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**NMFF: Flexible fitting of atomic structures into
low-resolution EM maps using elastic network
normal mode analysis**

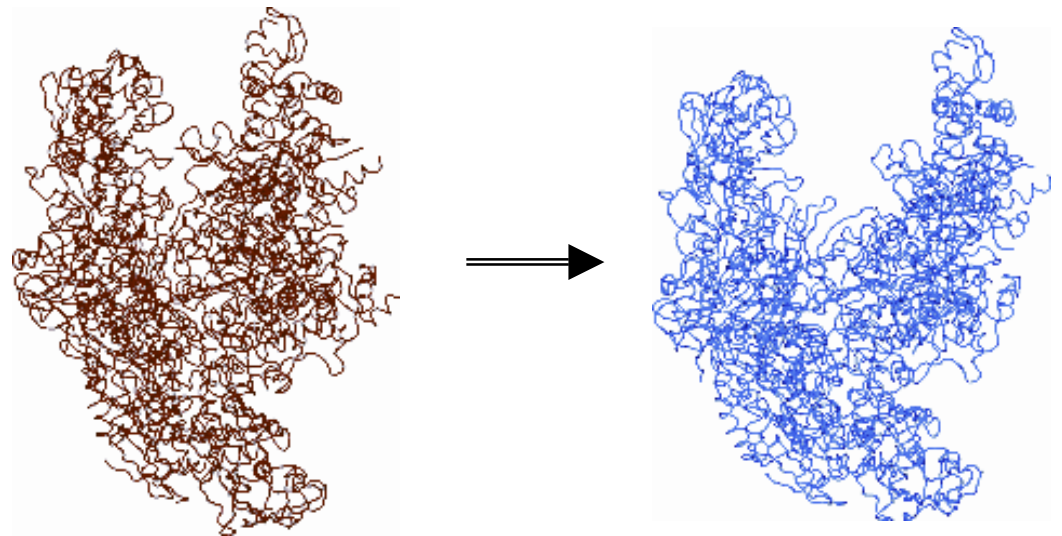
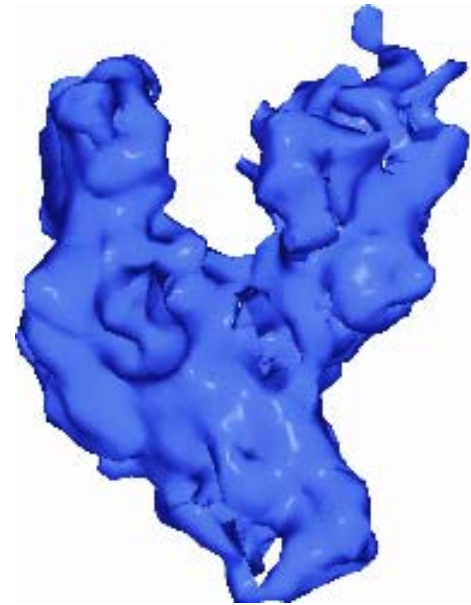
Florence Tama, Osamu Miyashita and Charles L. Brooks III

Flexible fitting

Growing number of low-resolution structure

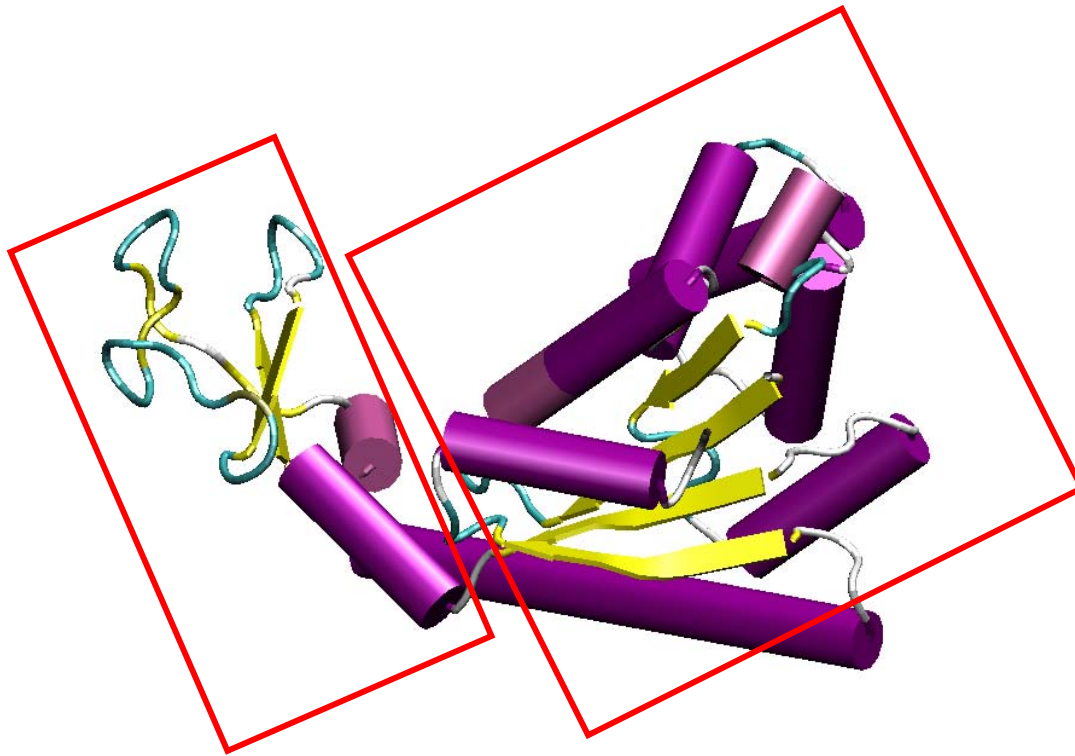
Fitting of high resolution structure
into low-resolution structure

Conformation between X-ray
structure and EM map may be
different => flexible fitting



Flexible fitting

Allow domains to rotate/translate as rigid body



➤ Division of protein is ad-hoc => may lead to different results

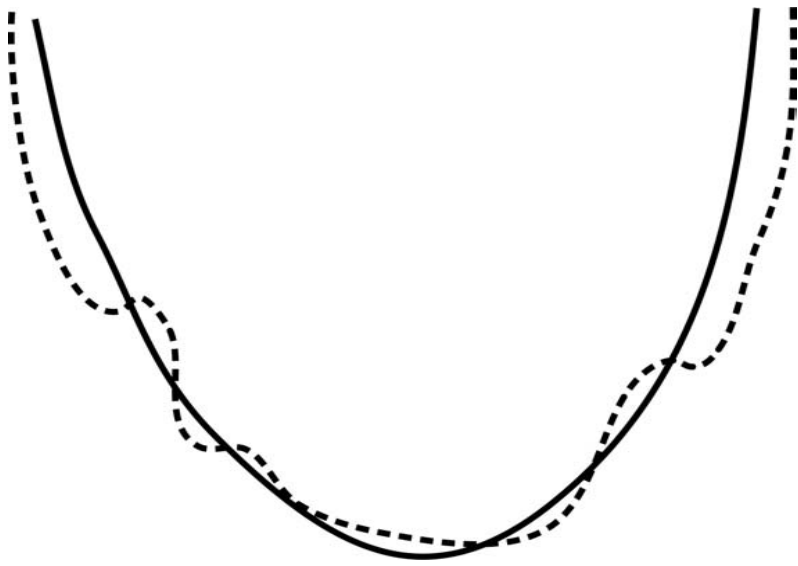
➤ correlated motions between domains are not taken into account

=> Quantitative techniques with reproducible results

Development of a refinement program using Normal mode Analysis

Normal mode analysis: Theory

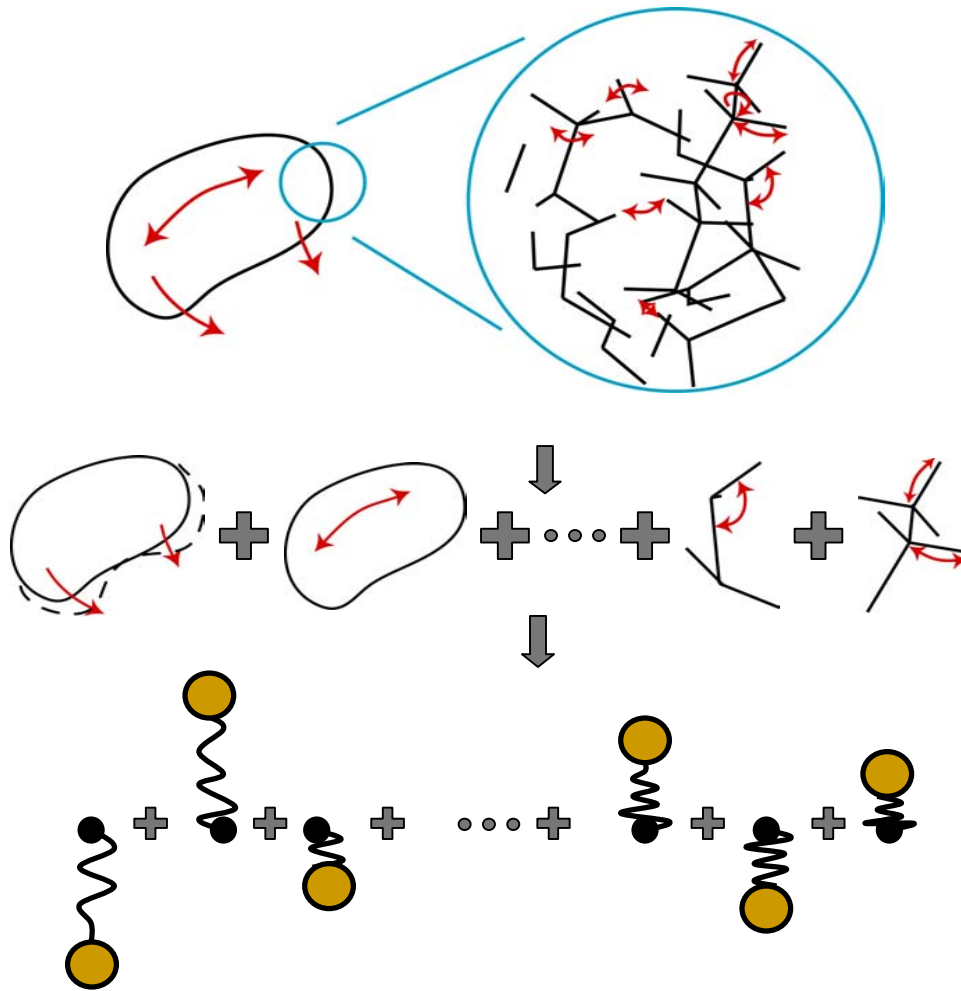
Potential energy \Rightarrow harmonic



Dynamics = harmonic potential
independent

$$L = \frac{1}{2} \sum_{i=1}^{3N-6} \dot{q}_i^2 - \frac{1}{2} \sum_{i=1}^{3N-6} \omega_i^2 q_i^2$$

Normal mode analysis



Low frequencies

High frequencies

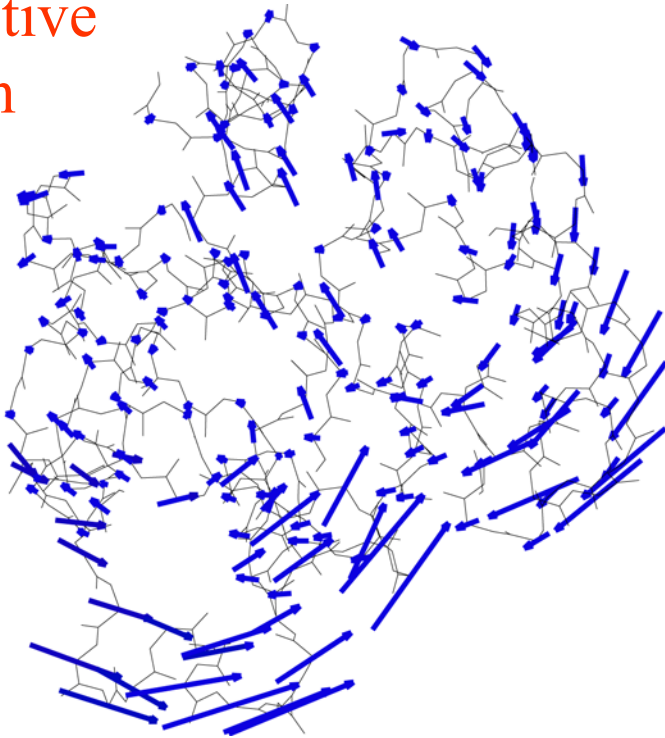
Large collective motions

Localized motions

Examples

Adenylate kinase:
Mode 1 (2.95 cm^{-1})

Collective
motion

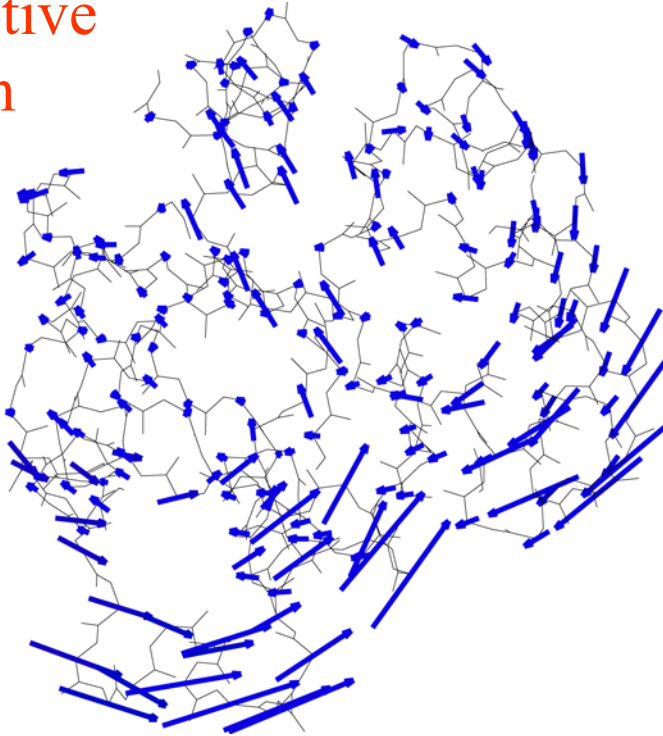


Examples

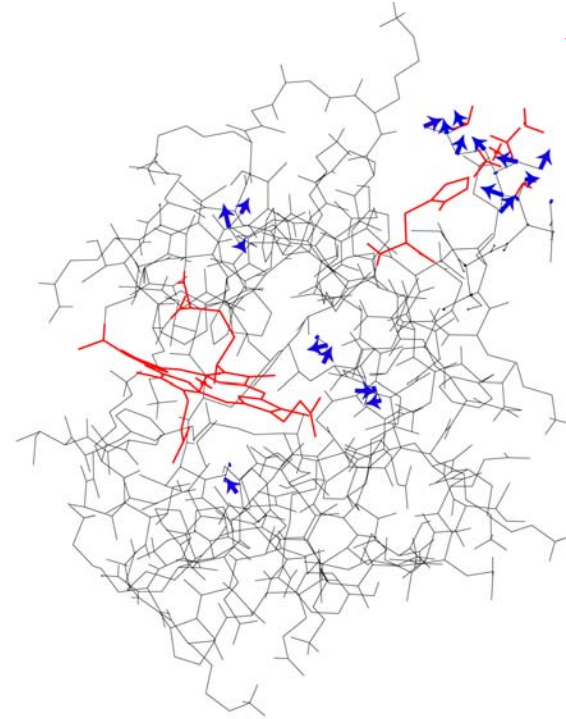
Adenylate kinase:
Mode 1 (2.95 cm^{-1})

Cytochrome c:
Mode 2757 (1519 cm^{-1})

Collective
motion



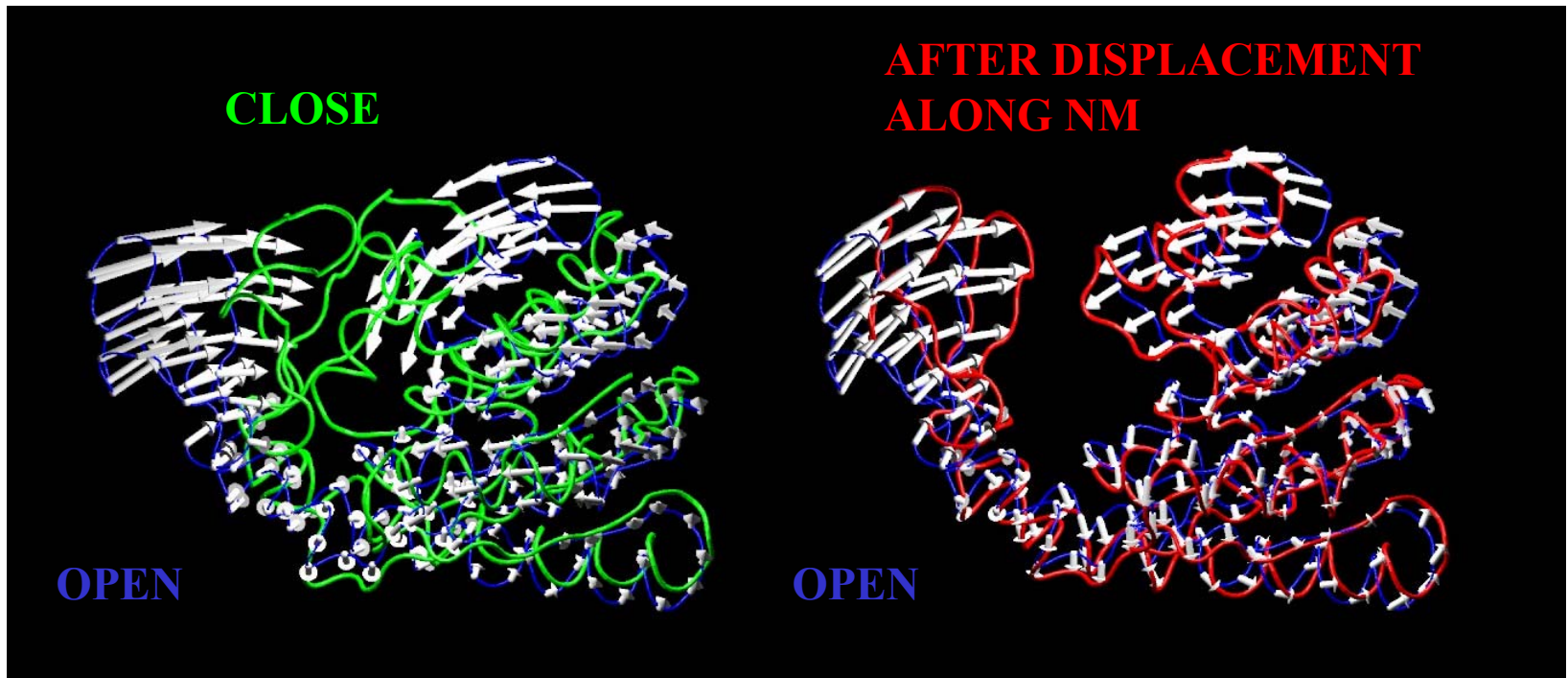
Localized
motion



NMA gives conformational changes

Adenylate Kinase =>

large conformational change upon ligand binding



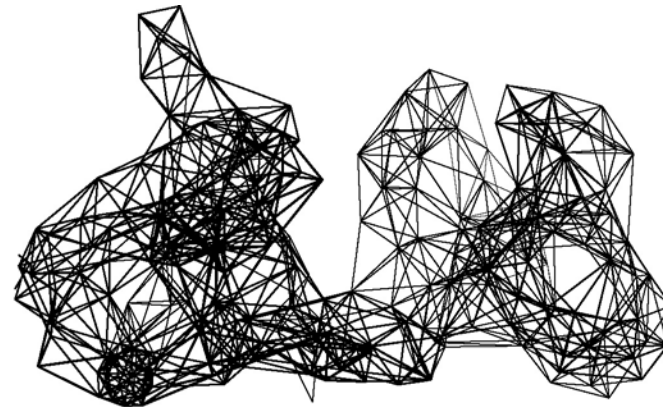
1 normal mode can represent up to 80-90 % of the overall conformational change

Elastic network model

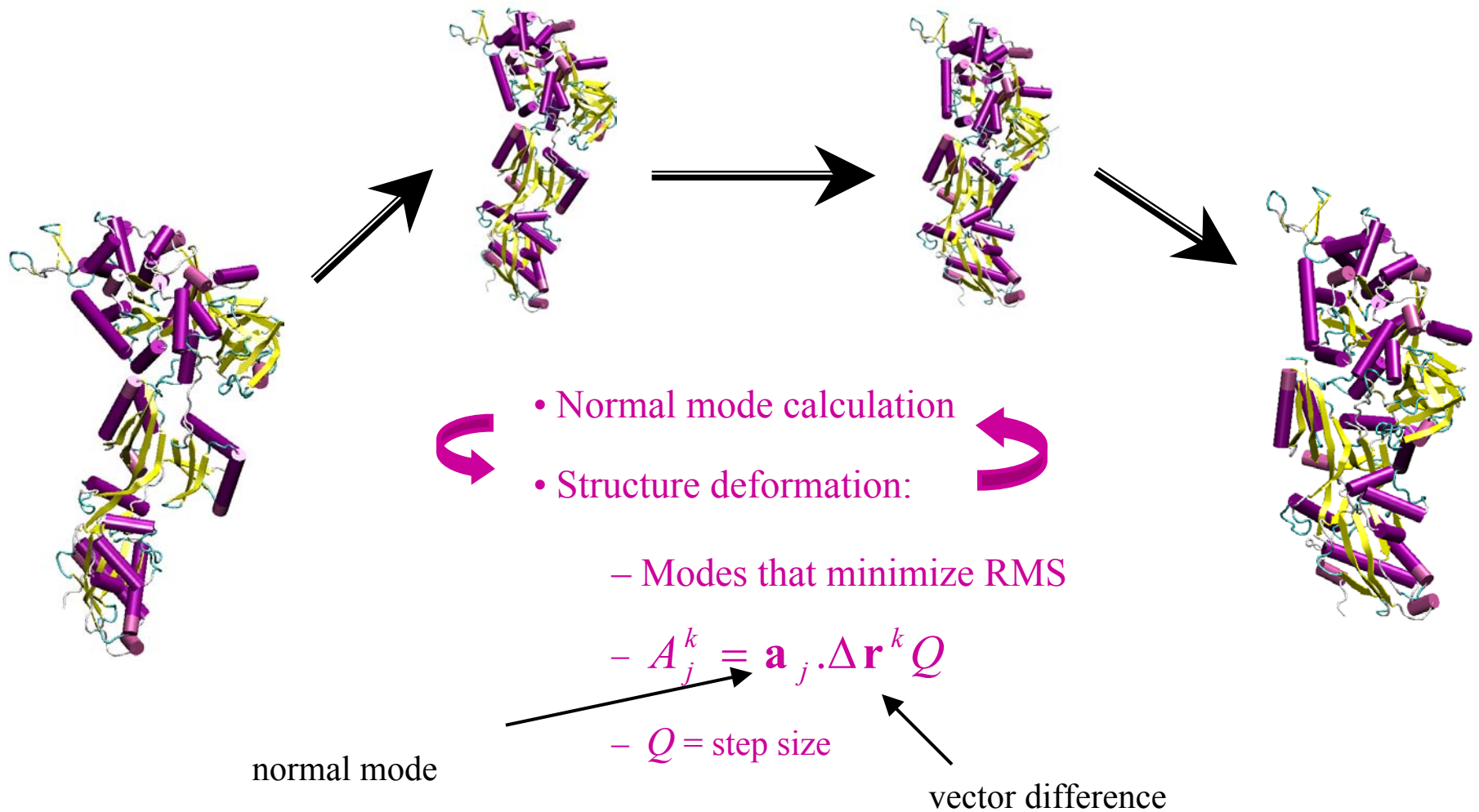
Atoms connected via elastic springs

$$E(r_a, r_b) = \frac{C}{2} \left(|r_{a,b}| - |r_{a,b}^0| \right)^2$$

$$E_p = \sum_{a,b} E(r_a, r_b)$$

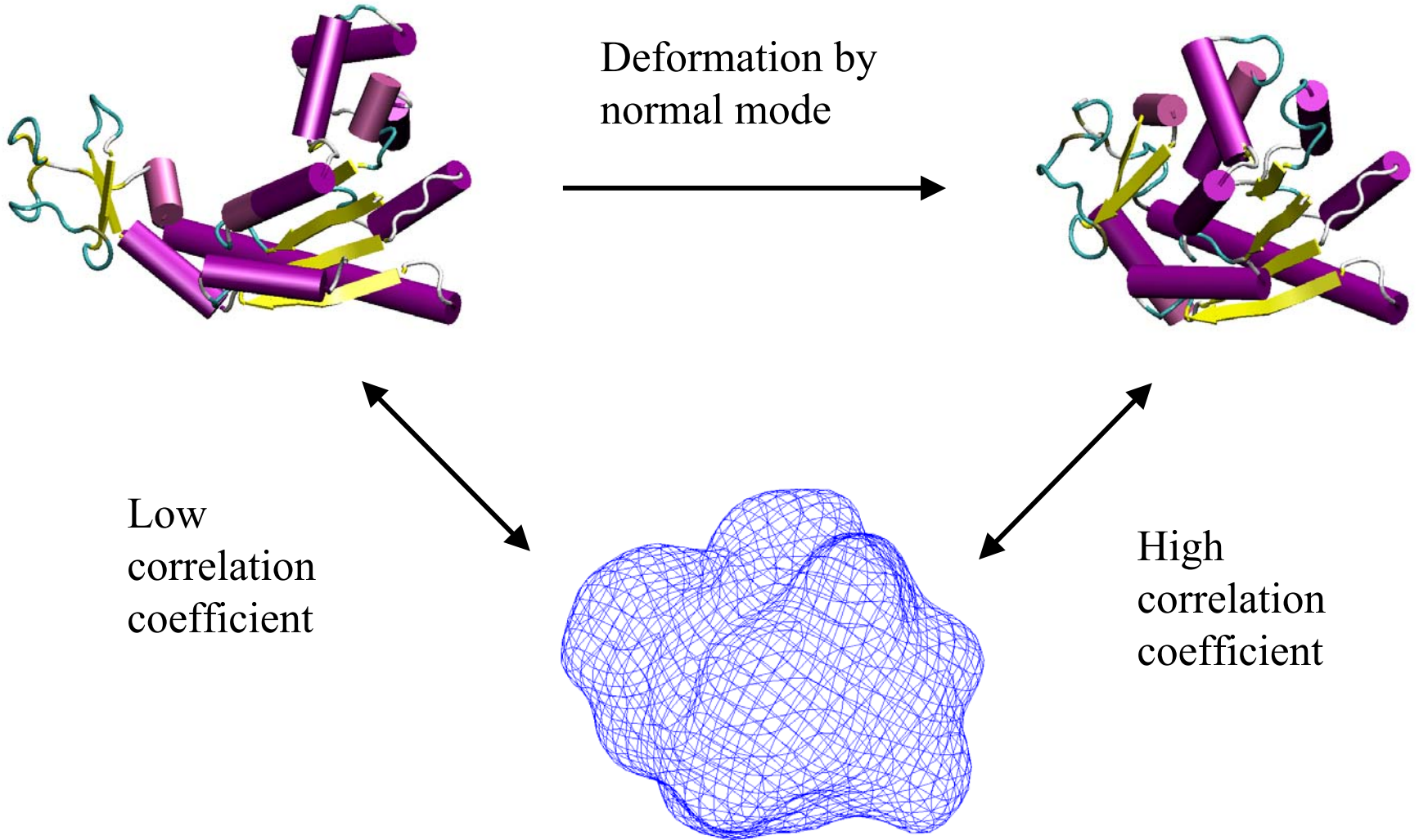


Exploring conformational transitions using NMA

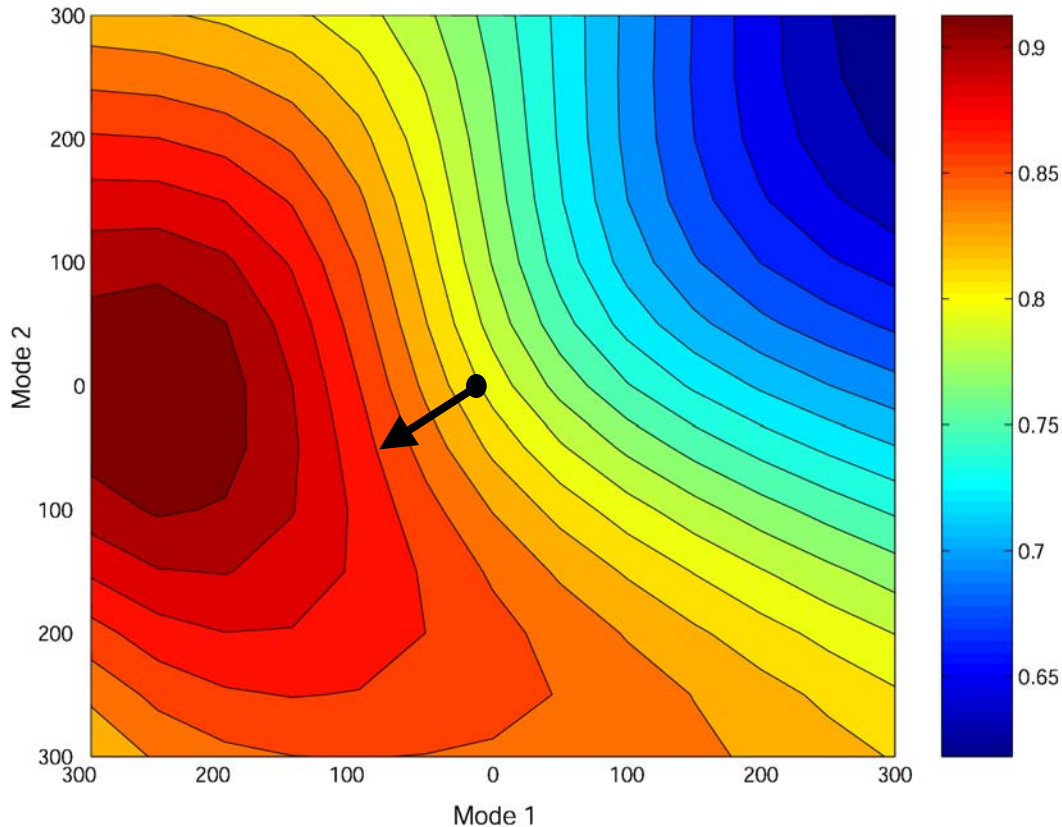


Selection of normal modes that may contribute to the transition

Normal Mode Flexible Fitting



Correlation coefficient: Maximization problem



➤ Maximization of the correlation coefficient as a function of normal mode coordinate

➤ Identification of the relevant normal modes

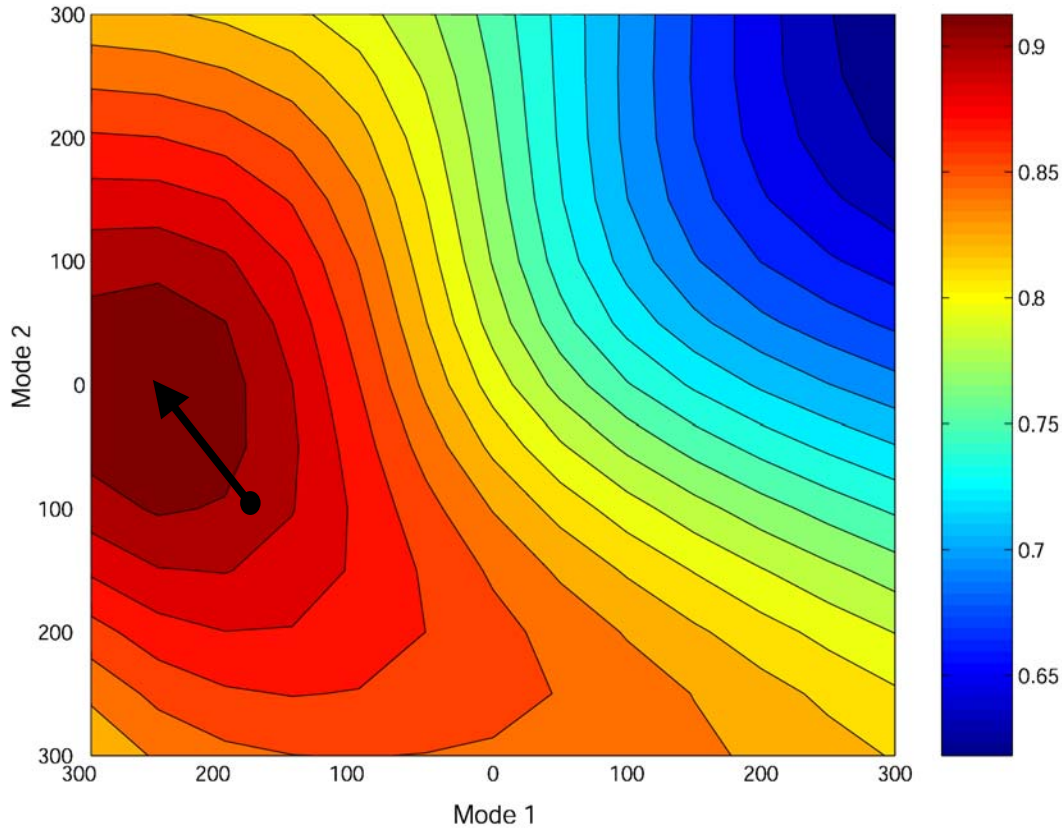
Steepest Descent
/Newton Raphson



Gradient/Hessian

$$F_k = \frac{\partial \text{c.c.}}{\partial q_k}$$

Correlation coefficient: Maximization problem



Near a maximum

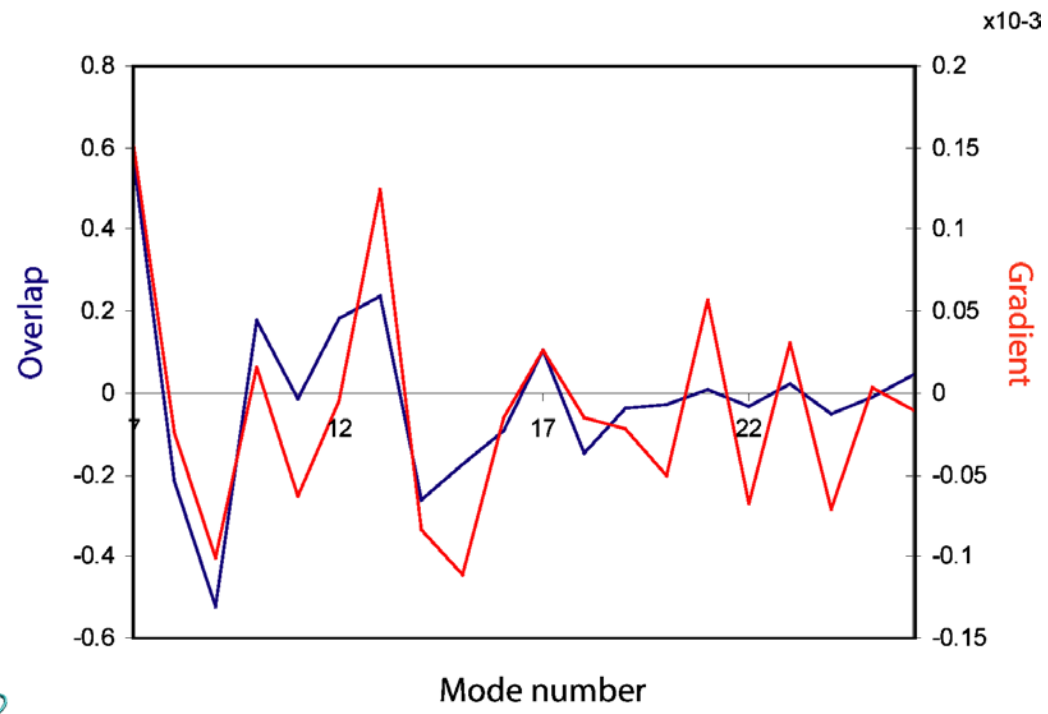
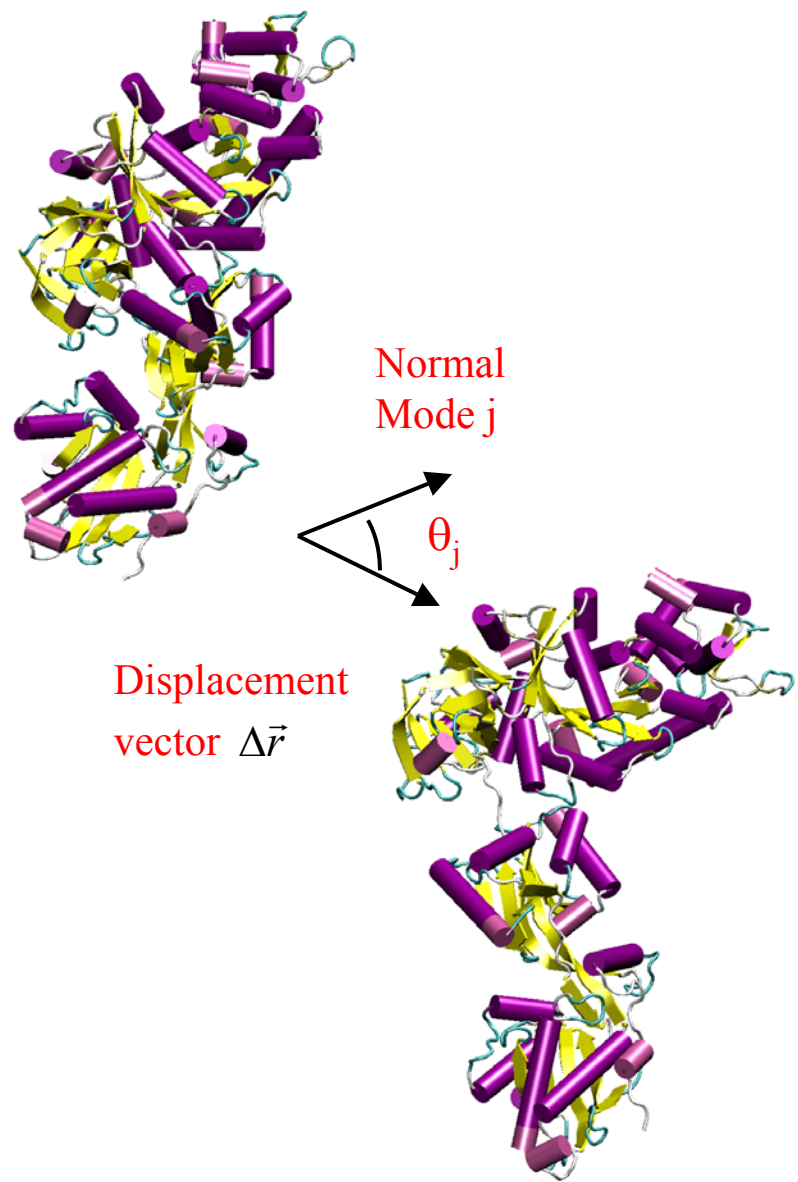
Steepest Descent
/Newton Raphson



Gradient/Hessian

$$F_k = \frac{\partial^2 \text{c.c.}}{\partial q_k \partial q_j}$$

Derivative of correlation coefficient by normal mode coordinates

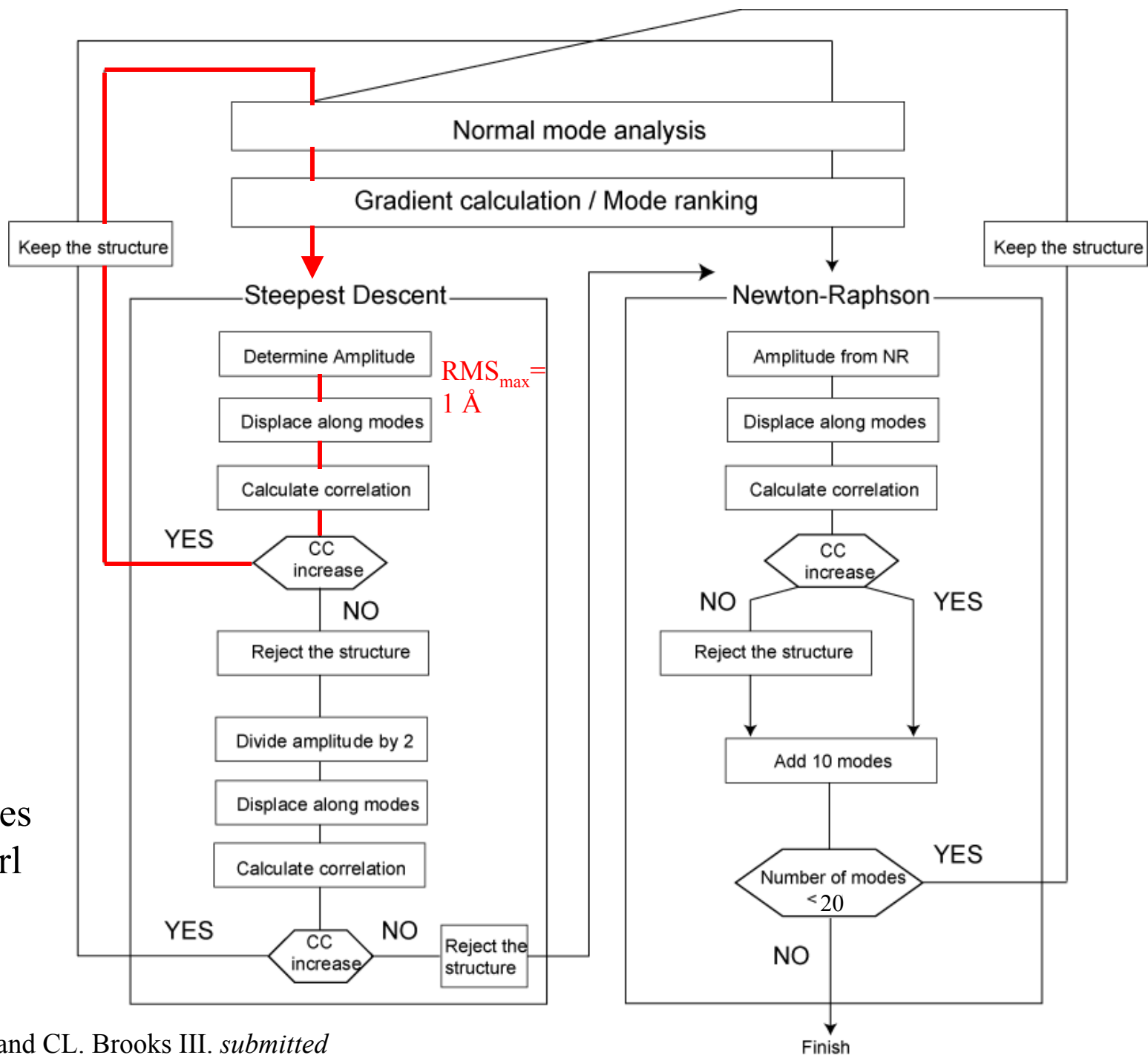


$$F_k = \frac{\partial \text{c.c.}}{\partial q_k}$$

NMFF: Normal Mode Flexible Fitting program

Iterative
procedure

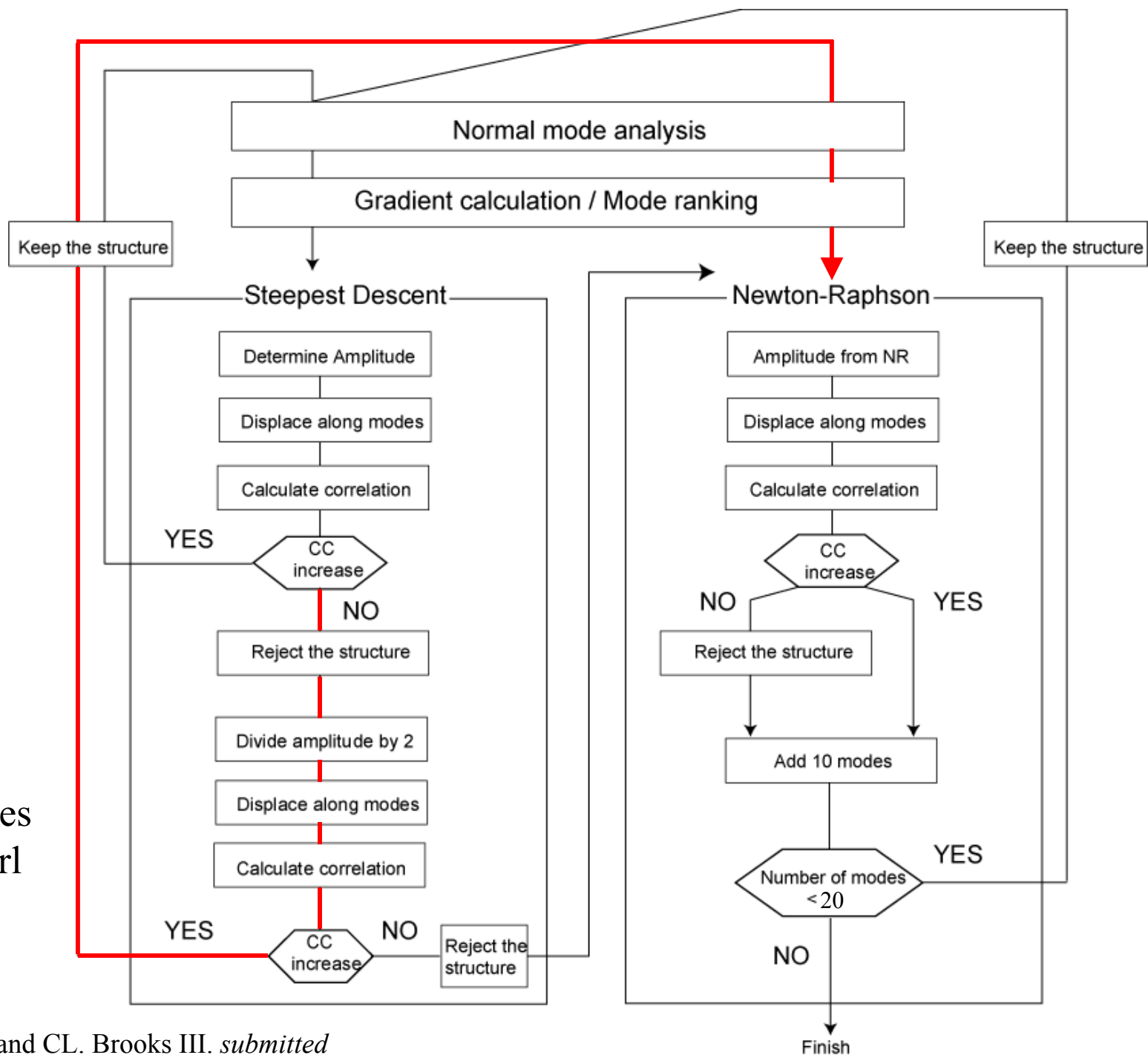
Fortran / C codes
managed by perl
script.



NMFF: Normal Mode Flexible Fitting program

Iterative
procedure

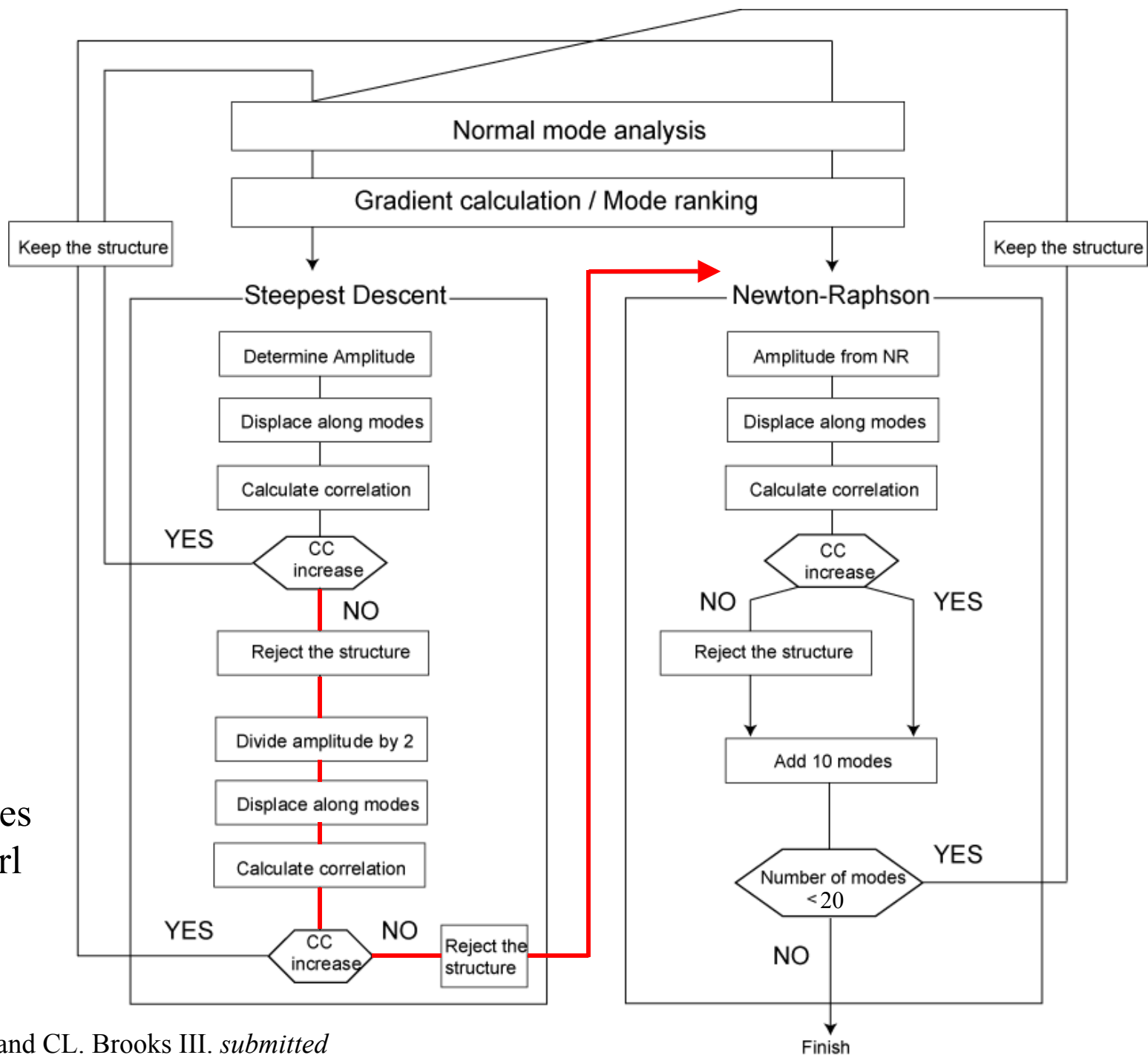
Fortran / C codes
managed by perl
script.



NMFF: Normal Mode Flexible Fitting program

Iterative
procedure

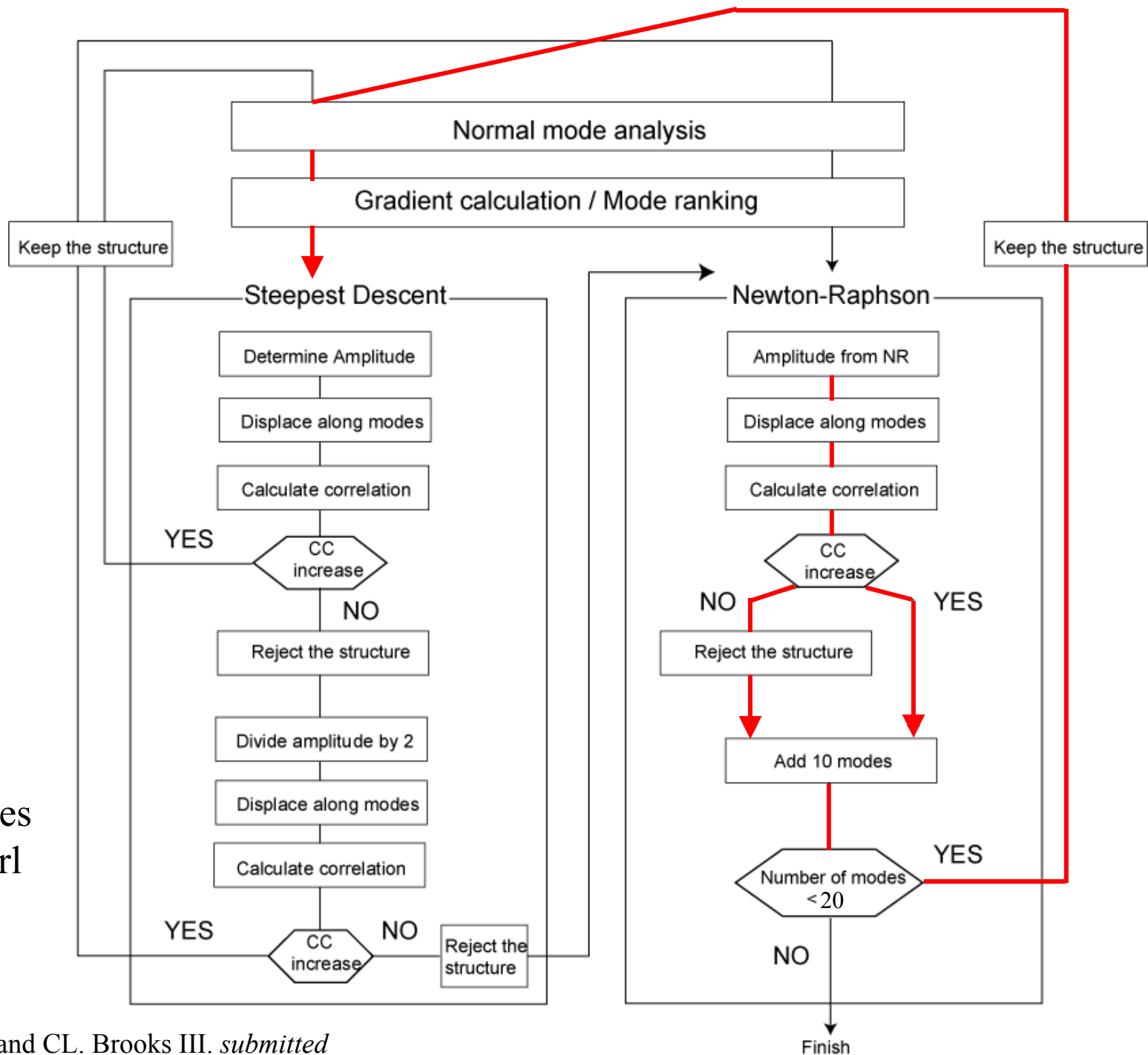
Fortran / C codes
managed by perl
script.



NMFF: Normal Mode Flexible Fitting program

Iterative
procedure

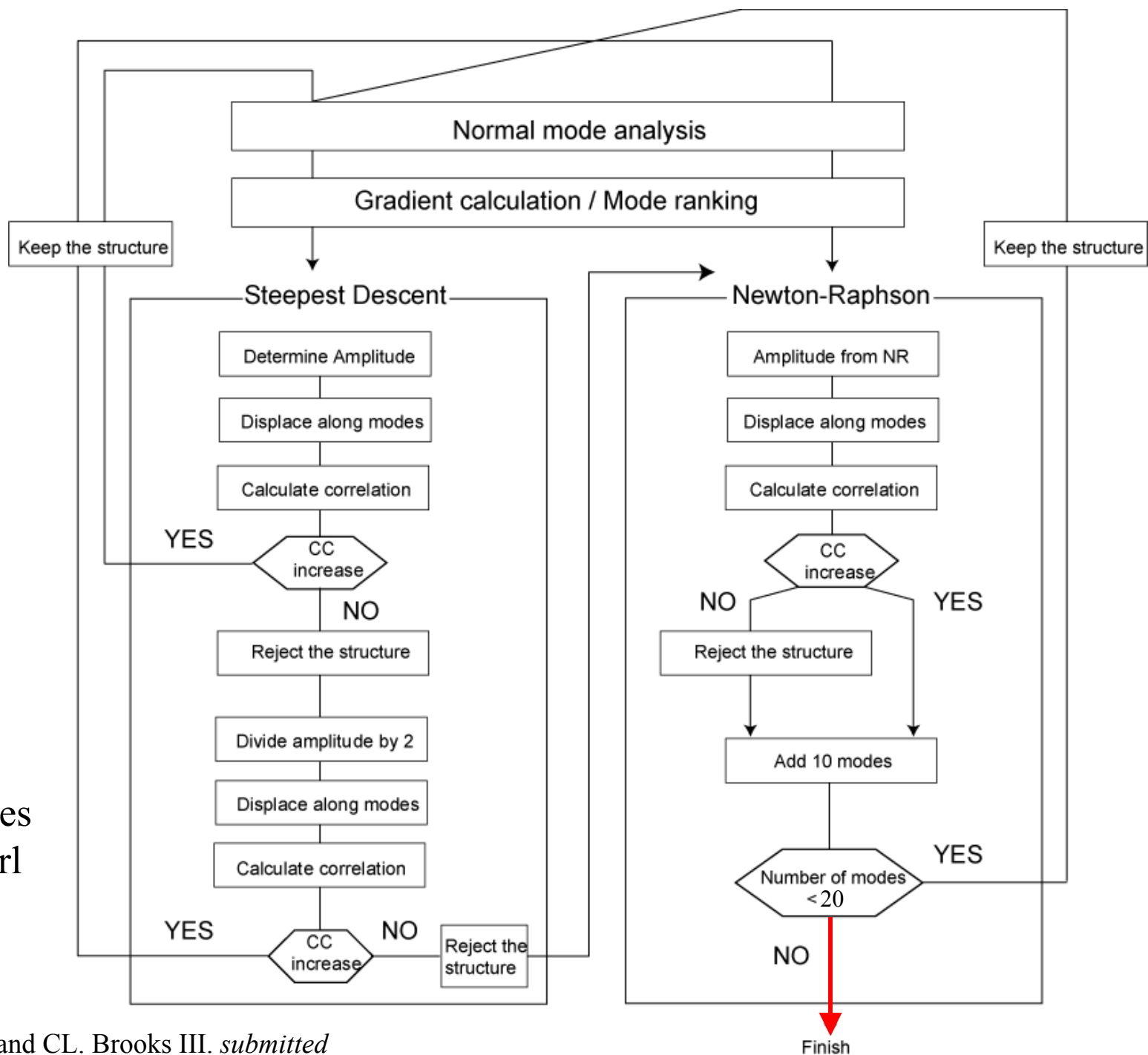
Fortran / C codes
managed by perl
script.



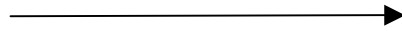
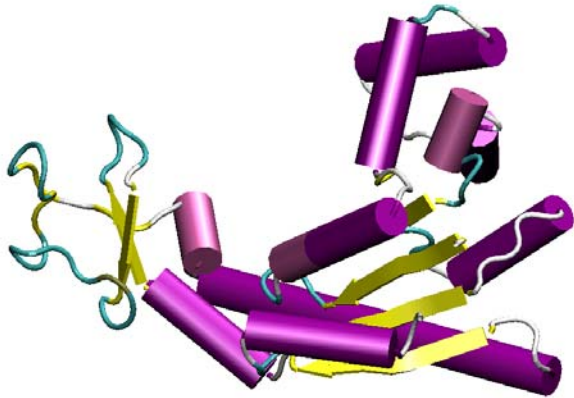
NMFF: Normal Mode Flexible Fitting program

Iterative
procedure

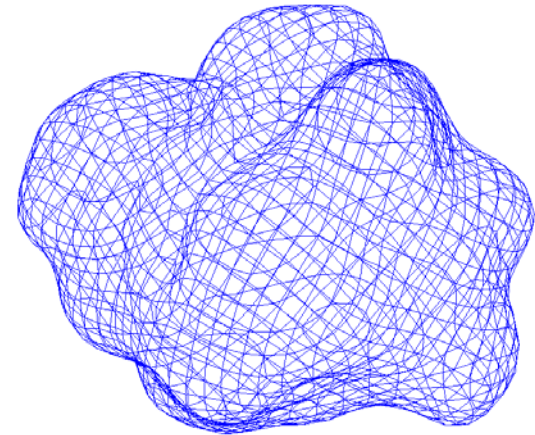
Fortran / C codes
managed by perl
script.



Validation of our approach

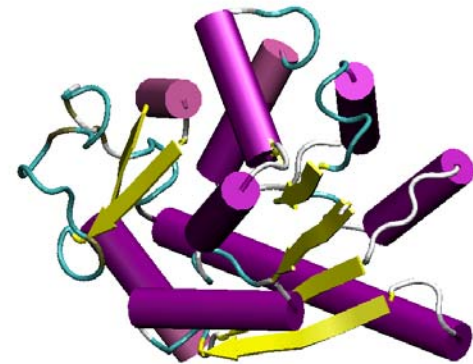


Deformation of
the structure
along NM



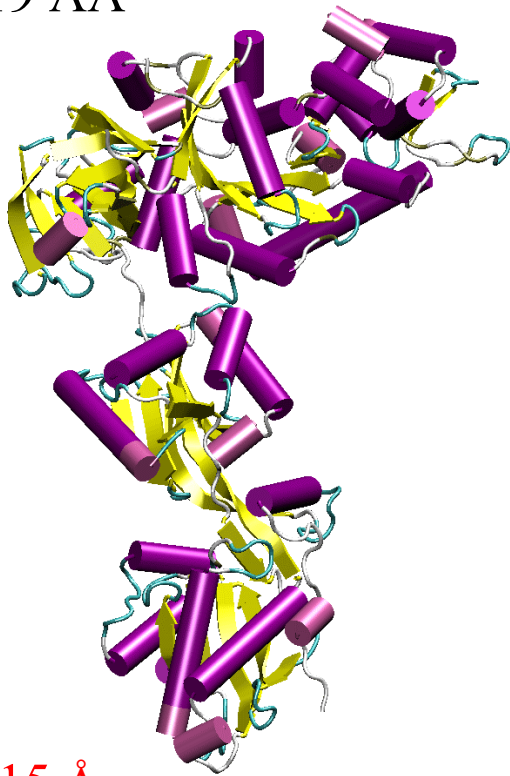
Lower
resolution

Conformation 1 and 2 are
superimposed by RMSD



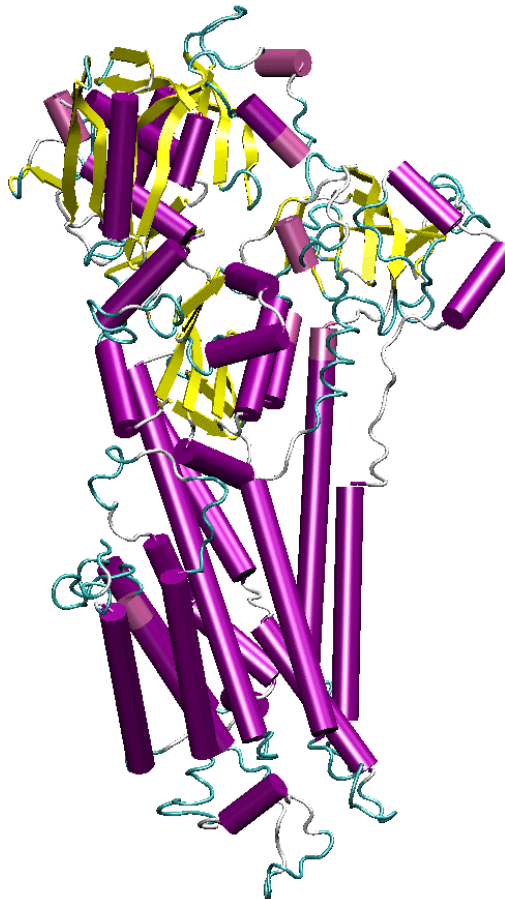
Proteins studied

EF2,
819 AA



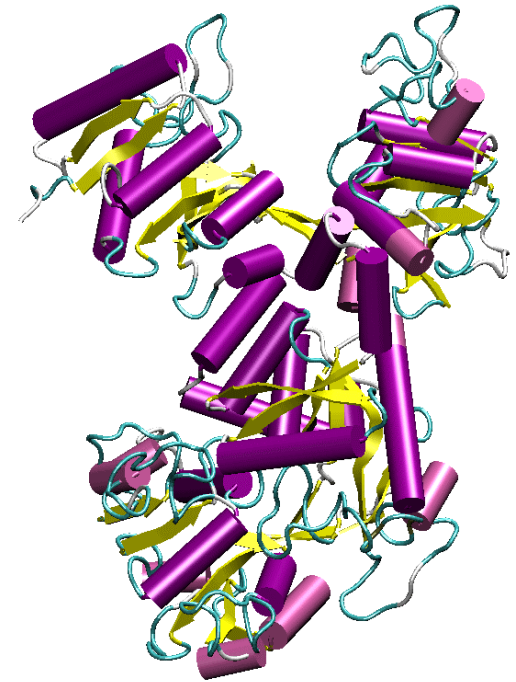
15 Å

Ca²⁺-ATPase,
994 AA



14 Å

6.5 Å



Lactoferrin,
691 AA

Elongation Factor 2

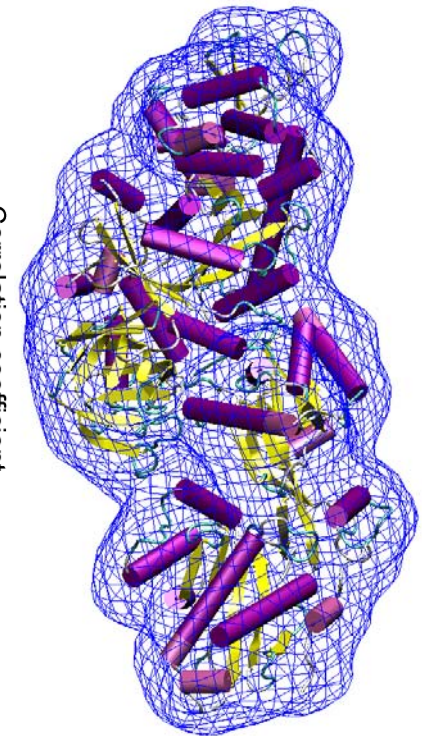
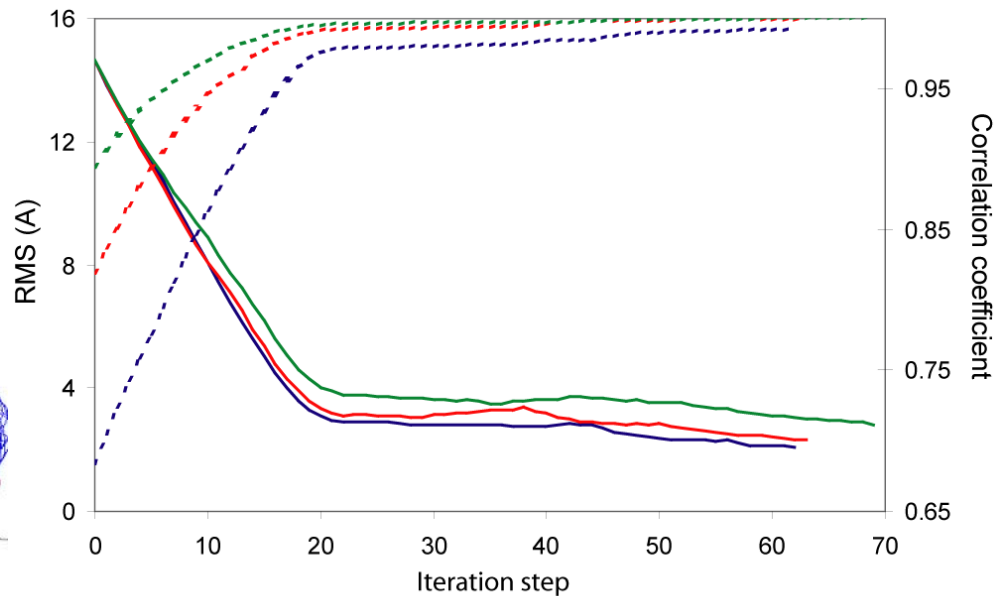
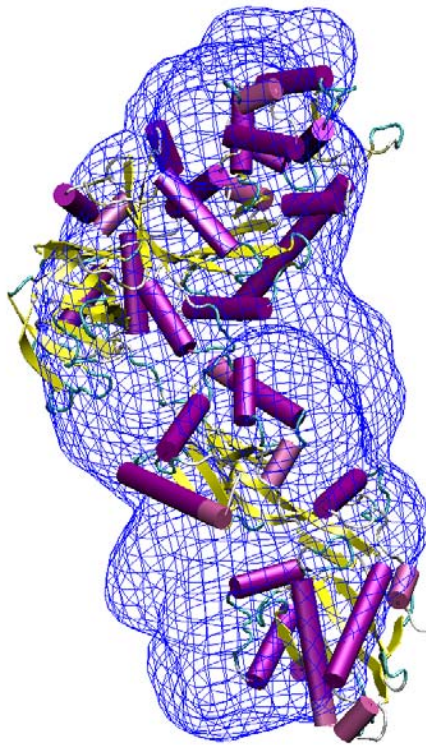
Normal modes => elastic network

All-atoms are included

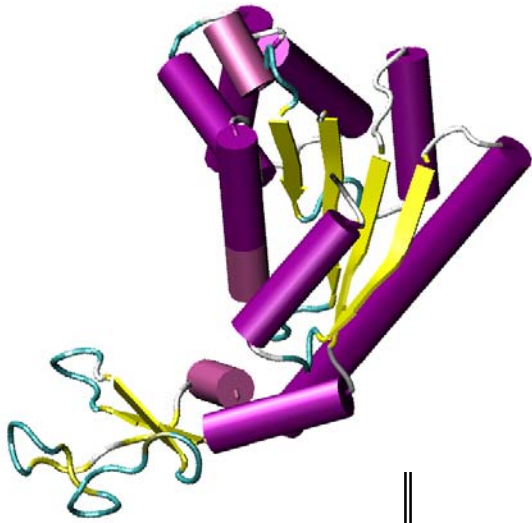
10 Å resolution

20 Å resolution

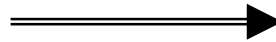
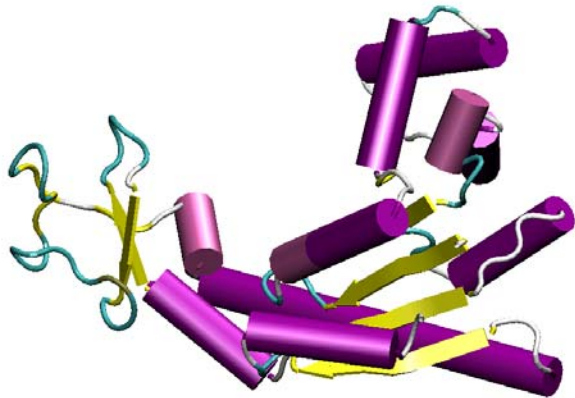
30 Å resolution



Starting from different orientation

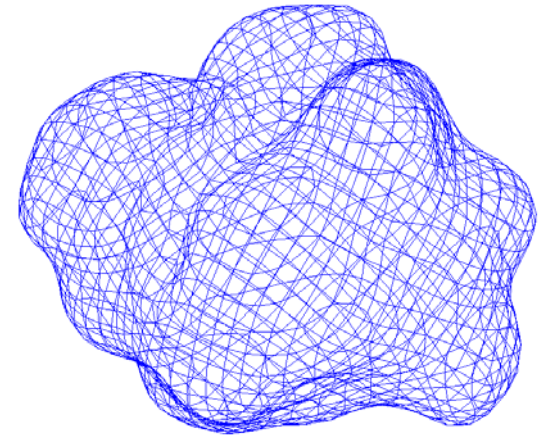


Rigid body
fitting with
Situs



Deformation of the
structure along NM

EM Map and X-ray
structure do not have
the same orientation



Rigid body fitting

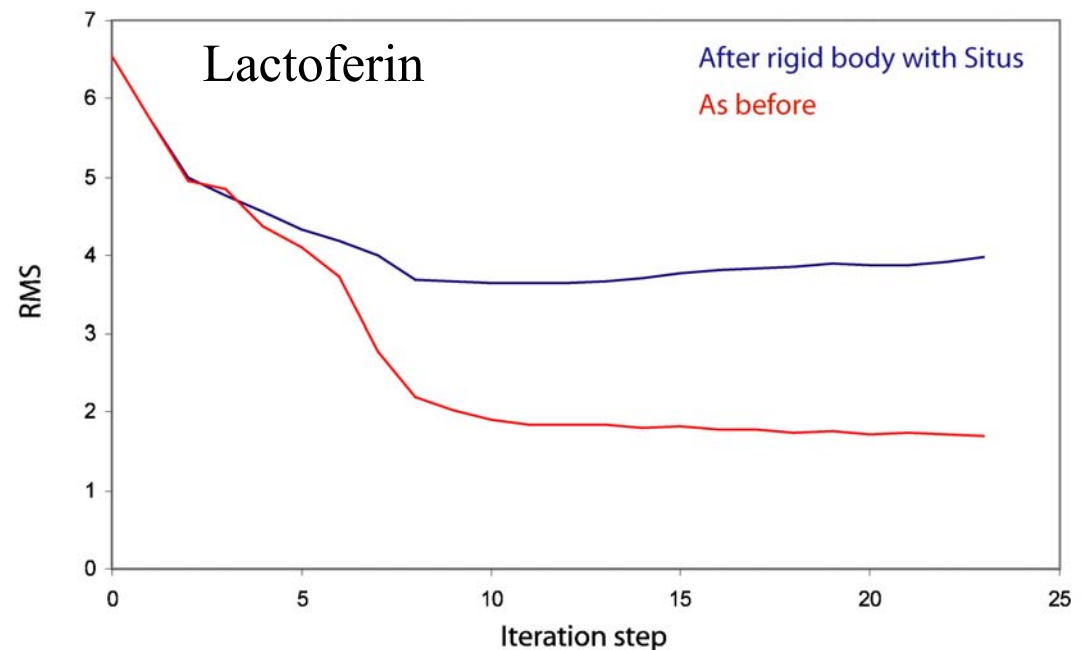
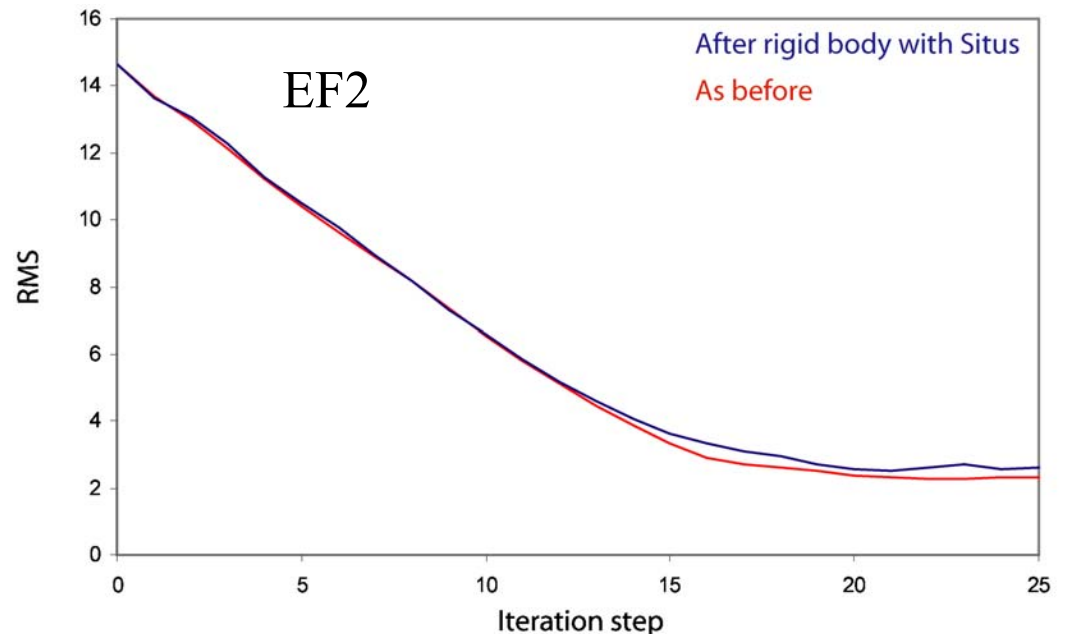


Structures (by rigid body fitting) are not perfectly oriented as previously



Problems

We need to consider rotation/translation degree of freedoms



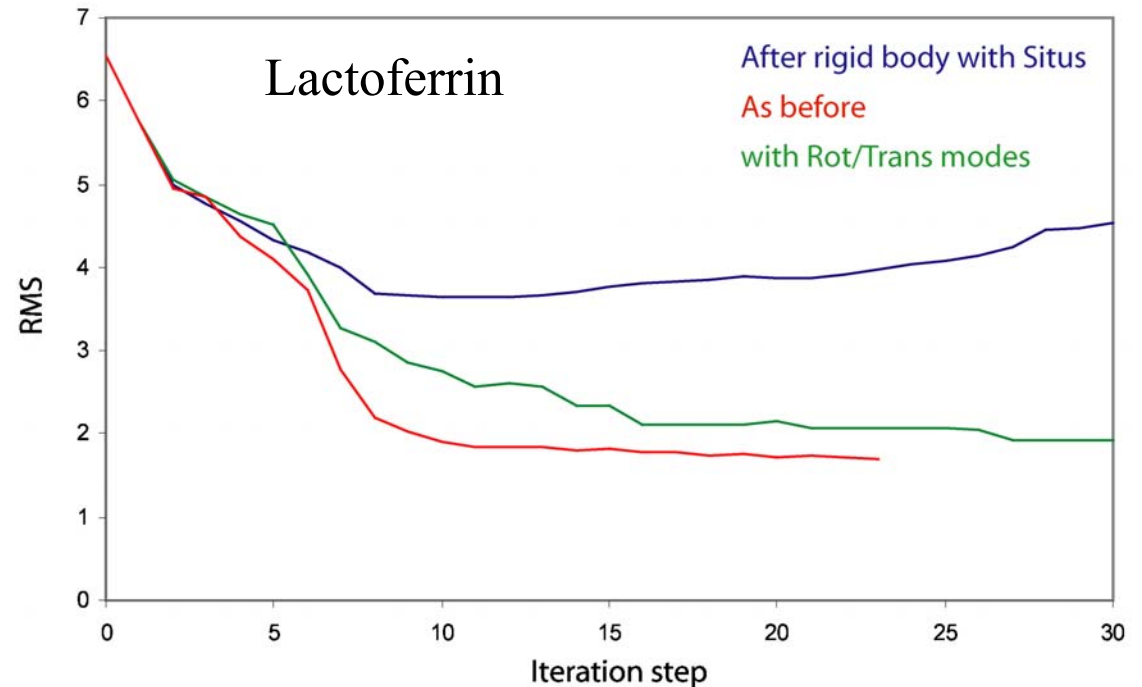
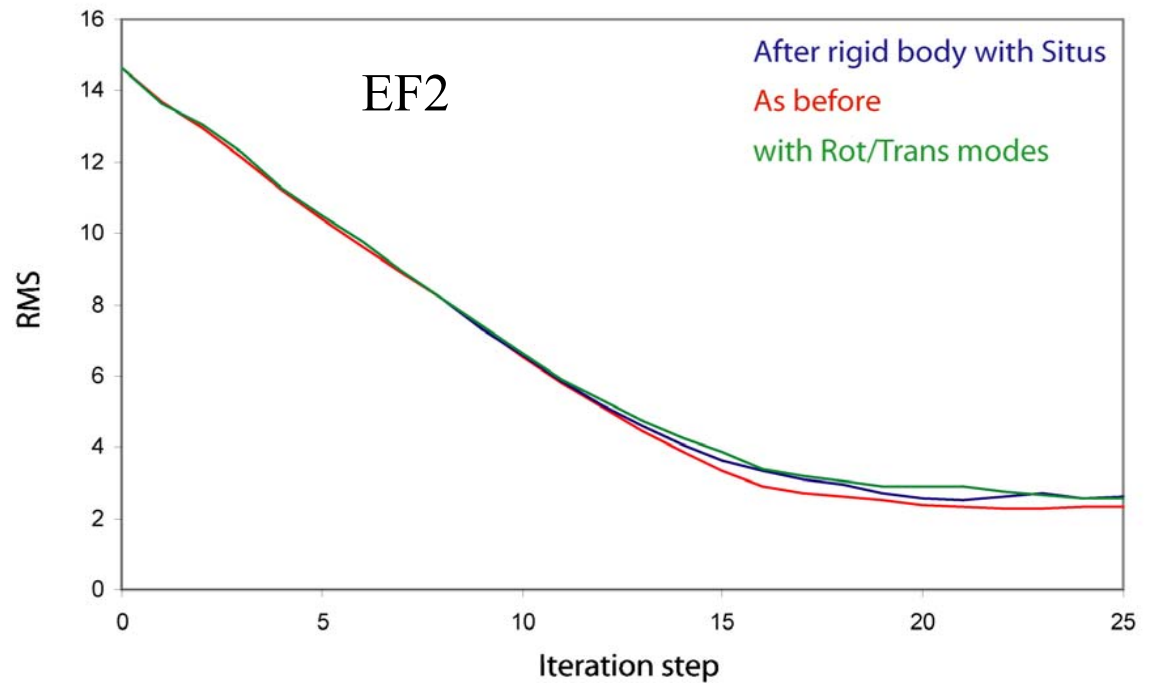
Normal mode provides 3 rotations /3 translations modes



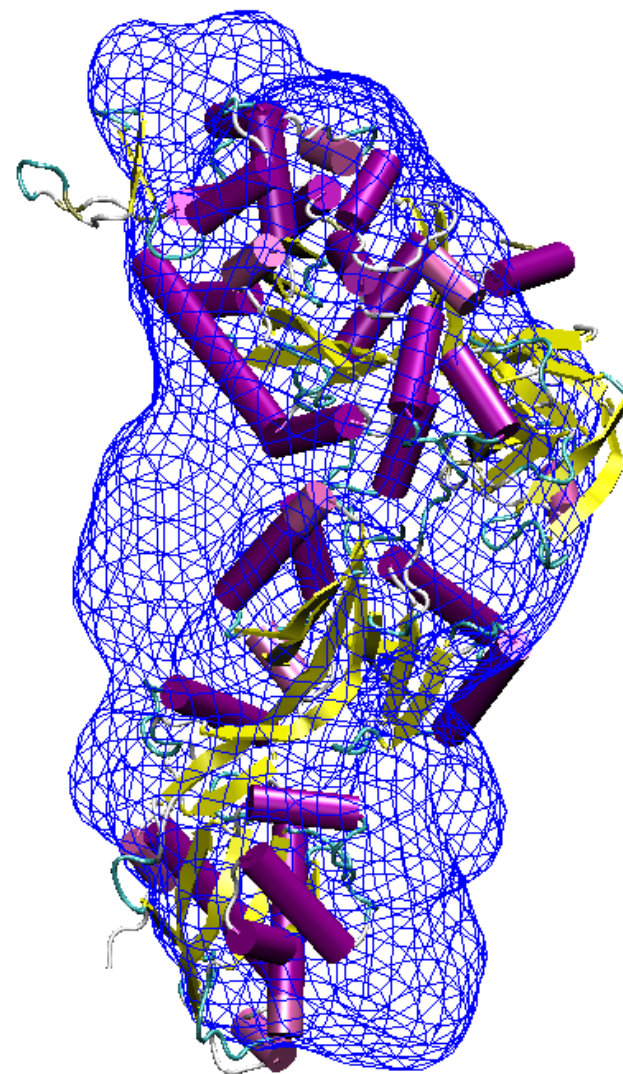
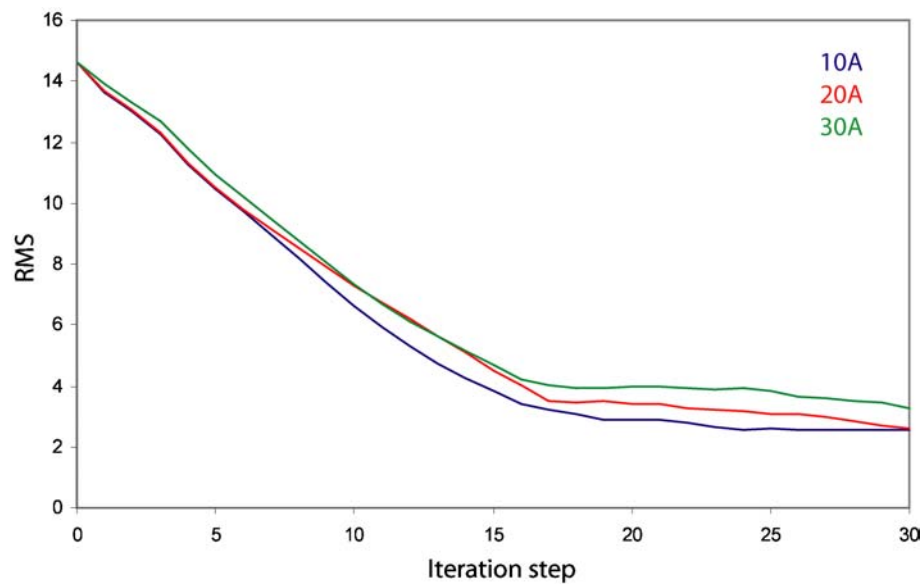
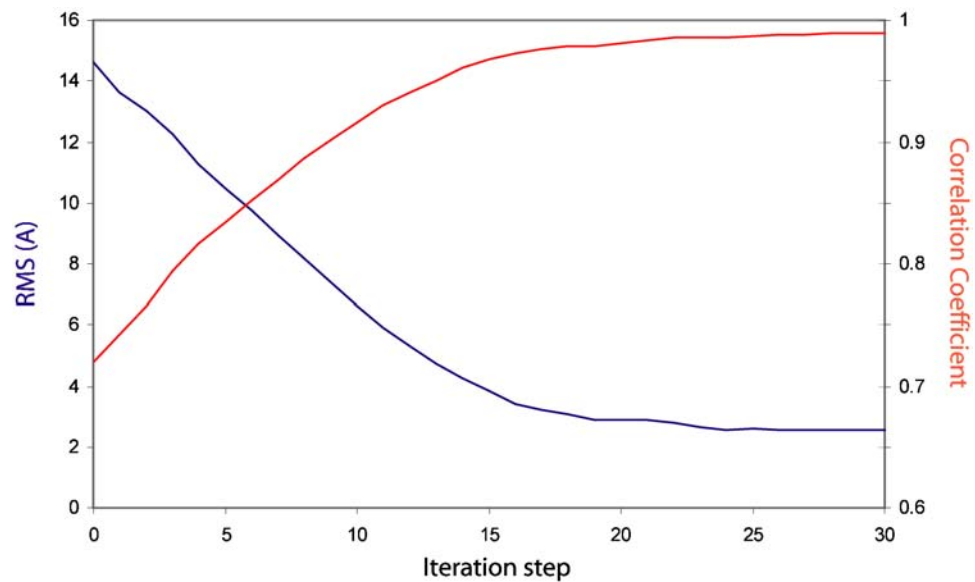
Rot/Trans modes can be included in the refinement



Fitting is improved:
automatic adjustment of rotation/translation



Elongation Factor 2



Elastic Network enables multi-scale flexible fitting

- Structures obtained are in good agreement with know structures (even at low resolution)
- Number of iteration step depend on the step size

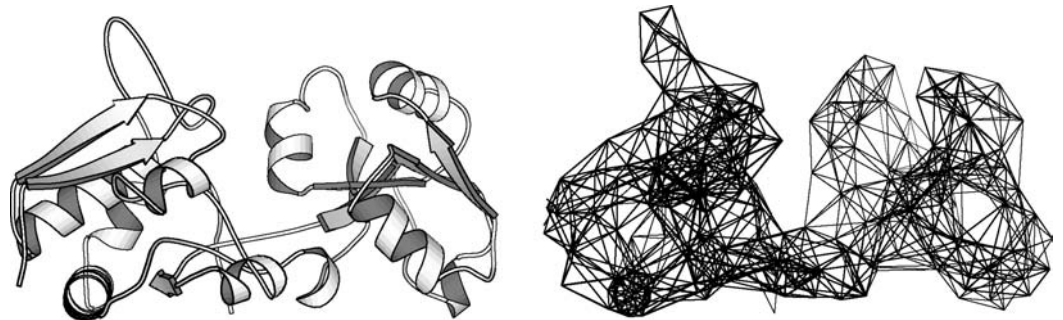
If step size is too large => may lead to some distortion
small => take longer

$$E(r_a, r_b) = \frac{C}{2} \left(|r_{a,b}| - |r_{a,b}^0| \right)^2$$

Coarse grained model

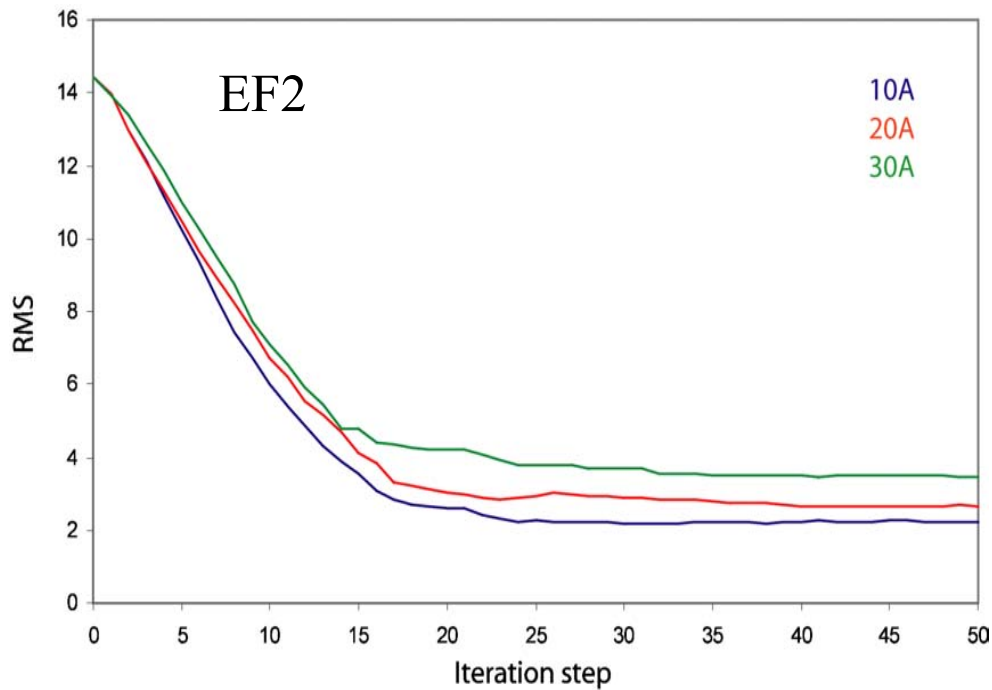


Reproduce well large conformational change of biological systems



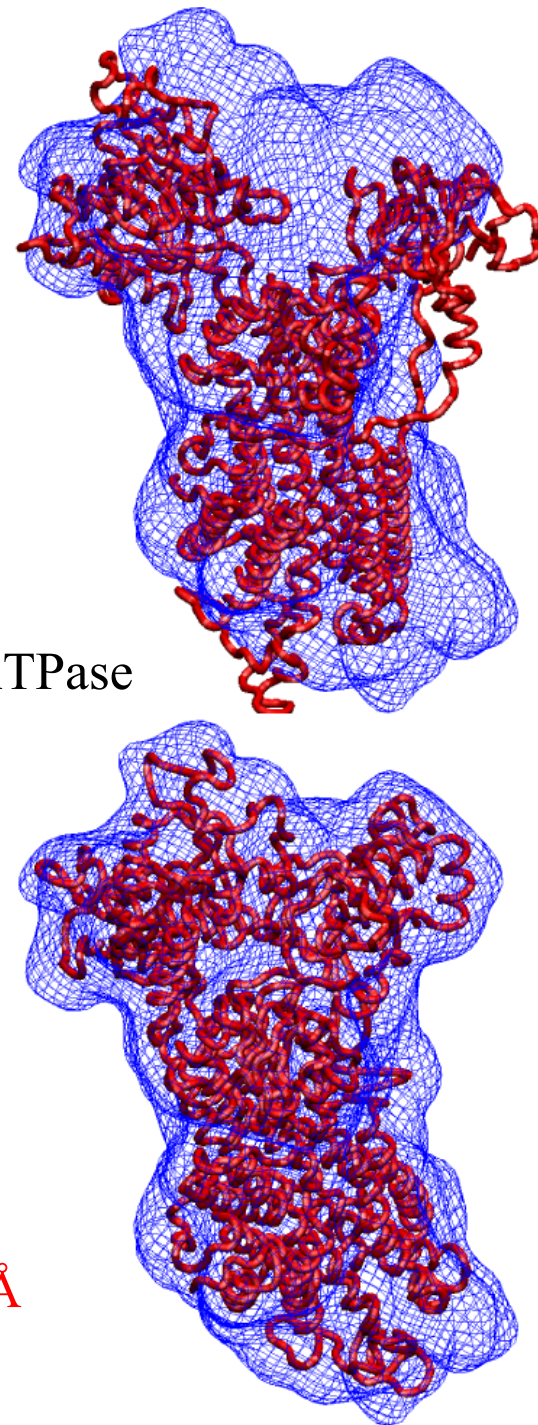
Use Cα atoms for fitting

Flexible fitting using C α only



Ca²⁺-ATPase

Final RMS = 4.8 Å



NMFF results/synthetic data

Initial structure	Atoms included	Resolution (Å)	Final RMSD (Å)		
			Lactoferrin	EF2	Ca ²⁺ -ATPase
RMSD fitted structure		10	0.8 (1.4)	1.8 (2.1)	4.3 (4.9)
	All atoms	20	1.1 (1.5)	2.1 (2.3)	5.0 (5.6)
		30	1.4 (1.9)	2.6 (3.1)	5.0 (5.5)
		10	1.0	1.8	5.1
	C α atoms	20	1.3	2.2	4.7
		30	1.8	2.8	5.4
Situs rigid body*		10	0.9 (1.4)	2.1 (2.3)	4.5 (5.0)
	All atoms	20	1.0 (1.5)	2.2 (2.4)	4.9 (5.5)
		30	1.4 (1.8)	2.9 (3.0)	5.2 (5.7)
		10	1.2	1.9	4.8
	C α atoms	20	1.4	2.2	4.7
		30	2.0	2.6	5.2
Original RMSD (Å)			6.5	14.6	14.4

Tests on experimental EM maps

Applications to experimental cryo-EM maps

- 1- Elongation Factor G bound to the ribosome (Valle *et al.* Cell **144**, 123 (2003))
- 2- E-coli RNA polymerase (Darst *et al.* PNAS **99**, 4296 (2002))
- 3 -Cowpea Chlorotic MottleVirus (swollen form) (Speir *et al.* Structure **3** 63 (1995))

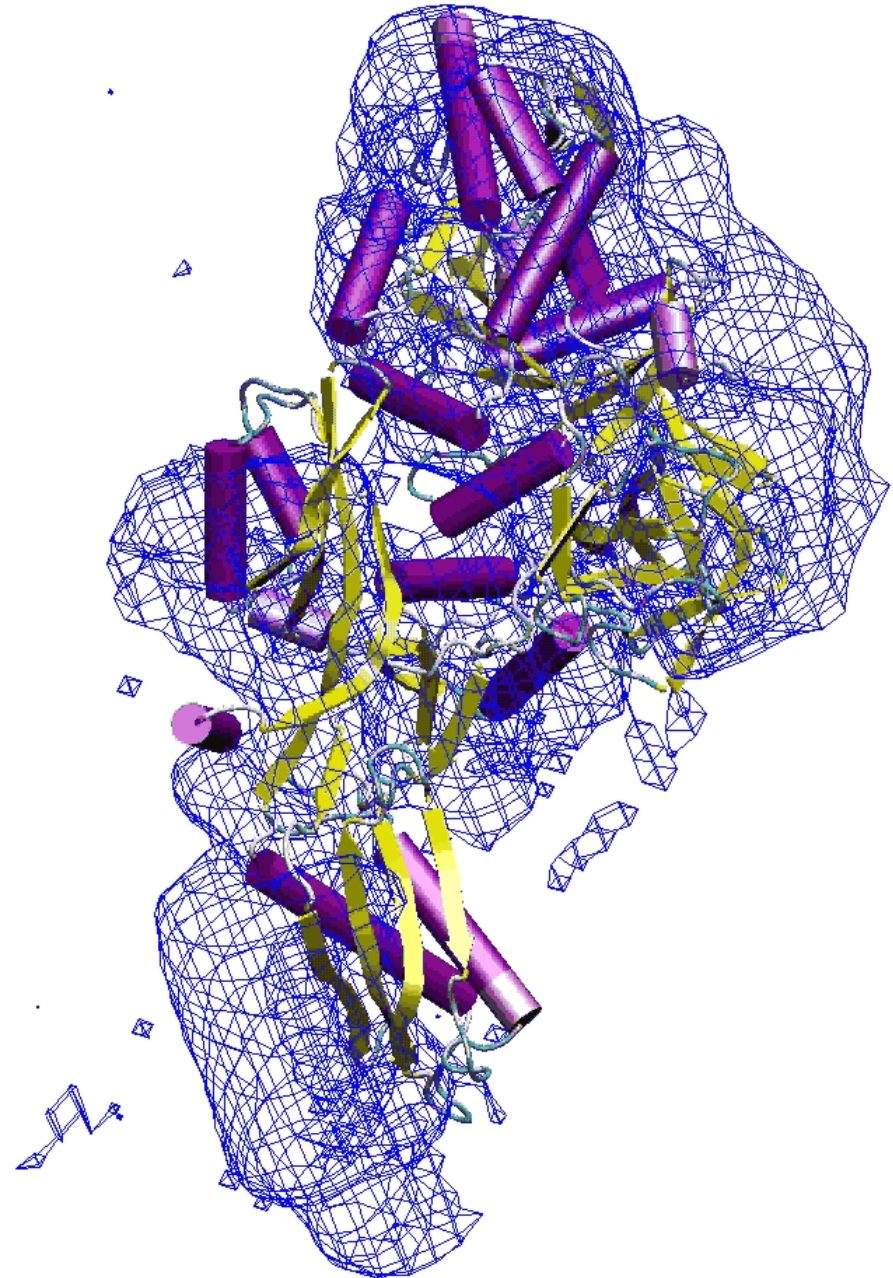
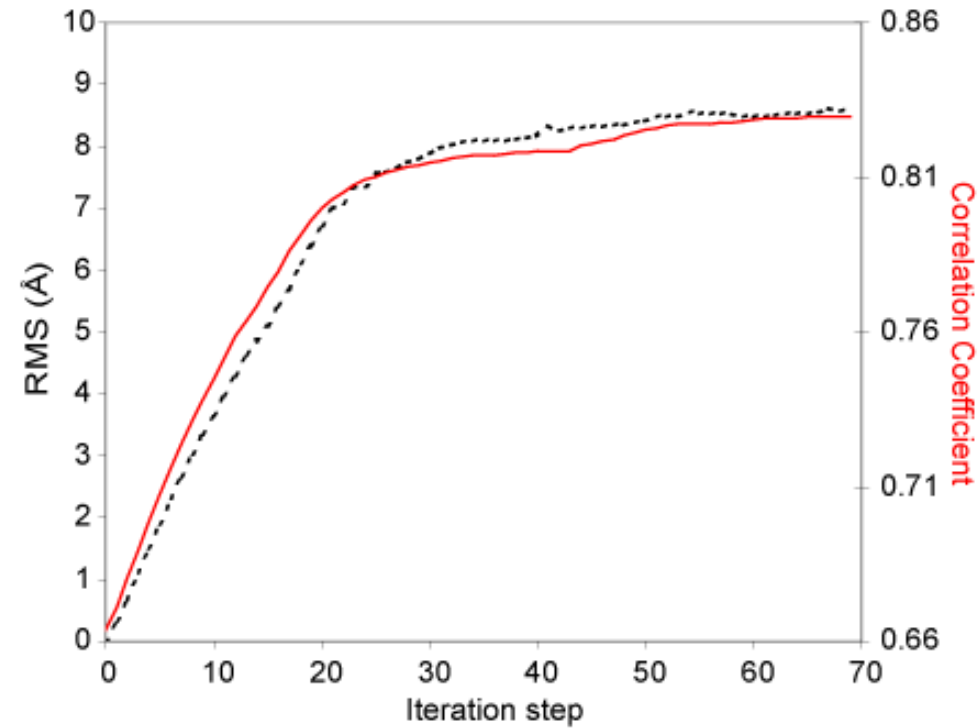
1 - Preliminary rigid body fitting

2- Flexible fitting

20 lowest frequency normal modes

Elongation Factor G bound to the ribosome

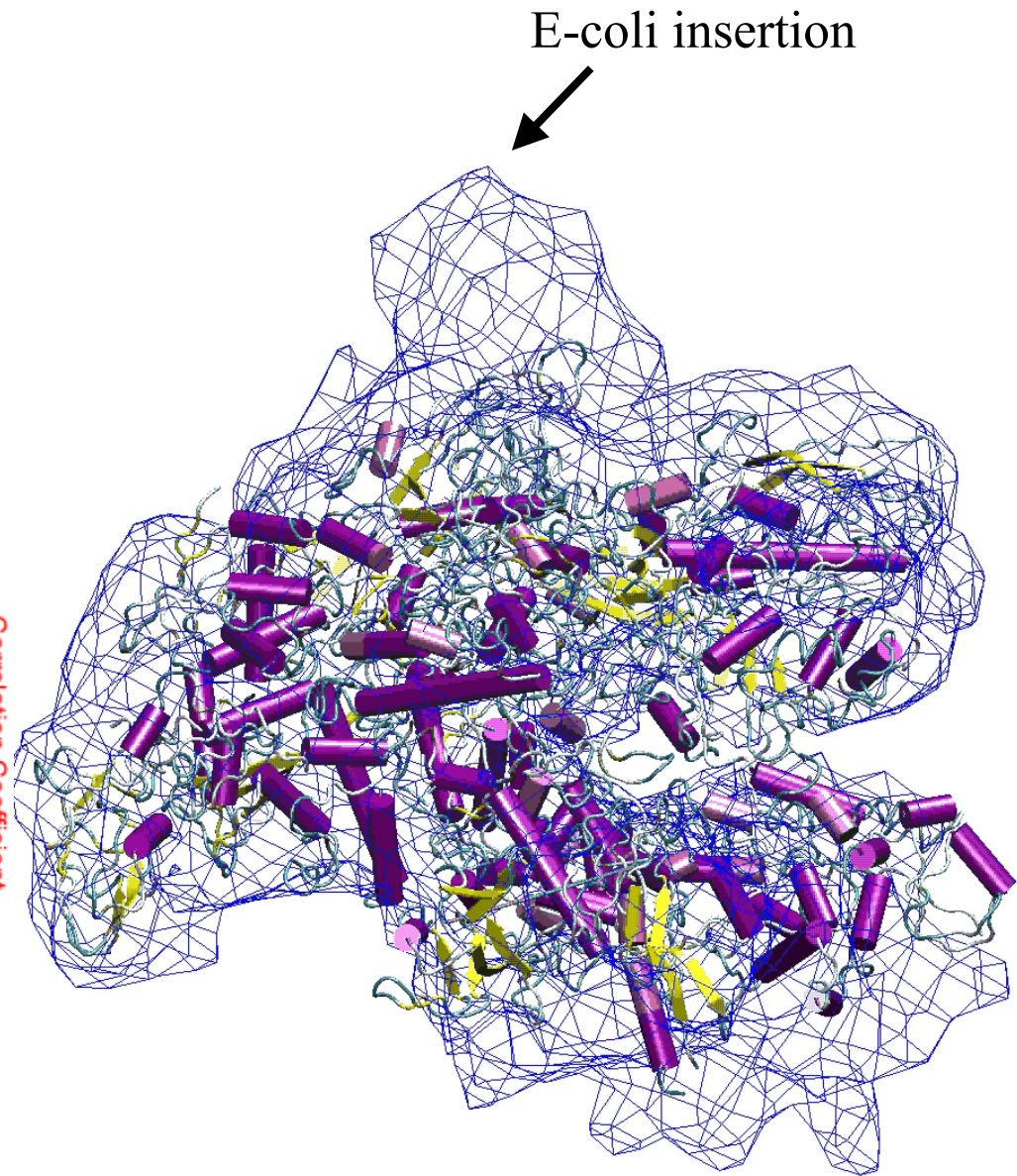
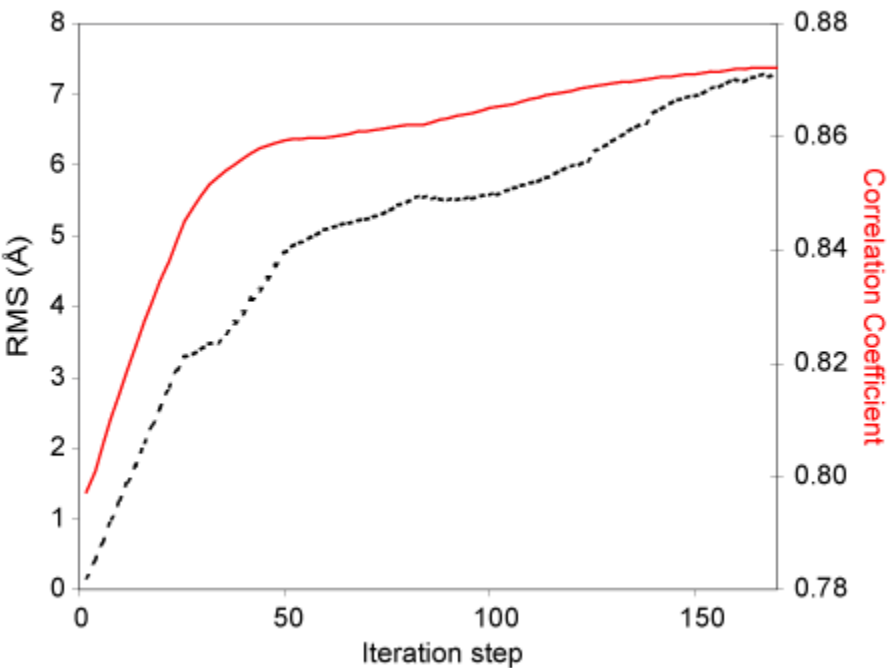
Resolution = 10.8 Å



E-coli RNA polymerase

Crystal structure of
Thermus Aquaticus

Resolution = 15 Å



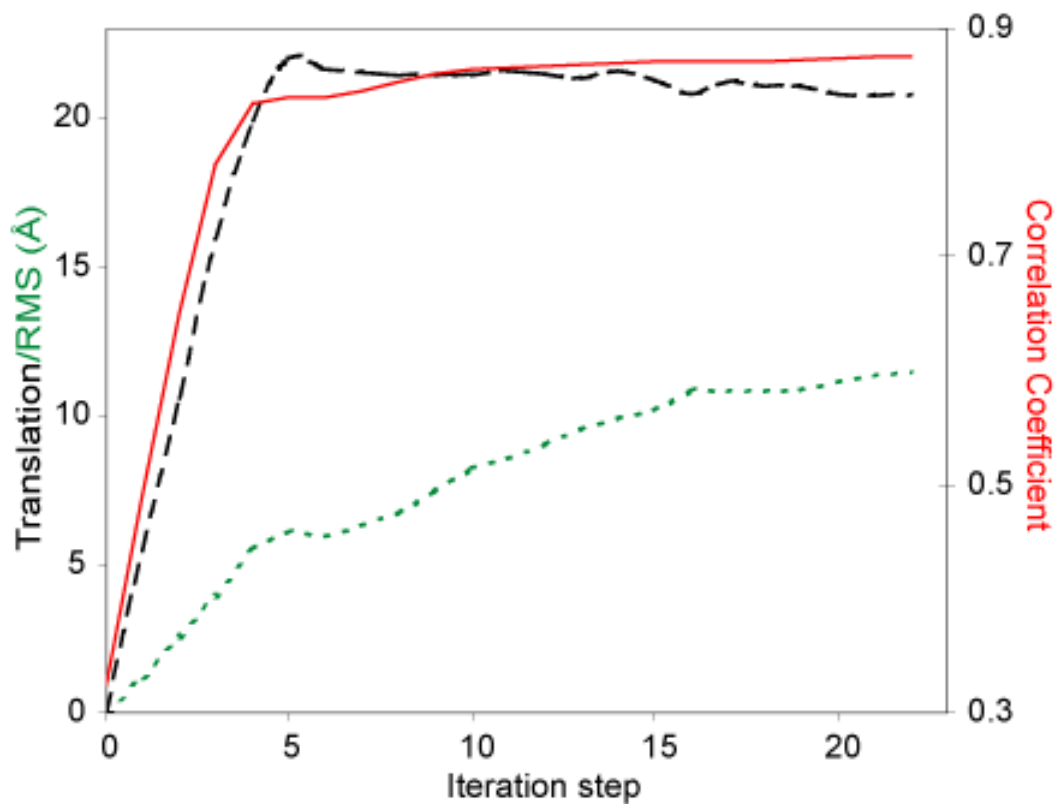
Clamp motion

Cowpea Chlorotic Mottle Virus

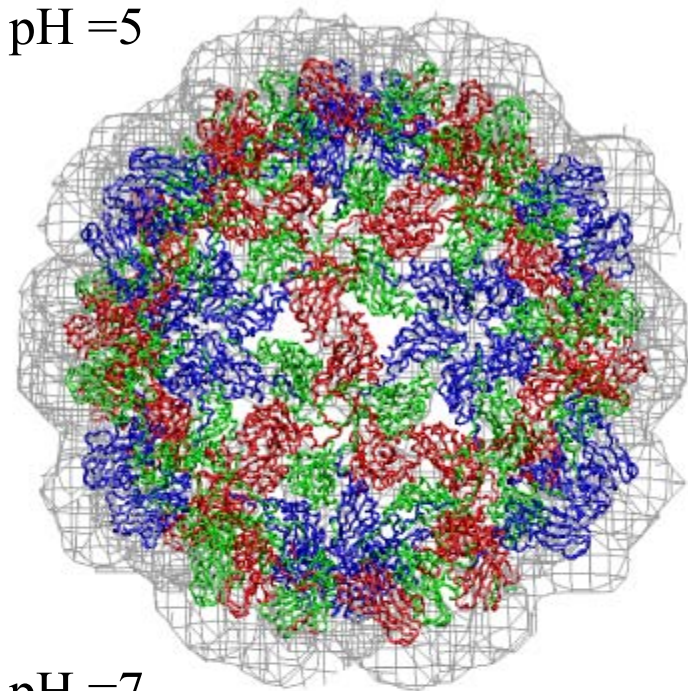
Use C α atoms only

Resolution = 28 Å

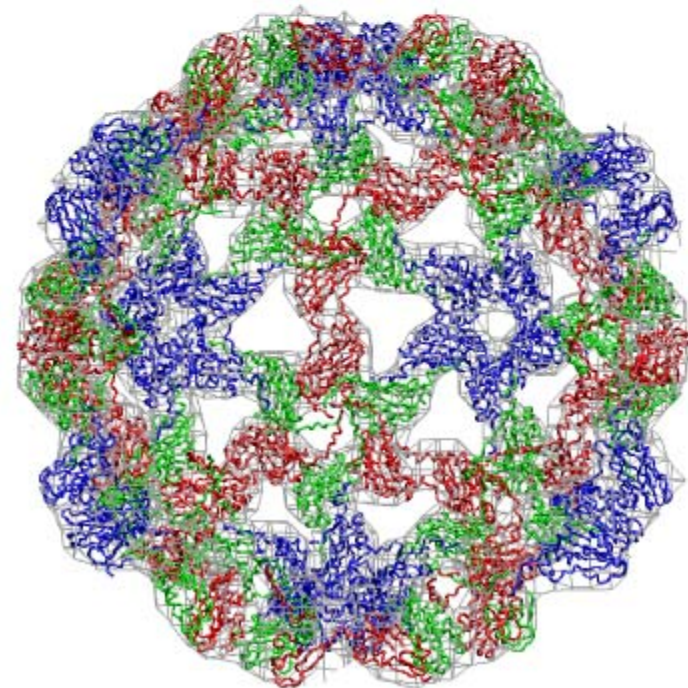
Icosahedral symmetry imposed



pH = 5



pH = 7



Summary

System	Resolution (Å)	Correlation Coefficient	
		Before	After
EF-G	10.8	0.62	0.81
RNA polymerase	15	0.79	0.88
CCMV	28	0.31	0.87

All-atoms or C α atoms

NMFF uses mechanical properties of biological systems for the flexible fitting

Acknowledgements

Seth Darst (The Rockefeller University)

Joachim Frank (HHMI, Wadsworth Center)

Jack Johnson (TSRI)