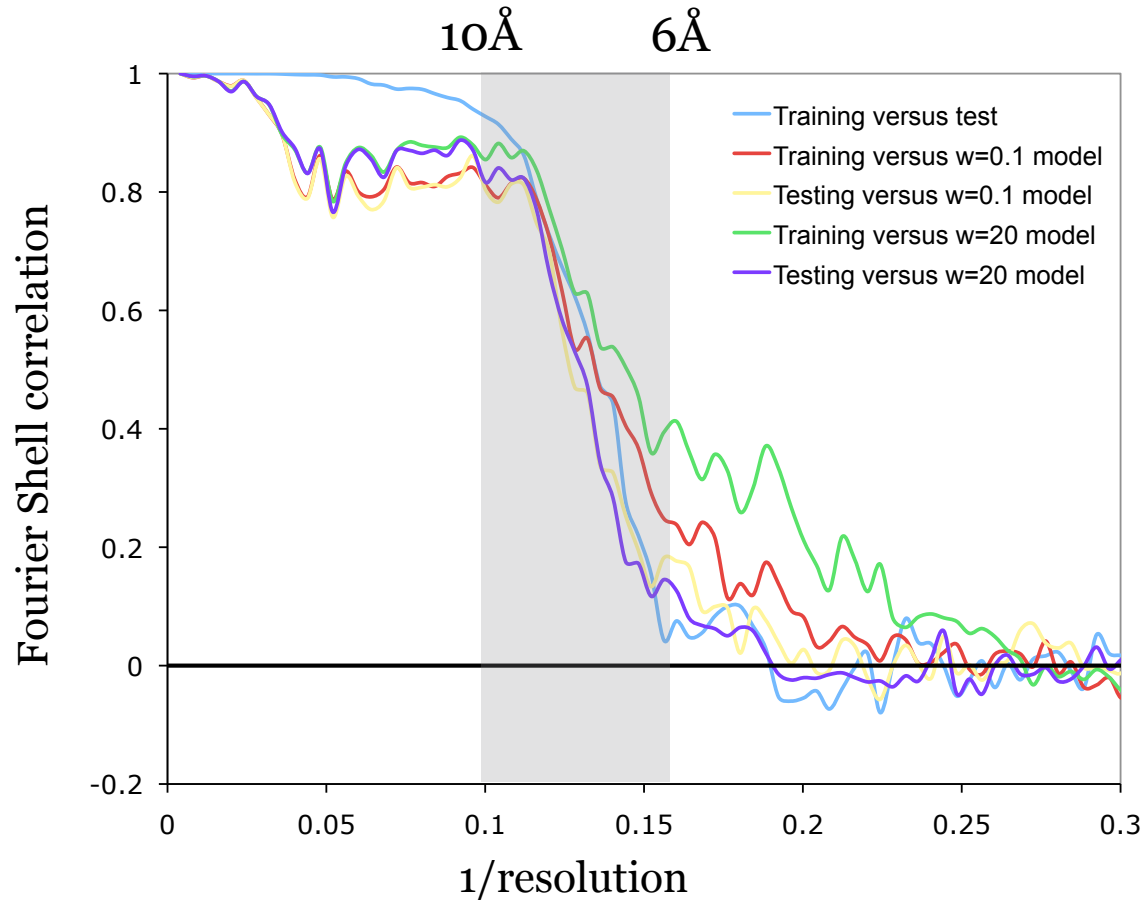
Independent validation



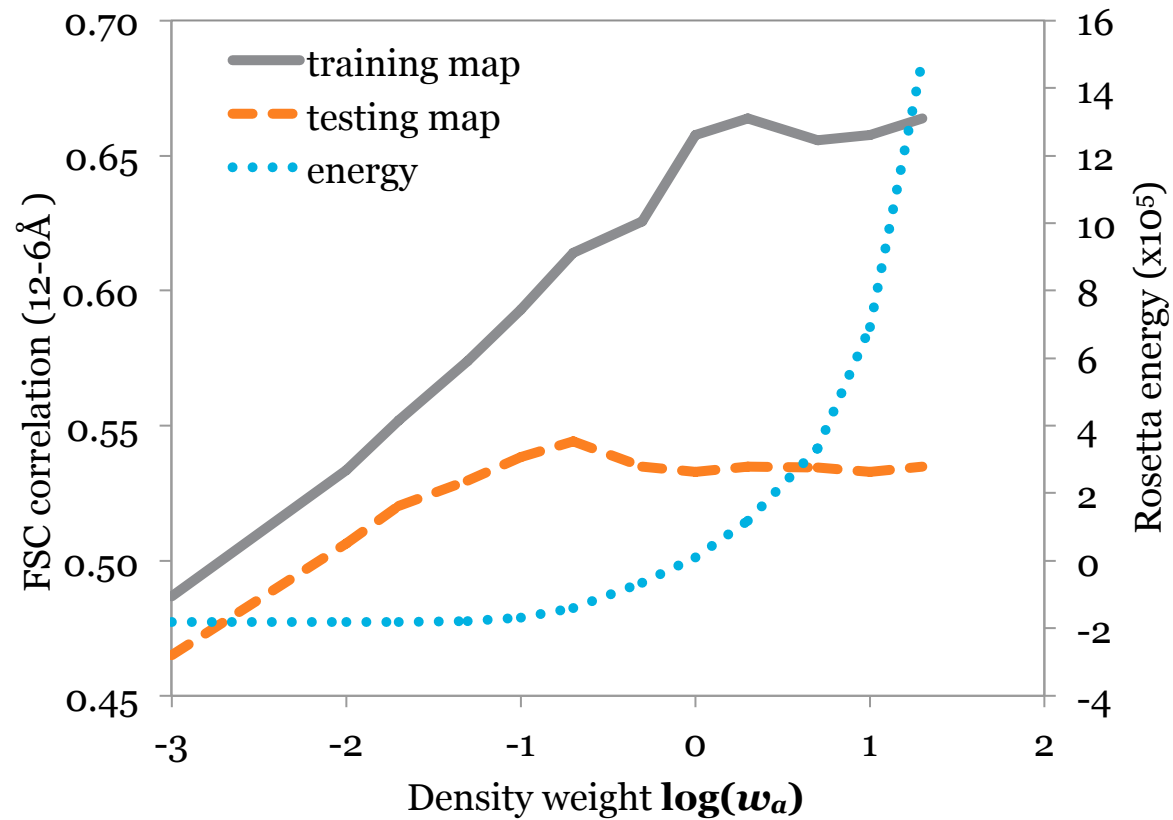
Refine models into
reconstruction 1
“train map”

Evaluate models against
reconstruction 2
“test map”

Independent validation

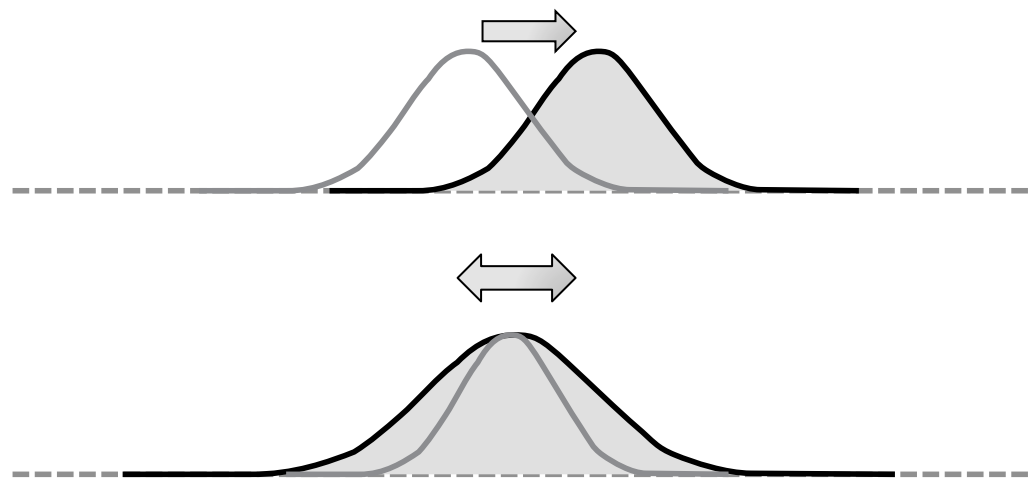


Independent validation



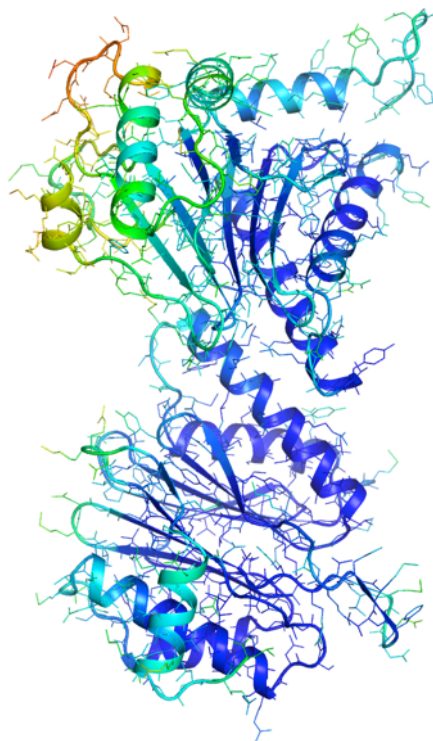
Fitting atomic B factors

- In addition to refining atomic coords, refine per-atom B factors (in real space)



- Alternate coordinate refinement and B factor refinement
- Constraint function keeps B factors of nearby atoms close

Model B's have good agreement with crystallographic Bs

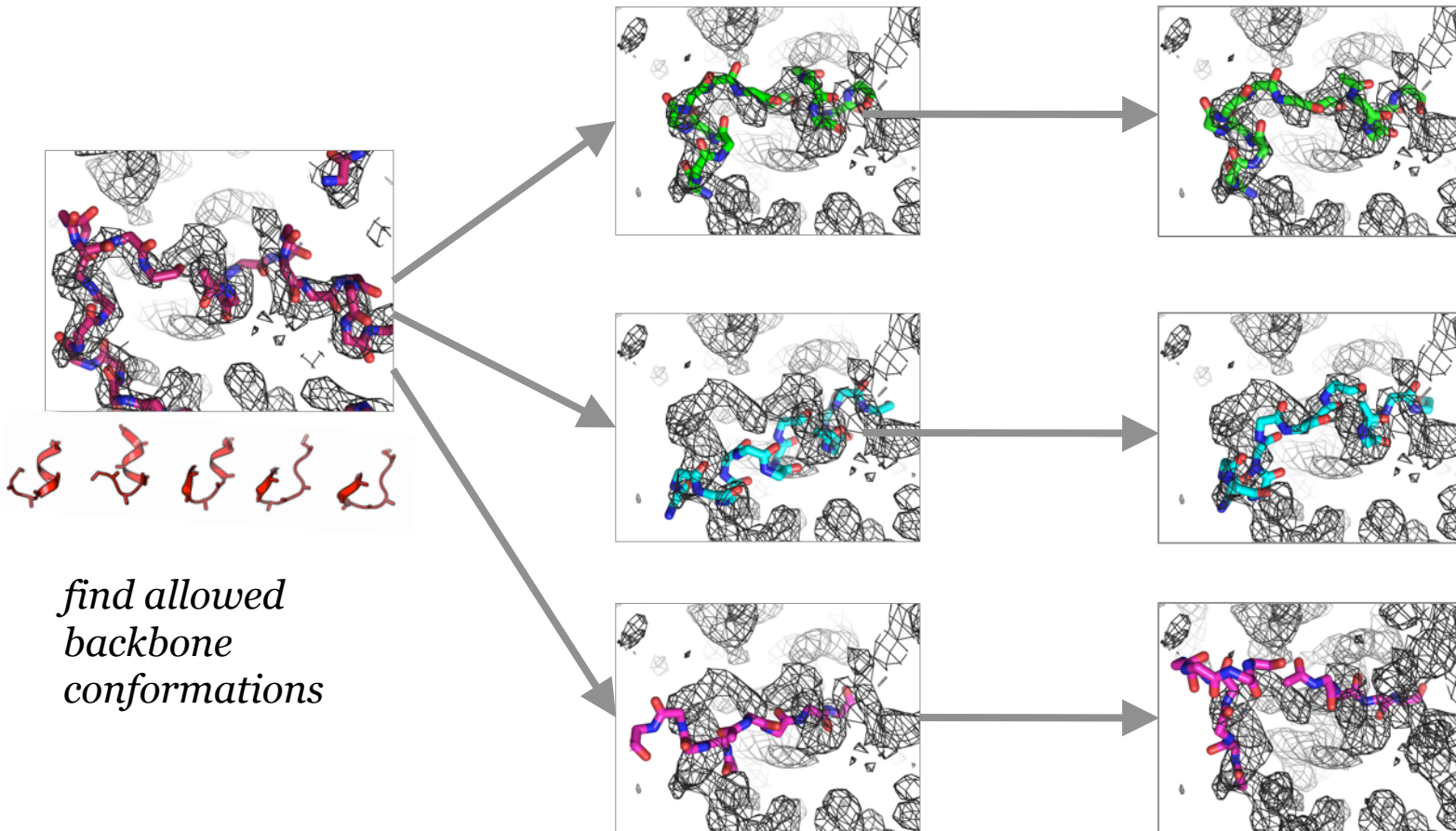


Deposited crystal structure
(1pma)



CryoEM map, real-space B factors

Iterative density-guided conformational sampling



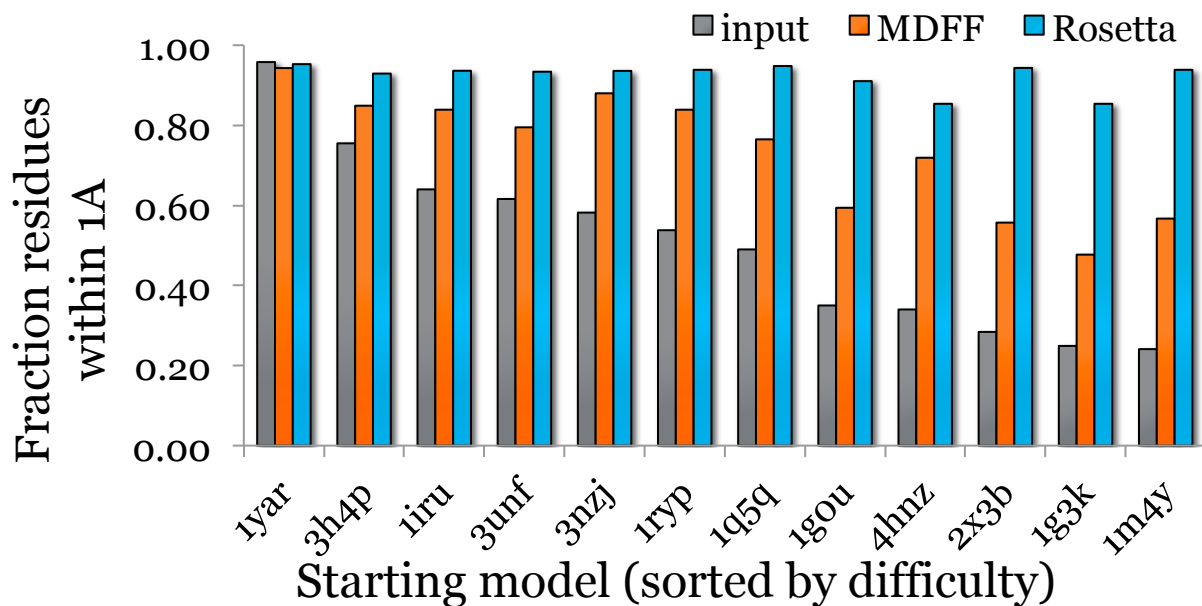
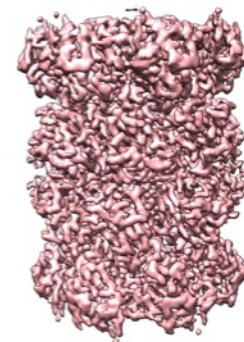
*find allowed
backbone
conformations*

*optimize into density
with minimal forcefield*

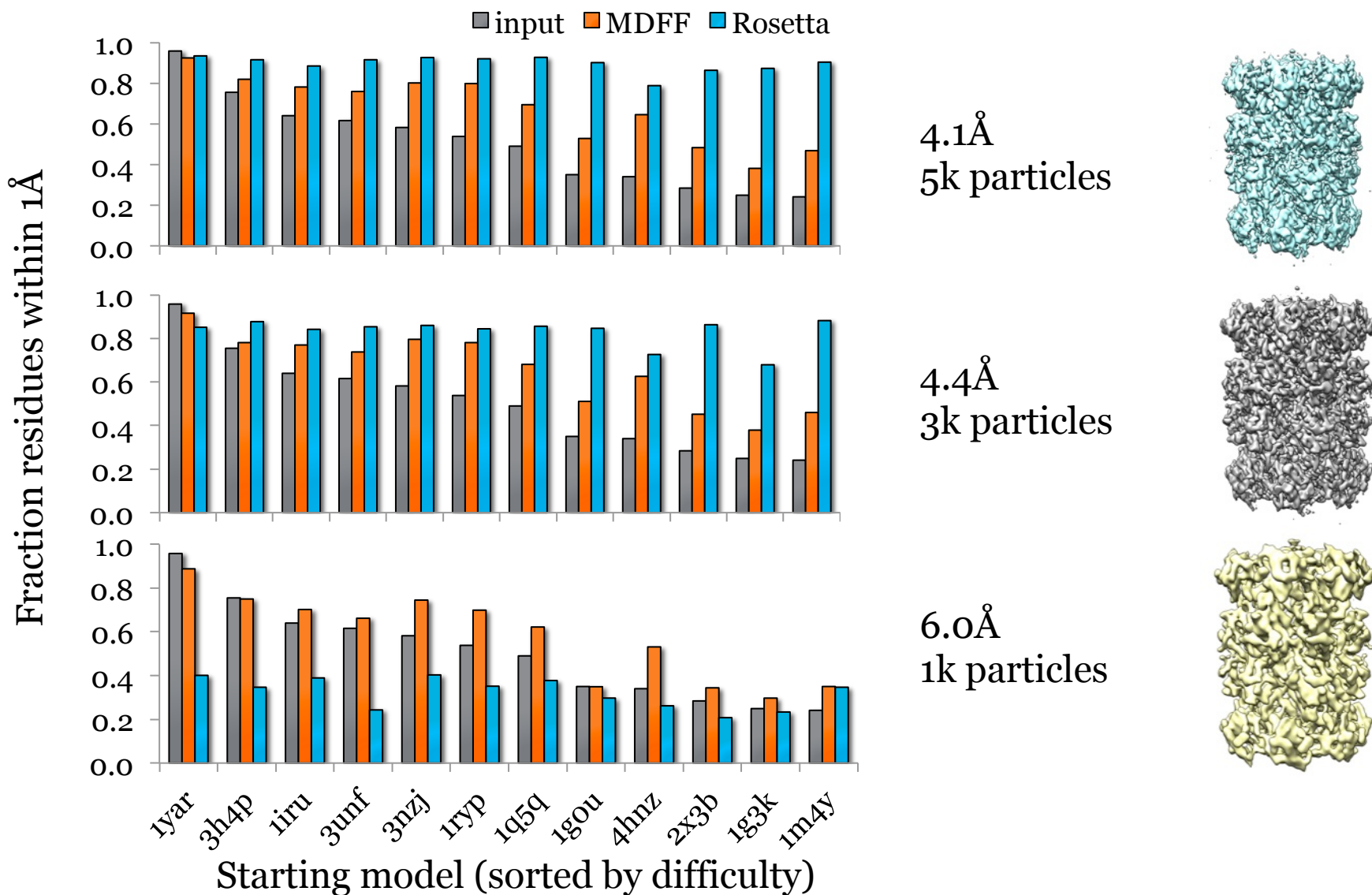
Assessing the role of starting-model quality on structure determination

| Template | Sequence ID |
|-----------|-------------|
| 1yar | 100% |
| 3h4p | 50% |
| 3nzj | 32% |
| 1iru | 30% |
| 1ryp | 30% |
| 1q5q | 26% |
| 3unf | 25% |
| 1m4y | 20% |
| 2x3b/2z3b | 19% |
| 4hnz | 17% |
| 1g3k | 17% |
| 1gou | 17% |

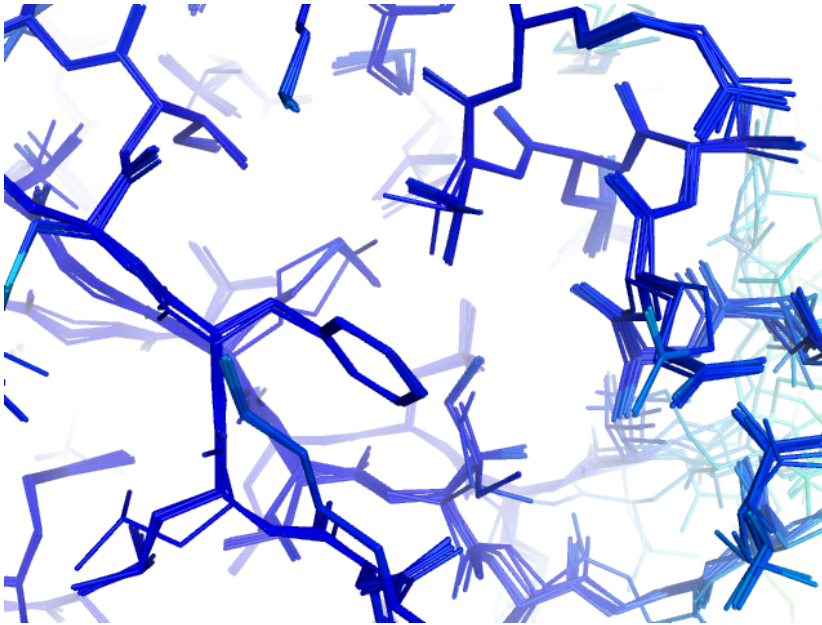
20S proteasome at
3.3Å resolution



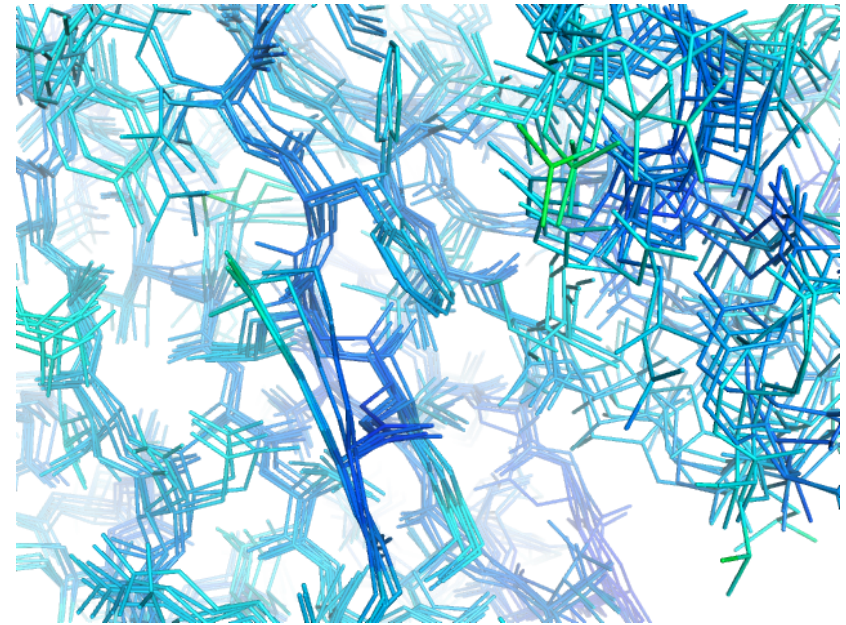
We can accurately determine structures to atomic resolution at 4.4Å or better



Model convergence is an indicator of accuracy

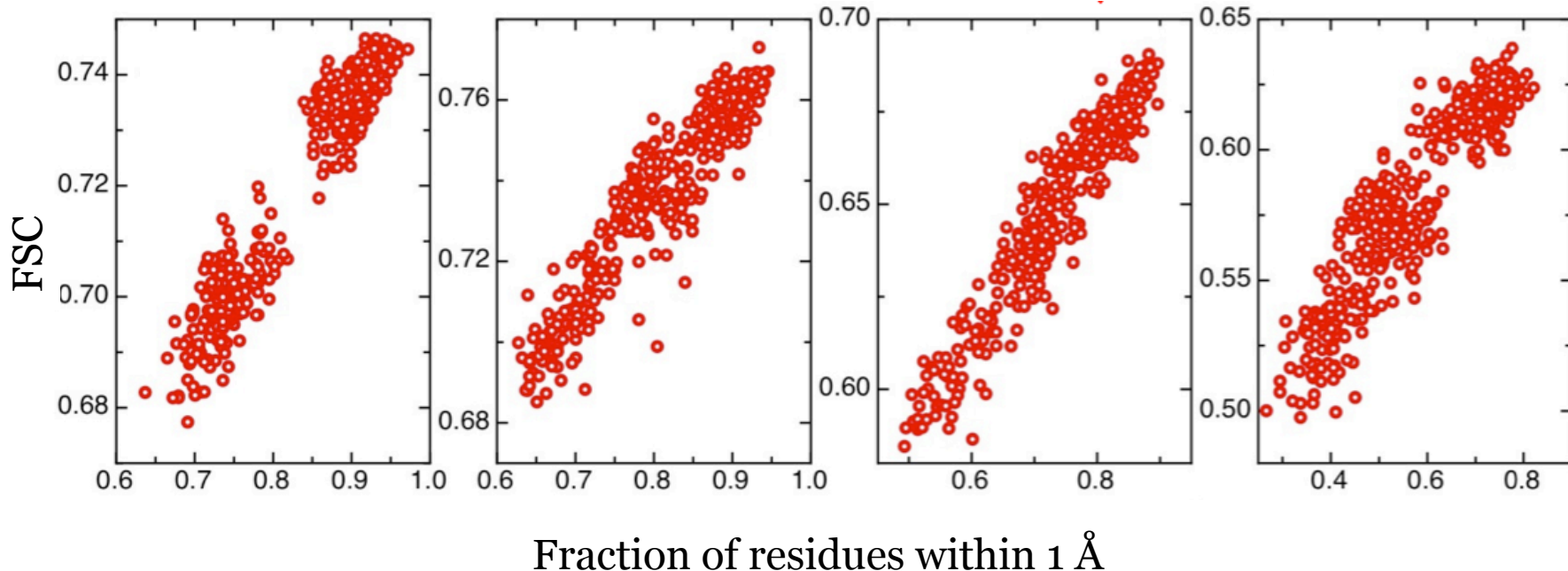


1gou (3.3Å)

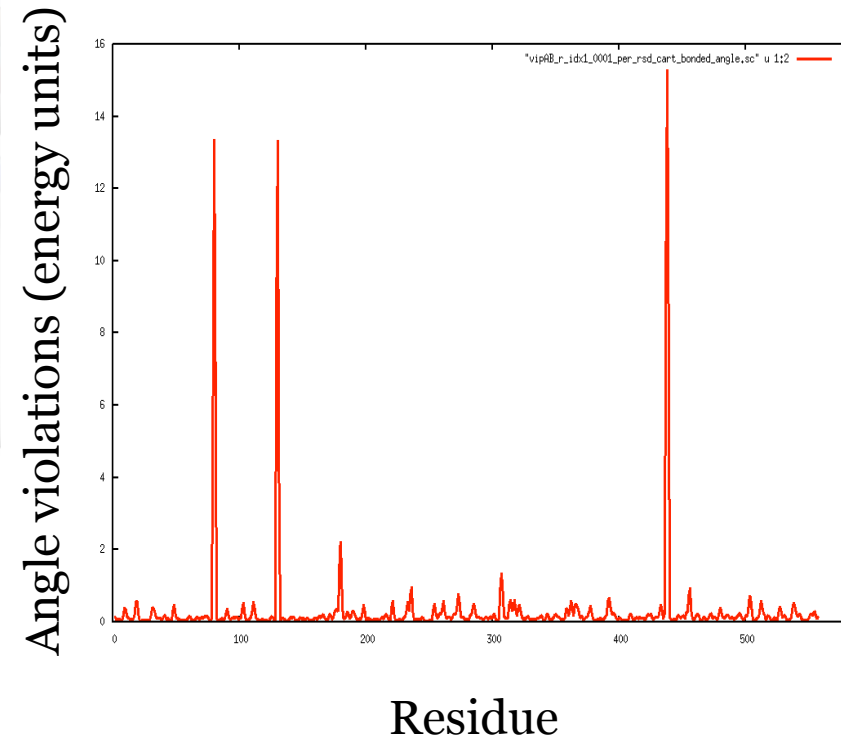
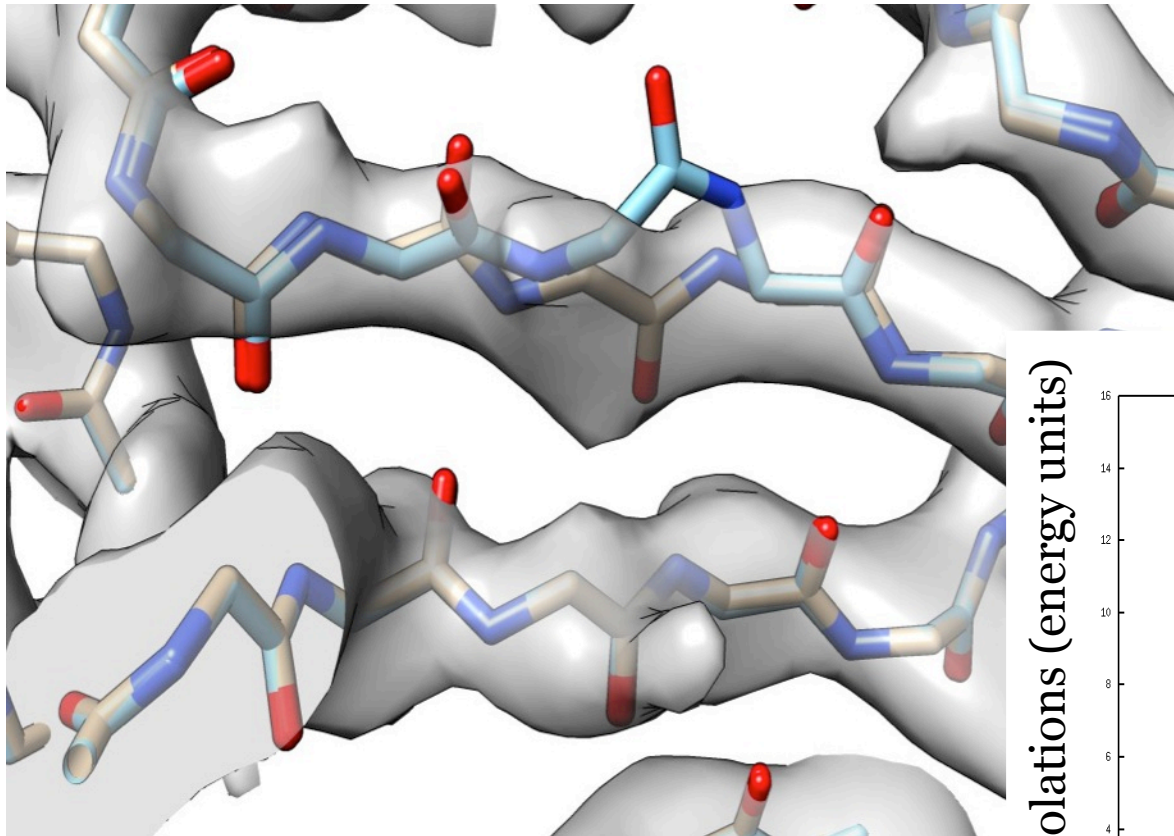


1gou (6.0Å)

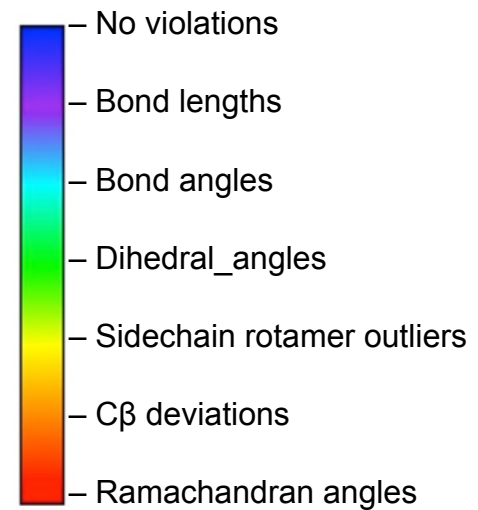
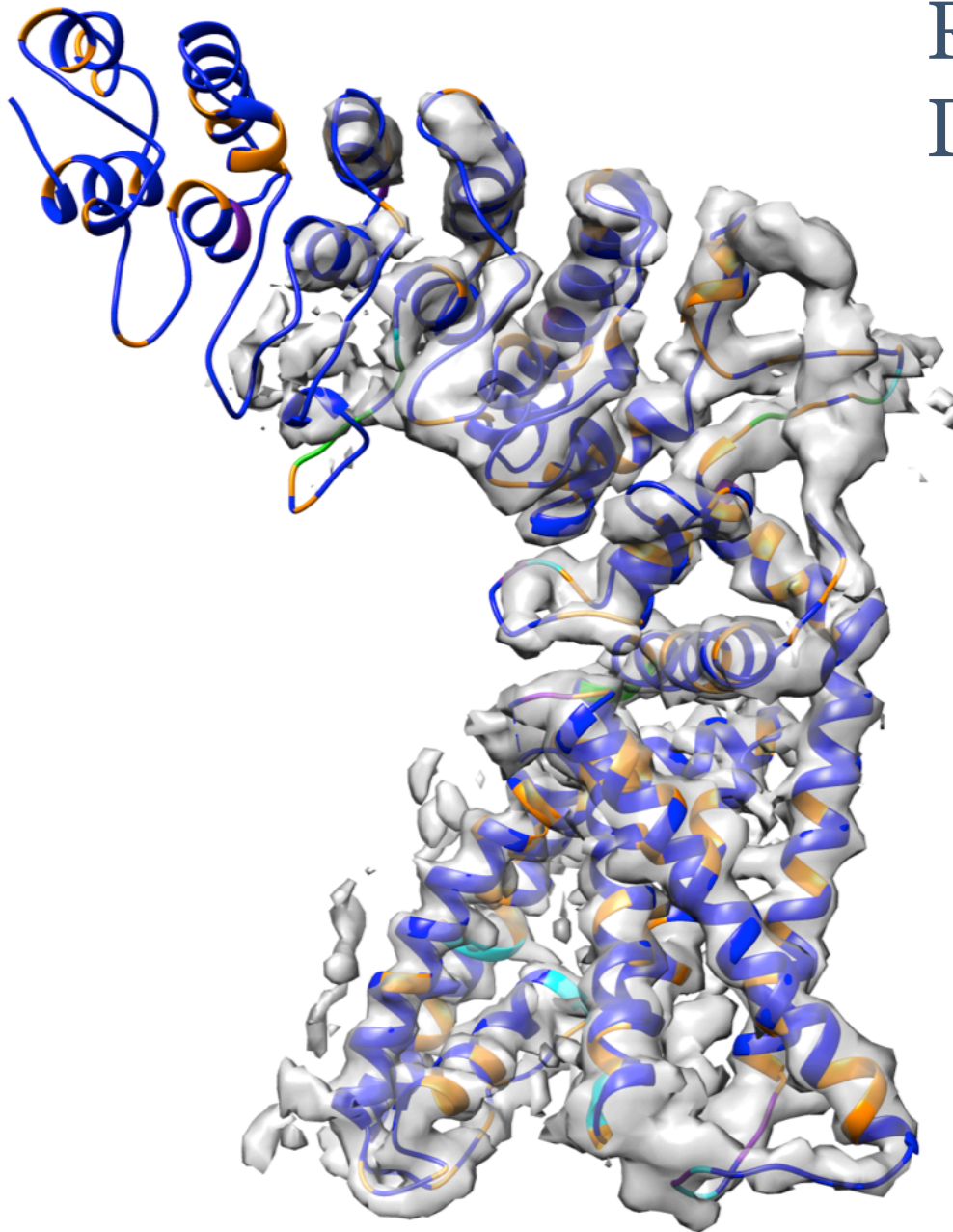
Independent FSC is an indicator of accuracy (though not absolute)



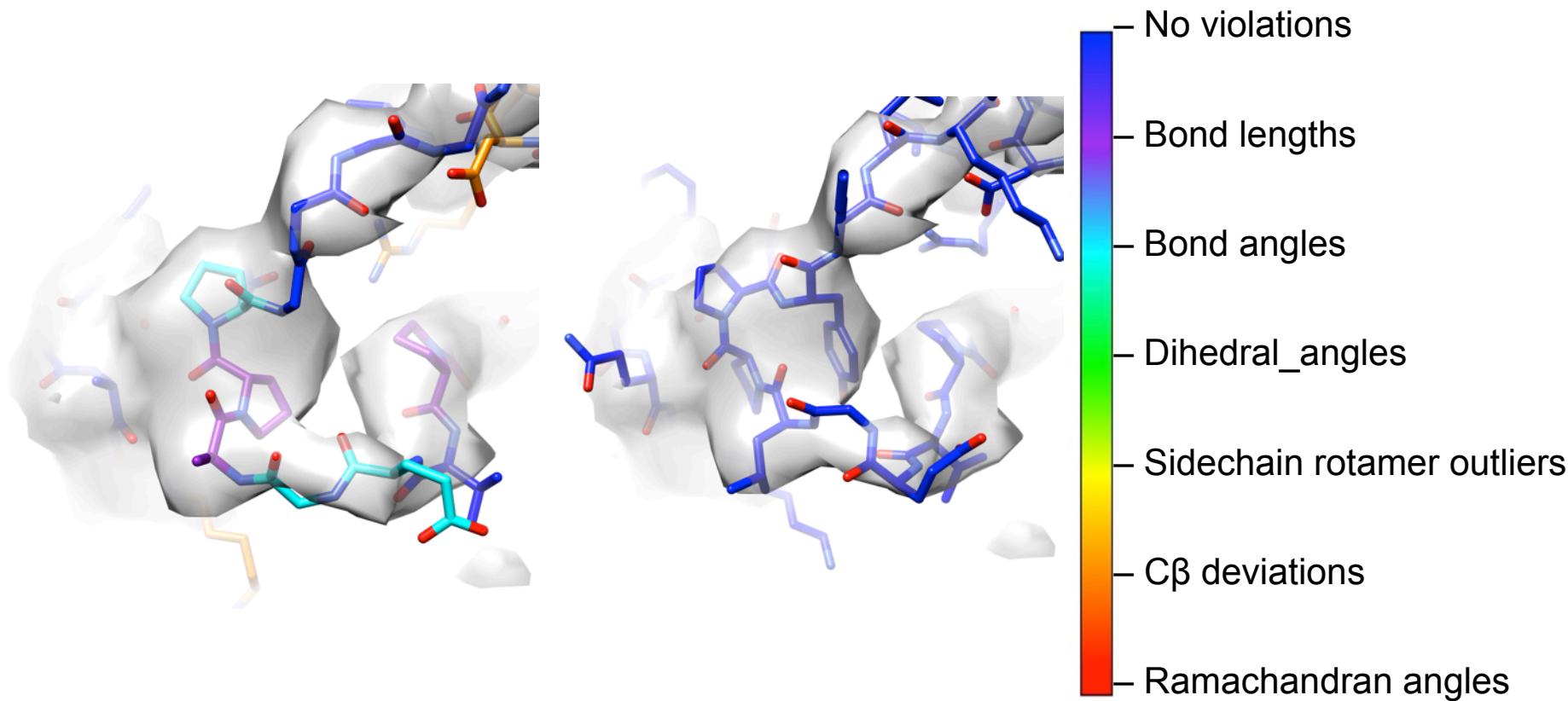
Model strain also can indicate errors



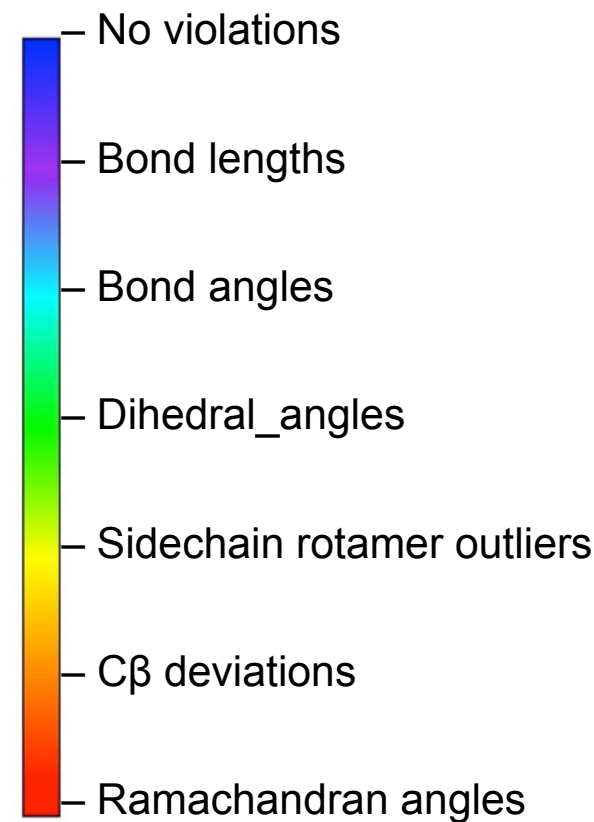
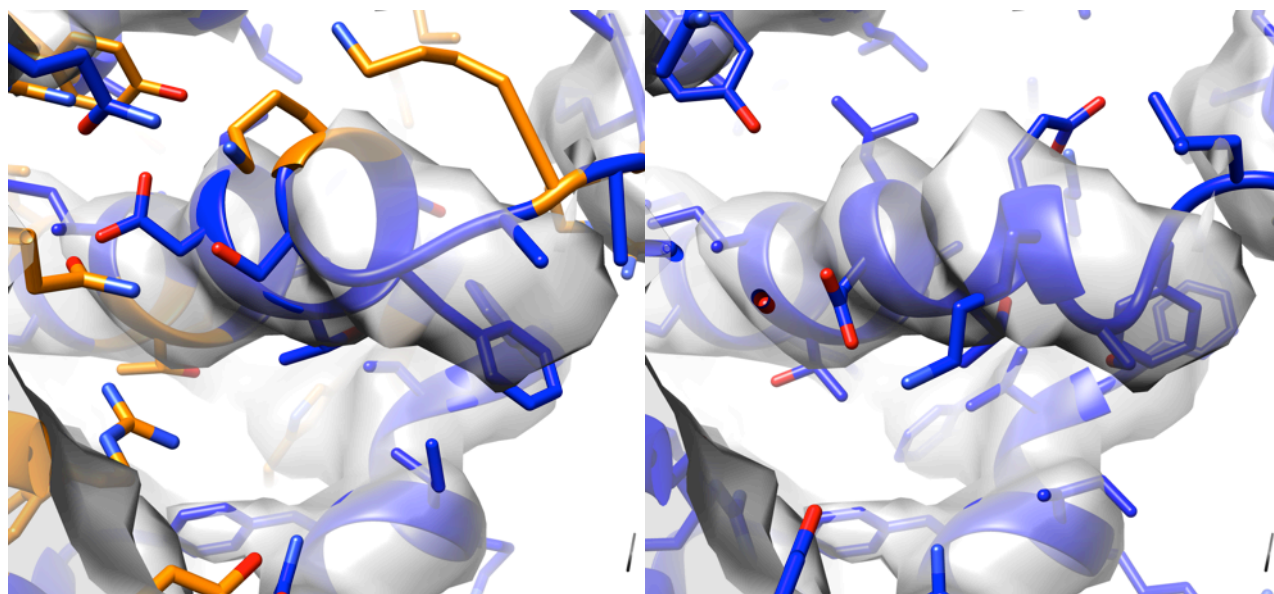
Refinement of TRPV1: Deposited structure



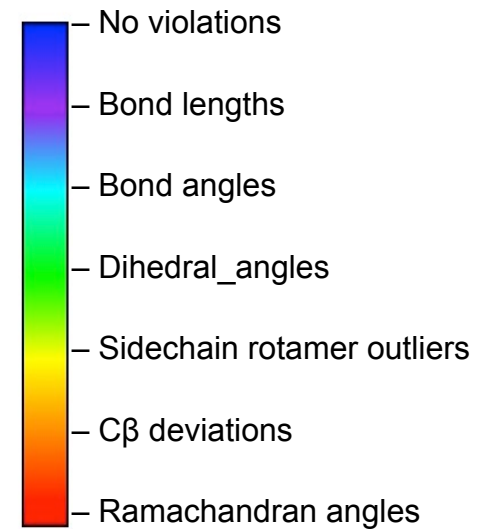
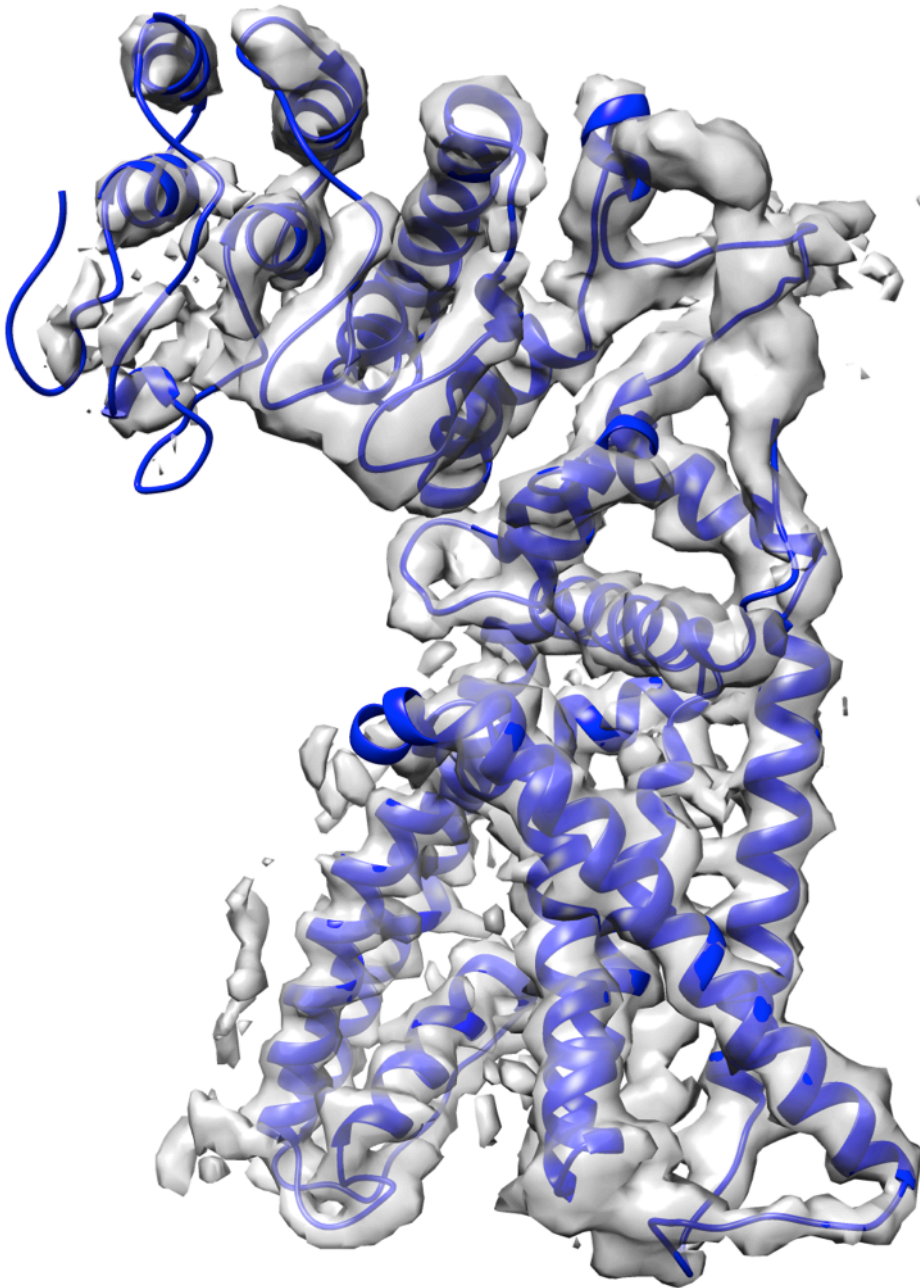
Local strain reveals errors



Local strain reveals errors

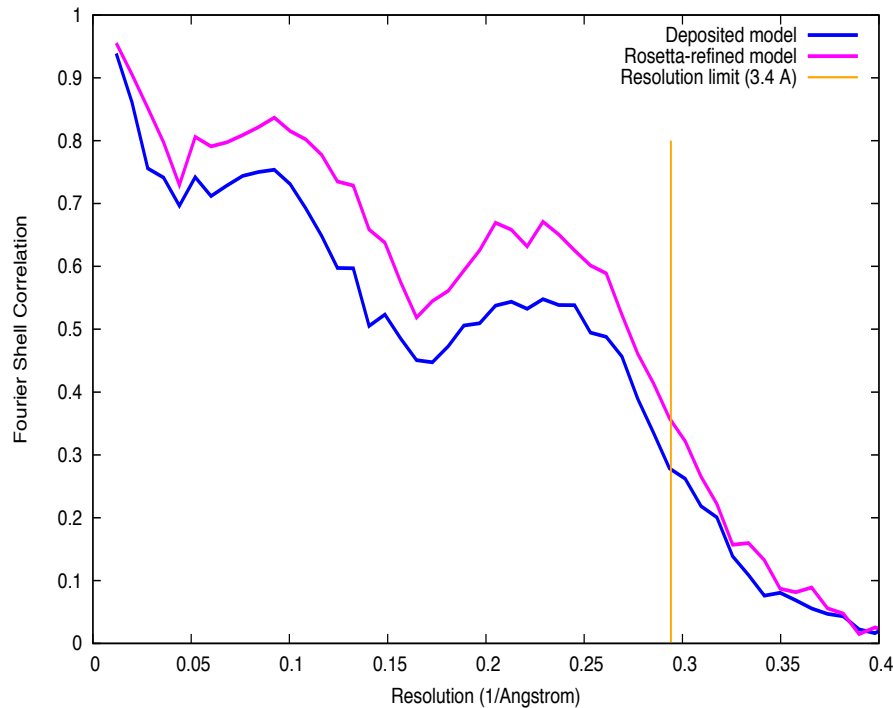


Final refined model

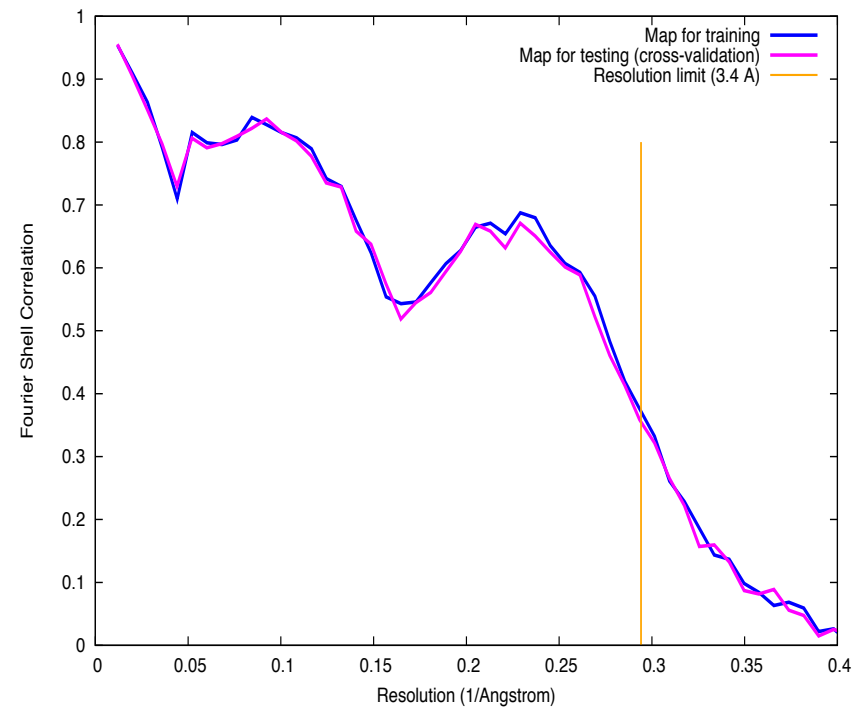


Cross-validation – low/no overfitting

model-map FSC
deposited versus refined



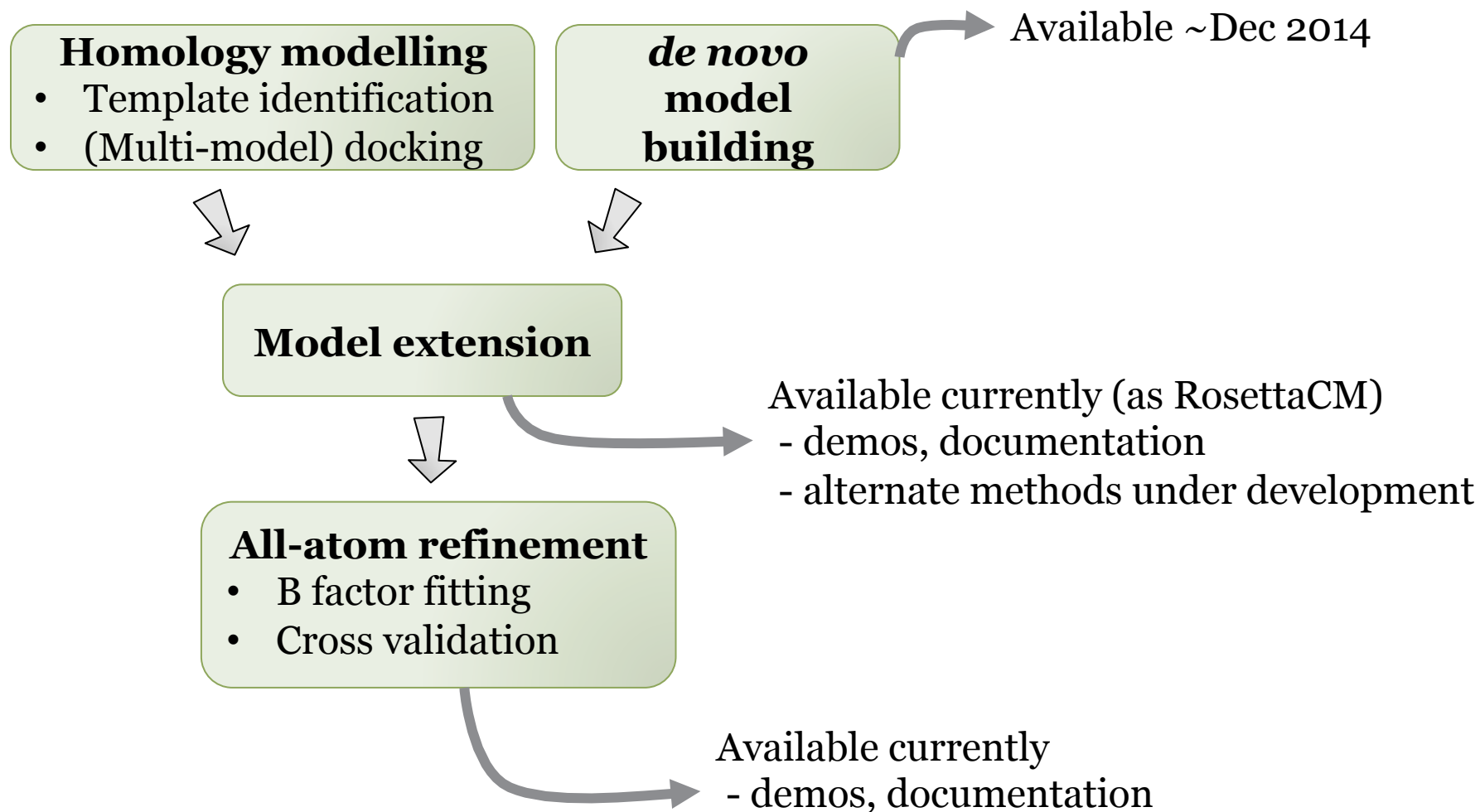
model-map FSC
train versus test



Conclusions

- Atomic accuracy is possible from near-atomic resolution (up to 4.5Å) data
- Have we solved it? Do we have...
 - Good fit to independent data (locally and globally)?
 - No model strain / molprobability outliers?
 - Well converged ensemble of solutions satisfying the above two?

Method availability



Acknowledgements

- Collaborators
 - Wah Chiu (Baylor), Junjie Zhang (Texas A&M)
 - Tom Marlovits (IMBA, Austria)
 - Ed Egelman (U. Virginia)
 - Misha Kudryashev, Marek Basler (U. Basel)
 - Xueming Li, Yifan Cheng (UCSF)
- Students & Postdoc
 - **Ray Wang**
 - Patrick Conway
 - Brandon Frenz
 - Zibo Chen
 - Ryan Pavlovicz